

(19)



Europäisches Patentamt
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Office européen des brevets



(11)

EP 0 185 759 B2

(12)

NEW EUROPEAN PATENT SPECIFICATION

(45) Date of publication and mention
of the opposition decision:
02.06.2004 Bulletin 2004/23

(51) Int Cl.7: **A61K 31/19**

(45) Mention of the grant of the patent:
19.11.1992 Bulletin 1992/47

(86) International application number:
PCT/US1985/001202

(21) Application number: **85903545.3**

(87) International publication number:
WO 1986/000227 (16.01.1986 Gazette 1986/02)

(22) Date of filing: **24.06.1985**

(54) ELECTROLYTE SOLUTIONS AND (IN VIVO) USE THEREOF

ELEKTROLYTLÖSUNGEN UND DEREN (IN VIVO) VERWENDUNG

SOLUTIONS ELECTROLYTIQUES ET LEUR UTILISATION (IN VIVO)

(84) Designated Contracting States:
AT BE CH DE FR GB IT LI LU NL SE

(56) References cited:

EP-A- 177 614 WO-A-81/03180
WO-A-85/01657 US-A- 3 676 553
US-A- 3 970 750 US-A- 3 993 750
US-A- 3 993 751 US-A- 4 308 255

(30) Priority: **22.06.1984 US 623510**

- **Physicians' desk Reference 28th Edition.** (1974), p. 1257
- **Informationsschreiben von Salvia** (07.05.1979)
- **Dialysis & Transplantation**, Vol. 8, S. 703, 704, 709 (1979)
- **Int. J. Artif. Organs** (1981) 148, 149, 308-311 (effectively:308-309)
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- **Int. J. Artif. Organs** (1983) 217,218
- **ASAIO 1982**, Seite 270-275
- **Dokumentation Geigy, Wissenschaftliche Tabellen**, 7. Auflage (1968), Seite 526
- **Merrill, J.P., et al., J. Clin. Invest.**, 29 (1950) 412-424
- **CRC, Handbook of Chemistry and Physic**, 61. Ausgabe, 1981, Seite D 167
- **Sargent J.A., et al., Proc. Dialysis Transplant Forum** (1977), Seite109-116

(43) Date of publication of application:
02.07.1986 Bulletin 1986/27

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Description

[0001] This invention lies in the field of in vivo techniques and compositions for replenishing fluid electrolytes and nutrients while regulating metabolic processes in living mammals.

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State of the Art

[0002] The vital functions of highly developed organisms are closely dependent on the internal aqueous medium and on the maintenance in it of extreme constance of chemical and physical properties.

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[0003] It has long been recognized that all animal intracellular and extracellular body fluids contain inorganic electrolytes, and that these electrolytes are involved in, and profoundly influence, various life processes. Attempts to make artificial electrolyte fluids which may bathe tissues or be administered to the human blood stream have been known since about 1880, and, although modern analytical tools and procedures have clarified compositional details of blood electrolytes, the use of various aqueous electrolyte solutions for in vivo purposes in human medicine and related fields has been extant for approximately one hundred years.

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[0004] Those inorganic electrolytes characteristically found in normal human blood serum at respective concentration levels above about 1 millimolar per liter of concentration are shown below in Table I. Also, for comparative purposes, in Table I are shown some representative compositions of various aqueous electrolyte solutions that have been previously prepared and used for in vivo purposes. In general, the philosophy behind the formulation of aqueous electrolyte solutions for in vivo use has been that such should mimic or closely resemble the chemical composition of electrolytes in blood and plasma. An electrolyte is a substance(usually a salt, acid or base) which in solution dissociates wholly or partly into electrically charged particles known as ions (the term is also sometimes used in the art to denote the solution itself, which has a high electrical conductivity than the pure solvent, e.g. water). The positively charged ions are termed cations while the negatively charged ions are termed anions. Strong and weak electrolytes are recognized. The dissociation of electrolytes is very markedly dependent on concentration: it increases with increasing dilution of the solution. The ions can be regarded as molecules in electrolyte solutions. Because of dissociation considerations, the term "sigma" or the greek letter for sigma ("Σ") is sometimes employed herein as a prefix to designate the total presence of a specified material, such as an electrolyte, whether or not all of the material is in an ionic form complexed with a heavy metal, or regardless of charge on the material in a given solution. A pair of brackets ([]) indicates the free concentration of the substance indicated as opposed to that bound to tissue components, such as proteins.

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Table I - Prior Art. Class Ia Solutions Containing 1 or 2 Cations, no Nutrients and No $\text{NaCO}_3/\text{CO}_2$

5	Units	Normal Plasma	I. a. 1 N.E.J.M. 1985, 1285 L fluid	I. a. 2 0.9% Saline U.S.	I. a. 3 0.95% Saline Na Lactate, U.K.; Salt
10	Na	136 - 145	155	162.5	160.3
K		3.5 - 5.0			
15	Ca free [Ca ²⁺]	2.1 - 2.6 [1.06]			
Mg free [Mg ²⁺]	0.75 - 1.25 [0.53]				
20	#Eq Cations	142.7-153.2	155	162.5	160.3
Cl	100 - 106	135		162.5	108.3
HCO ₃	26 - 28				
25	ΣPi	1 - 1.45			
SO ₄	0.32 - 0.94				
L-lactate	0.6 - 1.8				52.0 (d,l)
30	pyruvate				
Lact/pyr			ca		
0 & 0Hbutyrate					
acetoacetate					
35	8 HB/ acac				
acetate					
Other					
40	$\Sigma \text{#Eq anions}$	128.7-139.4	155	162.5	160.3
Na/Cl	1.28 - 1.45	1.00		1.00	1.48
Glucose or others	3.9 - 5.6				
CO ₂	0.99 - 1.39				
45	pH	7.35 - 7.45	5.5 - 6.5	5.5 - 6.5	6.5
ΣmOsm	285 - 295	310	325	321	
50	Uses:				
I. a. 1. Most common U.S. I.V. electrolyte solution, Herdt Manual. Causes hyperchloremic acidosis with Na/Cl = 1.00. See Black DAK, Lacet i, 1952.					
I. a. 2. Used as "normal" saline in U.K. and Canada. Geigy Handbook.					
I. a. 3. Darrow et al. J. Am. Med. Ass. 143: 365, 432, 1944. Normal Na/Cl ratio but causes abnormalities.					

Table I (Cont'd) - Prior Art, Class Ib. Solutions Containing 1 or 2 Cations, HCO_3^- , and No Nutrients.

5	Units	Normal	I. b. i.
	Plasma	Isotonic	
	moles	0.E.J.M.	NaHCO_3
		283, 1295	Salt
	L fluid	1970	
10	Na	136 - 145	160.3
	K	3.5 - 5.0	
	Ca free [Ca ²⁺]	2.1 - 2.6 [1.06]	
15	Mg free [Mg ²⁺]	0.75 - 1.25 [0.53]	
	$\Sigma \text{Eq Cations}$	142.7-153.2	160.3
20	Cl	100 - 106	108.3
	HCO_3^-	26 - 28	52
	ΣPi	1 - 1.45	
	SO_4^-	0.32 - 0.94	
25	L-lactate	0.6 - 1.8	
	pyruvate		
	Lact/pyr		
30	D, D, CHbutyrate		
	acetoacetate		
	BHB/acac		
	acetate		
35	Other		
	$\Sigma \text{Eq anions}$	128.7-139.4	160.3
	Na/Cl	1.28 - 1.45	1.48
40	Glucose or others	3.9 - 5.6	
	CO_2	0.99 - 1.39	
	pH	7.35 - 7.45	8.6
45	Σosm	285 - 295	321
	Uses:		
	I. b. i. Darrow et al. J. Am. Med. Ass. 143: 365, 432, 1944. Use of bicarbonate alone to correct Na/Cl ratio gives a solution with an abnormal pH, and one which will cause Ca^{2+} or Mg^{2+} added to the solution to precipitate as MgCO_3 or Ca CO_3 .		
50	Is the common alternative to Na lactate, salt; I. a. 3.		

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Table I (Cont'd) - Prior Art. Class Ic Solutions Containing 1 or 2 Cations, with Non-ionic Nutrients.
Typically 2.5%, 5%, 10%, 20% Glucose or Fructose in the U.S. and 2.62%, 5.25%, 10.5%, 20% Glucose or Fructose in the U.K.

5	Units	I. c. 1. Normal Plasma W.E.J.A. 283, 1295 L fluid	I. c. 2. 5% Dextrose in H ₂ O, U.S. 1970	I. c. 3. Isotonic Glucose 2 + NaCl	I. c. 4. Glucose + NaCl	I. c. 10. 0 - 5 - W + 0.9% NaCl	I. c. 11. 10% Glucose + 0.9% NaCl	I. c. 12. 2.5% Glucose + 0.45% NaCl in Electro- lyte 73	I. c. 13.
10	Na	136 - 145		54.1	53.4	154	154	77	40
K	3.5 - 5.0							55	
Ca	2.1 - 2.6 free [Ca ²⁺] 1.06 1								
Mg	0.75 - 1.15 free [Mg ²⁺] 0.53 1								
Σ Eq Cations	142.7-153.2 0	0		54.1	53.4	154	154	77	75
Cl	100 - 106			54.1	53.4	154	154	77	47.5
HCO ₃ ⁻	26 - 28								
Σ Pi	1 - 1.45							7.5H ₂ PO ₄ ⁻	
SO ₄ ²⁻	0.32 - 0.94								
L-lactate	0.6 - 1.8				17.3 (d,1)			20 (d,1)	
pyruvate									
Lact/pyr					00			00	
D, B, OHbutyrate									
acetoacetate									
BHB/acac									
acetate									
Other									
Σ Eq anions	129.7-139.4 0	0		54.1	53.4	154	154	77	75
Na/Cl	1.28 - 1.45			1.00	1.48	1.00	1.00	1.00	0.84
Glucose or others	3.9 - 5.6 278	292	195	195	278	556	139		
CO ₂	0.99 - 1.39								279 (Fructose)
pH	7.33 - 7.45 76.5	76.5	76.5	76.5	75.5 - 8.5	75.5 - 8.5	75.5 - 8.5		
Σ Oss	285 - 295 278	292	301	302	581	813	293	428	
Uses:									
45	I. c. 1.	Most used I.V. solution in the U.S. Merck Handbook, 1966, p.1867. This is combined with NaCl in varying proportions so long as the osmolarity is not below 270 mOsm.							
	I. c. 2.	Same solution in the U.K., where "isotonic" differs. Geigy Handbook, 1970, p. 334.							
	I. c. 3.	Geigy Handbook, 1970, p. 334, has Na/Cl = 1.00							
	I. c. 4.	Geigy Handbook, 1970, p. 334, has reasonable Na/Cl ratio but induces an abnormal redox state.							
	I. c. 10. through I. c. 12.	See Facts and Comparisons p. S1, Oct '81, Lippincott							
50	I. c. 13.	Facts and Comparisons p. S2b Aug '83, Lippincott. Used in parenteral nutrition.							

Table I (Cont'd) - Prior Art. Class II Solutions Containing 1 or 2 Cations, Nutrients, and $\text{HCO}_3^-/\text{CO}_2$.
None in prior art.

5	Units	Normal Plasma
	anoles	B.E.J.B.
	-----	283, 1285
	L fluid	1970
10	Na	136 - 145
	K	3.5 - 5.0
	Ca free [Ca ²⁺]	2.1 - 2.6 [1.06]
15	Mg free [Mg ²⁺]	0.75 - 1.25 [0.53]
	Σ Eq Cations	142.7-153.2
20	Cl	100 - 106
	HCO_3^-	26 - 28
	Σ Pi	1 - 1.45
	SO_4^-	0.32 - 0.94
25	L - lactate	0.6 - 1.8
	pyruvate	
	Lact/pyr	
30	D D CHbutyrate	
	acetoacetate	
	B HB/ acac	
	acetate	
35	Other	
	Σ aEq anions	128.7-139.4
	Na/Cl	1.28 - 1.45
40	Glucose or others	3.9 - 5.6
	CO_2	0.99 - 1.39
	pH	7.35 - 7.45
45	Σ osm	285 - 295
	Use	

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Table I - Prior Art, Class 2a Electrolyte Fluids Containing 3 or 4 Cations Suitable for Contacting Cells, Containing
(Cont'd) No HCO₃⁻/CO₂ and No Glucose; eg. after S.J. Ringer, *Physiol Rev* 29, 223, 1883.

	Units	Nossal Plasma H.E.D.F. 283, 1295 L. Fluid	2. a. 1. Ringer's Injection U.S.	2. a. 2. Lactated Ringer's	2. a. 3. Lactated (Commercial)	2. a. 4. Acetated Ringer's U.S.	2. a. 5. Lact/Acet Ringer's	2. a. 10. Ionomol D-CN (Abbott)	2. a. 11. Plasmalyte (Travenol)	2. a. 12. Isolyte S (McGaw) Polyionic 148(Cutter)
5	Na	136 - 145	147	129.8	130	130	140	139	140	140
10	K	3.5 - 5.0	4	5.4	4	4	10	12	10	5
15	Ca free [Ca ²⁺] [1.06]	2.1 - 2.6	2.5	0.9	1.5	1.5	2.5	2.5	2.5	2.5
20	Mg free [Mg ²⁺] [0.53]	0.75 - 1.25		1.0			1.5	1.5	1.5	1.5
25	Σ aEq Cations	142.7-153.2	156	139	137	137	158	158	158	148
30	Cl	100 - 106	156	111.8	109	109	103	108	103	78
35	HCO ₃ ⁻	26 - 28								
40	Σ Pi	1 - 1.45								
45	SO ₄ ²⁻	0.52 - 0.94								
50	L-lactate pyruvate	0.6 - 1.8		27.8 (d,l)	28 (d,l)		27.5 (d,l)	50 (d,l)	8 (d,l)	
	Lact/pyr			oo	oo		oo	oo	oo	
	D,B-OHbutyrate									
	acetoacetate									
	B-HB/acac									
	acetate					28	27.5		47	27
	Other									23 (glucconate) 148
	Σ aEq anions	128.7-139.4	156	139	137	137	158	158	158	
	Na/Cl	1.28 - 1.45	0.94	1.16	1.19	1.19	1.36	1.28	1.36	1.43
	Glucose or others	3.9 - 5.6								
	CO ₂	0.99 - 1.39								
	pH	7.35 - 7.45								
	Σ aOsm	285 - 295	309	276	272	272	312	312	312	294
	Use:		I.V. fluid	I.V. fluid	I.V. fluid	I.V. fluid	I.V. fluid	I.V. electro -lyte therapy	I.V. electro -lyte therapy	I.V. elec -trolyte therapy

- 2. a. 1. Facts and Comparisons p50, Oct'81, Lippincott
- 2. a. 2. Hartmann AF. *J. Am. Med. Ass.* 105: 1349, 1934.
- 2. a. 3. Facts and Comparisons p50, Oct'81, Lippincott.
- 2. a. 4. Facts and Comparisons p50, Oct'81, Lippincott.
- 2. a. 5. Fox et al. *J. Am. Med. Ass.* 148: 827, 1952.
- 2. a. 10. Facts and Comparisons p50, Oct'81, Lippincott.
- 2. a. 11. Facts and Comparisons p50, Oct'81, Lippincott.
- 2. a. 12. Facts and Comparisons p50, Oct'81, Lippincott.

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Table I - Prior Art. Class 2a (Cont'd).

5	Units	Normal Plasma NaOles L fluid	2. a. 13. Isolyte E N.E.J.R. 203, 1285 1970	2. a. 14. Debecco's Phosphate Saline	2. a. 15. Kreb's Ringer Phosphate
10	Na	136 - 145	140	152.2	150.76
K	3.5 - 5.0	10		4.17	5.92
Ca free [Ca ²⁺] [1.06]	2.1 - 2.6	2.5		0.9	2.54
Mg free [Mg ²⁺] [0.33]	0.75 - 1.25	1.5		0.49	1.18
Σ mEq Cations	142.7-153.2	158		159.15	164.12
Cl	100 - 106	103		140	131.51
HCO ₃	26 - 29				
Σ Pi	1 - 1.45			7.83	17.38
SO ₄	0.32 - 0.94			0.48	2.35
L - lactate	0.6 - 1.8				
pyruvate					
Lact/pyr					
D B OHbutyrate					
acetoacetate					
B HB/ acac					
acetate		49			
Other		4 citrate			
Σ mEq anions	128.7-139.4	158		159.18	165.15
Na/Cl	1.28 - 1.45	1.40		1.08	1.15
Glucose or others	3.9 - 5.6				
CO ₂	0.99 - 1.39				
pH	7.35 - 7.45			7.4	7.4
Σ mOs	285 - 295	315		308	311.16
Use:	I.V. electrolyte therapy	Usually tissue culture, sometimes cardiac surgery		Biochemical experiments	
2. a. 14. Debecco R, Vogt M. J Exp Med 1954; 99: 167 - 182					
2. a. 13. Facts and Comparisons Oct, 1981, p.50, Lippincott, St.Louis					
2. a. 15. Krebs HA. Hoppe S Z Physiol Chem 1933; 217: 193					

Table I - Prior Art. Class 2b Solutions Containing 3 or 4 Cations, $\text{HCO}_3^-/\text{CO}_2$, and No Glucose or Other Non-Ionic Nutrients.

5	Units	Normal	Z. b. i.
	Plasma	Krebs	
	H.E.J.A.	Henseleit	
	-----	283, 1285	
	L fluid	1970	
10	Na	136 - 145	143
	K	3.5 - 5.0	5.9
	Ca	2.1 - 2.6	2.5
	free (Ca^{2+})	[1.06]	
15	Mg	0.75 - 1.25	1.2
	free (Mg^{2+})	[0.53]	
	Σ mEq Cations 142.7-153.2 156.3		
20	Cl	100 - 106	127.8
	HCO_3^-	26 - 28	25
	Σ Pi	1 - 1.45	1.18
	SO_4^-	0.32 - 0.94	1.18
25	L-lactate	0.6 - 1.8	
	pyruvate		
	Lact/pyr		
	0.8 CHButyrate		
30	acetoacetate		
	3 HB/acac		
	acetate		
35	Other		
	Σ mEq anions 128.7-139.4 157.3		
	Na/Cl	1.28 - 1.45	1.12
40	Glucose	3.9 - 5.6	
	or others		
	CO_2	0.99 - 1.39	1.24
	pH	7.35 - 7.45	7.4
	Σ osm	285 - 295	308
45	Use:	Multiple Biochemical Uses	
50	Z. b. i. Krebs H.A., Henseleit K., Hoppe-Seyler's Z. Physiol Chem 1932; 210: 33-66. This is the second major advance in fluids after S.J. Ringer, Physiol 1883; 4: 29, 223. This fluid became the basis for most tissue culture "balanced salt mixtures," was used in dialysis. It is known to contain twice too much Ca and Mg. It also has an abnormal Na/Cl ratio which Krebs himself unsuccessfully attempted to correct in 1950. (See Krebs H.A. & A 1950; 4: 249- 269, or Table I class 2d.)		

Table I - Prior Art. Class 2c Solutions Containing 3 or 4 Cations, No $\text{HCO}_3^-/\text{CO}_2$ to Which is Added Non-Ionic Nutrients.

Units	Normal Plasma N.E.J.M. 283, CBS L fluid 1970	2. c. 1. Lactated Ringer's + 5%Glucose + 2.5%Glucose	2. c. 2. 1/2 Strength Acetated Lact-Ringer Ringer's + 5%Glucose + Glucose	2. c. 3. Ionosol B (Abbott)	2. c. 4. Dianeal + 5%Glucose	2. c. 5. (Travenol)	2. c. 6. Peritoneal Dialysis + 4.25%Glucose	2. c. 7. (Ae. McGraw) Dialysis + 4.25%Glucose (Travenol)
Na	136 - 145	150	65	130	57	141	141.5	132
K	3.5 - 5.0	4	2	4	25			4
Ca free [Ca ²⁺] [1.06]	2.1 - 2.6	1.5	0.75	1.5		1.75	2.0	1.875
Mg free [Mg ²⁺] [0.53]	0.75 - 1.25				2.5	0.75	0.75	0.75
Σ mEq Cations 142.7-153.2	137		66.5	137	87	146	147	141
Cl	100 - 106	109	55	109	49	101	102.5	106
HCO_3^-	26 - 28							
Σ Pi	1 - 1.45				6.5 H_2PO_4^-			
SO_4^-	0.32 - 0.94							
L - lactate	0.6 - 1.8	29(d,l)	14(d,l)		25(d,l)	45(d,l)		35(d,l)
pyruvate								
Lact/pyr	00	00		00	00		00	
0.8 Dihbutyrate								
acetoacetate								
BHB/ acac								
acetate			28			44.5		
Other								
Σ mEq anions 128.7-139.4	137	69	137	87	146	147	141	
Na/Cl	1.28 - 1.45	1.19	1.18	1.19	1.16	1.40	1.38	1.25
Glucose or others	3.9 - 5.6	278	139	278	278	83	236	236
CO_2	0.99 - 1.39							
pH	7.35 - 7.45					75.5-6.5	75.5-6.5	75.5-6.5
Σ osm	285 - 295	524	263	523	443	366	510	494
Use:	I.V. fluid nutrition & for de- hydration	I.V. fluid same as 2.c.1.	Parenteral Nutrition	Dialysis	Peritoneal Dialysis	Peritoneal Dialysis	Peritoneal Dialysis	Peritoneal Dialysis

2. c. 1. Multiple Manufacturer's Facts and Comparisons p.52, Oct '81

2. c. 2. Multiple Manufacturer's Facts and Comparisons p.52, Oct '81

2. c. 3. Multiple Manufacturer's Facts and Comparisons p.52, Oct '81

2. c. 4. (Abbott) Facts and Comparisons p.52b, Aug '83

2. c. 5. (Travenol) Facts and Comparisons p.704, Oct '82

2. c. 6. (American McGraw) Facts and Comparisons p.704, Oct '82

2. c. 7. (Travenol) Facts and Comparisons p.704, Oct '82

Table I - Prior Art. Class 2d Solutions Containing 3 or 4 Cations, Plus Non-Ionic Nutrients and HCO_3/CO_2

5	Units	Normal Plasma N.E.J.A. 283, 1285 L fluid	Z. d. 1. Krebs Serum Substitute 1970	Z. d. 2. Tyrade's Solution I (Schiaassek)	Z. d. 3. Tyrade's Solution	Z. d. 4. Locke's Solution
10	Na	136 - 145	141	151.54	150.1	157.57
K	3.5 - 5.0	5.93		5.9	5.9	3.57
Ca free [Ca ²⁺] [1.06]	2.1 - 2.6	2.54		1.8	1.8	2.16
Mg free [Mg ²⁺] [0.53]	0.75 - 1.25	1.18		0.45	0.45	0
$\Sigma \text{mEq Cations}$	142.7-153.2	154.37		162.07	160.5	165.46
Cl	100 - 106	104.8		147.48	147.48	163.92
HCO_3	26 - 28	24.9		11.9	11.9	3.57
ΣPi	1 - 1.45	1.23		1.22	1.22	--
SO_4	0.32 - 0.94	2.36				
L - lactate	0.6 - 1.8			1.33		
pyruvate		4.9		0.09		
Lact/pyr				14.8		
D B OHbutyrate						
30	acetoacetate					
BHB/acac						
acetate						
35	Other		2.45 glutamate 5.4 fumarate			
$\Sigma \text{mEq anions}$	128.7-139.4	154.47		162.81	161.6	167.49
Na/Cl	1.28 - 1.45	1.35		1.03	1.02	0.98
40	Glucose or others	3.9 - 5.6	9.2	5.45	5.6	5.6 - 13.7
CO_2	0.99 - 1.39	1.0		1.17		
pH	7.35 - 7.45	7.4		7.1	7.1	?
Σosm	285 - 295	308.2		328	318.3	336
45	User		Artificial Serum for Tissue Slices Normal Na/Cl			
50						
		2. d. 1. Krebs HA. <i>8.B.A.</i> 4: 249 - 269, 1950. Not used <i>in vivo</i> but presented for comparison of composition.				
		2. d. 2. Tyrade's solution as modified for liver perfusion by Schiaassek H. <i>Biochim Z</i> 336: 460, 1963. Not used <i>in vivo</i> but presented to show prior art in composition. Same for 2.d.3. Tyrade's, and 2.d.4, Locke's.				
		2. d. 3. Tyrade HV. <i>Arch Int Pharmacodyn Ther</i> 20: 205 - 223, 1910.				
		2. d. 4. Locke FS. <i>Zentral Physiol</i> 14: 670 - 672, 1900.				

55 [0005] Contemporarily, a large number of different aqueous electrolyte solutions are prepared, sold in commerce, and used as in vivo fluids, such as for electrolyte and fluid replacement, parenteral nutrition, and dialysis (both hemoperitoneal).

[0006] Even a cursory examination of Table I will confirm the medical dicta that "plasma is an unmakable solution".

The solutions listed in Table I illustrate this belief. The essential problem is that plasma contains, in addition to major inorganic electrolytes, trace quantities of various electrolytes plus various metabolites including plasma proteins. In practice, it has not been possible to construct synthetically a replication of plasma because of its complexity. Blood, extracellular fluid, and even plasma can be regarded as tissues.

[0007] In most prior art electrolyte solutions, the concentration of chloride anions (Cl^-) is higher than in human plasma or serum. For example, the Krebs Henseleit solution (see Table I) contains a concentration of Cl^- which is about 20% higher than in human serum. This anion gap, that is, the difference between the positive cations and the negative anions, is now known to be due largely to the anionic metabolites in normal plasma plus the contribution of acidic amino acid groups found on plasma proteins. Referring to Table I, it is seen that the total positive cations in plasma is 142-154 meq/l while the total anions is only about 128-137 meq/l leaving a deficit of about 14-17 meq/l of anions. For convenience, the anion gap in human plasma can be expressed as the ratio of sodium cation milliequivalents per liter to chloride anion milliequivalents per liter.

[0008] From Table I, it is clear that the Krebs Serum substitute (Krebs, H.A. Biochem, Biophys. Acta 4, 249-269, 1950) comes closest to approximating the electrolyte composition of human plasma. In this solution, Krebs attempted to correct the excessive Cl^- content in Krebs Henseleit solution (Hoppe-S. Z. Physiol. Chem. 210, 33-66, 1932) using metabolic experiments with tissue slices. Because of the law of electrical neutrality, Na^+ cannot be added to a solution without some anion (such as Cl^-) being added also; the sum of cations and anions must be equal in any solution. In his 1950 attempt, Krebs chose pyruvate⁻, L-glutamate⁻, and fumarate²⁻ as anions to be added.

[0009] An alternative to Krebs selection of anions came about at the same time. In 1949, the use of high concentrations of acetate as a metabolizable organic anion was advocated (Mudge G.H., Mannining J.A., Gilman A.; Proc. Soc. Exptl. Biol. Med. 71, 136-138, 1949). This idea led in 1964 to the advocacy of the use of 35-45mM (millimolar) acetate in commercial hemodialysis fluids (Mion C.M. Hegstrom R.M., Boen S.T., Scribner B.H.; Trans. Am. Soc. Artif. Internal Organs 10, 110-113, 1964).

[0010] In addition to the above organic anions, the current reference work "Facts and Comparisons" indicates various commercial electrolyte fluids which contain lactate anion.

[0011] All of the prior art electrolyte solutions (with or without nutrients) as exemplified in Table I are now believed to lead to undesirable and pathological consequences particularly through extended usage. As regards acetate, editorials recently appearing in the British Medical Journal, 287, 308-309, 1983) present evidence that acetate leads to fatigue, nausea, malaise, sudden hypotension, increased atherosclerosis, hypoventilation, and hypoxia. Also, the originator of acetate dialysis now advocates its use only in "healthy" patients (Pagel M.D., Ahmed S. Vizzo J.E. and Scribner B.H.; Kidney Int. 21, 513-518, 1982).

[0012] Krebs choice of glutamate⁻ and fumarate²⁻ is incorrect because these anions do not penetrate cell membranes in a predictable manner, but, like citrate³⁻, exhibit severe gradients of six fold or greater between plasma H_2O and cell H_2O . The alternate use of d,L-lactate⁻ (Hartmann AF. J Am Med Asso 103 1349-1354, 1934) is now known to induce severe abnormalities, particularly coma at levels far below the 28 to 35 mM d,L-lactate contained in these solutions (Oh MS et al, N. Eng J Med 301 249 251, 1979; Stolberg L, et al N Eng J Med 306: 1344-1348. 1982; Ballabriga A, et al Helv Paediatr Acta 25:25-34, 1970) in to the induction severe abnormalities in redox and phosphorylation state induced by the use of L-lactate alone. The use of gluconate⁻ induces abnormalities in the hexosemonophosphate pathway. Indeed, all previous used organic ions violate the "safe entry points" or the normal Na:Cl ratio as herein defined.

[0013] In addition to the use of d,L-lactate, gluconate, fumarate, glutamate, pyruvate, and citrate anions in current commercially available prior art electrolyte fluids, and wherein such anions are typically employed at levels above those found in the (plasma or serum) of healthy humans, many such prior art commercial fluids also employ high levels of nonionic metabolites, such as fructose and glycerol, which induce separate redox state and phosphorylation potential abnormalities in phosphorylation potential with rapid destruction of liver purine nucleotides and their release into blood sometimes leading to renal shutdown due to uric acid deposition in the kidneys (see Woods H.F., Eggleston L.V. and Krebs H.A.: Biochem. J. 119, 501-510, 1970). Fructose in plasma above 0.2mM must be considered to violate the "safe entry point". Likewise, use of intravenous glycerol at levels above 5mmol/l as currently practiced leads, in tissue containing glycerol kinase, such as kidney and liver, to accumulation of 10 mM glycerol phosphate (over 100 times normal). See Bruch H.B. et al.: J. Biol. Chem. 257, 3676-3679, 1982).

[0014] In addition to failing to solve the anion gap problem (or to provide a normal milliequivalent ratio of sodium cation to chloride anions) without causing profound and adverse physiological effects (including disruption of normal redox state and normal phosphorylation potential), many prior art aqueous electrolyte solutions for in vivo usage fail to have a pH which approximates the pH of mammalian intracellular and extracellular fluids, especially plasma or serum.

[0015] Mammalian systems normally operate at temperatures between about 37-38°C where, by common thermodynamic convention, neutral pH is taken to be about 7 at 25°C. It is clear that changes in pH, (the negative log 10 of $[\text{H}^+]$ concentration) necessarily affect the fundamental energetic relationships occurring in living cells. Also, enzymes have sharply defined ranges of $[\text{H}^+]$ concentration in which they perform their catalytic functions in a normal manner. Deviation of mammalian plasma pH down to 6.9 or above 7.7 from its normal range of 7.35 - 7.45 is therefore fatal to

most mammalian organisms. Massive changes in the cellular redox and phosphorylation states also disorder cellular homeostasis.

[0016] The pH of human plasma is normally maintained by the human body in the range from about 7.35 to 7.45 while the pH of human cellular cytoplasm is about 7.2 (see Veech et al in *J. Biol. Chem.* 254, 6538-6547, 1979). If blood pH drops to 6.8 in man, then death ensues from cardiac arrest, and if blood pH increases to above pH 7.7, then death ensues from convulsions.

[0017] The major chemical system maintaining body pH within this narrow normal range is the $[\text{CO}_2]/[\text{HCO}_3^-]$ buffer system. The $[\text{CO}_2]$ of blood is maintained minute to minute by a portion of the mammalian brain called the respiratory center which senses brain cell pH and adjusts the depth and speed of respiration to change pH by increasing or decreasing $[\text{CO}_2]$ according to the famous Henderson Hasselbalch equation (Henderson L.J., *Silliman Lectures*, Yale U. Press, New Haven, 1928).

[0018] Even though pH is thus seen to be a critical factor in mammalian blood, many commercial electrolyte solutions as administered have pH values which deviate substantially from normal. Others give excessive Cl^- relative to Na^+ which results in hyperchlloremic acidosis, (Black D.A.K.: *Lancet* i 305-12, 1953), or give organic anions in a manner which causes measurable deviations from normal in the metabolic processes of the cell. Also, many commercially available electrolyte solutions contain no carbon dioxide which can result in a loss of respiratory drive and consequent hypoxia in patients.

[0019] The compositions and methods disclosed herein overcome the above indicated prior art problems. These compositions and methods employ definite ratios of $[\text{bicarbonate}^-]/[\text{carbon dioxide}]$, $[\text{L-lactate}^-]/[\text{pyruvate}^-]$, and $(\text{d-betahydroxybutyrate}^-)/[\text{acetoacetate}^-]$. Each of these mixtures constitute a near equilibrium couple which is known to be a normal constituent of mammalian plasma. While each of these pairs of components has been previously employed at least on a laboratory basis in solutions used for animal (mammalian) experiments, these mixture pairs have never previously been used in an electrolyte solution to obtain a normal Na:Cl milliequivalent ratio or to solve the anion gap problem.

[0020] All previous electrolyte solutions, and plasma substitutes, induce severe and measurable pathogenic abnormalities and no prior art electrolyte solution or plasma substitute has both (a) employed at least one of the three mixture pairs of this invention and (b) achieve a normal Na: milliequivalent ratio as taught herein. Thus, for example, the Krebs Henseleit solution contains the $[\text{HCO}_3^-]/[\text{CO}_2]$ buffer system (but contains excessive chloride ions). Schimassek (Schimassek H.; *Bio. Chem. Z.* 336, 460, 1963) added about normal blood levels of lactate and pyruvate to what is essentially Tyrode's solution (see Tyrode, M.J.: *Arch. Int. Pharmacodyn* 20, 205, 1910) containing 2.5% albumin in an attempt to create a physiological solution for perfusion. It should be noted that Schimassek added 1.33mM/L D-Z-lactate, which is definitely abnormal (see normal blood lactate levels shown in Table 1). Further, the Na^+ of 151 mmol/l and Cl^- of 147.5 mmol/l in Schimassek's modified Tyrode's solution approximates the concentration of 155mmol/l Na and 155mmol/l Cl in so-called normal (0.9%) saline, the most widely used electrolyte infusion solution, and thus obtained a grossly abnormal Na:Cl milliequivalent ratio of about 1.24 - 1.45 with a mean of about 1.38. Infusions of electrolyte solutions with a Na:Cl milliequivalent ratio of less than about 1.38 have long been known to cause hyperchlloremic acidosis in the treated organism. (See Levinsky N.G. in Harrison's *Textbook of Medicine* pp 230-236, McGraw-Hill, N. Y., 1983). It is the attempt to avoid this problem that leads to the wide use of such solutions as Ringer's lactate or acetate dialysis fluids which overcome the Na:Cl ratio problem, but which in turn create gross abnormalities of other types. It is the attainment of a normal Na:Cl milliequivalent ratio in a manner which avoids the pathological consequences inherent in all currently known or practiced methods which is a major part of the invention herein disclosed.

[0021] The making of a Krebs Henseleit electrolyte solution (or other prior art electrolyte solution) and the incorporation thereinto of a mixture of L-lactate and pyruvate anions, or of a mixture of D-betahydroxybutyrate and acetoacetate anions did not, and could not, result in the making of an electrolyte solution wherein the anion gap problem was overcome (or wherein the milliequivalent ratio of sodium cations to chloride anions was normalized), in accordance with the teachings of the present invention, because each of such resulting solutions would still contain excessive chloride anions and so would inevitably cause hyperchlloremia if and when used in human or mammalian therapy.

[0022] In general summary, the prior art describes a series of electrolyte solutions typically of about 270-320 milliosmoles (or higher) comprised of: (a) 1 to 4 metallic cations of sodium, potassium, magnesium, and calcium in amounts greater than 0.5 mmol/l, (b) 1 to 5 inorganic anions of chloride plus also HPO_4^{2-} , (c) 0 to several organic carboxylic or bicarbonate anions, (d) 0 to 5 nonionic materials in concentrations of greater than about 0.5mM/L from the group comprising CO_2 gas, glucose, urea, glutamine, and others, and (e) sometimes one or more high molecular weight substances, such as albumin, hemocel, and the like. None of these solutions, for the reasons herein above explained, either normalize the milliequivalent ratio of Na:Cl at all, or normalize this ratio without causing profound and adverse physiological consequences. In the present invention, there are provided compositions of a complex fluid nature for in vivo usage which can substantially completely eliminate all of such prior art problems. While the components of these new solution compositions are known solution components, no one has heretofore formulated the solutions of the present invention which not only tend to achieve a normal plasma milliequivalent ratio of sodium cations to chloride

anions, but also tend to achieve a normalization of plasma pH and a normalization of the cellular redox state and the cellular phosphorylation potential. Also, these new solutions permit one to avoid usage of the previously employed carboxylic anionics, as acetate, or lactate alone, which cause adverse effects.

[0023] US-A-3993751 (Zinke), and other prior publications of which US-A-3993751 is typical, discloses a physiologically compatible aqueous salt solution for mammalian administration which contains bicarbonate anions but not nutrient and has CO₂ only in the atmosphere above the solution.

[0024] US-A-3676553 (Reynolds) discloses an injectable aqueous solution which contains bicarbonate anions but no dissolved CO₂.

[0025] Other prior publications include US-A-4308255, US-A-3970750 and US-A-3993750, Physician's Desk Reference, 28th edition (1974) page 1257, EP-A-0177614 and WO-A-85/01657. The last two documents were published after the priority date of the present application.

BRIEF SUMMARY OF THE INVENTION

[0026] The present invention provides salt solutions as defined in the claims of this specification, to which reference should now be made.

[0027] This invention thus provides electrolytes for use in accomplishing electrolyte and water therapy while simultaneously normalizing blood composition in a mammal (including man) by introducing in a physiologically effective amount by any means, including parenterally, intravenously, intraarterially, intramuscularly, intravascularly, and the like, by dialysis, or orally, and the like into such mammal an aqueous solution wherein:

- (a) the ratio of sodium cation milliequivalents per liter are so selected as to tend to produce the range found in normal mammalian blood plasma,
- (b) there is a physiologically effective amount of the near equilibrium couple d-betahydroxybutyrate⁻ and acetoacetate⁻, and
- (c) the pH ranges from 5 to 9.

[0028] There are also disclosed electrolytes of the class indicated wherein physiologically normal concentrations of the diivalent cations Mg²⁺ and Ca²⁺ may be included without precipitation. No one has previously made solutions for *in vivo* use which contain the correct Na⁺:Cl⁻ ratio and which also contain the physiologically normal respective amounts of Mg²⁺ and Ca²⁺.

[0029] When used for mammalian administration, such a solution:

- (a) tends to maintain and normalize in plasma the milliequivalent ratio of sodium cations to chloride anions in the normal range, and
- (b) tends to maintain and normalize plasma pH, and
- (c) tends to maintain and normalize the redox state and the phosphorylation potential.

[0030] One (first) class of such solutions characteristically utilizes (contains) an inorganic class of anions comprised of chloride and bicarbonate. These solutions have a physiological pH which is broadly in the range from 5 to 9, and preferably in the range from 6.9 to 8.6, and more preferably in the range from 7.35 to 7.45, and most preferably is 7.4 (for human use). Dissolved carbon dioxide is also present in these solutions. When administered, these solutions not only tend to maintain the treated mammal's normal blood (and plasma) ratio of sodium to chloride, but also tend to set (regulate) the treated mammal's blood (plasma) pH at a normalized value. In addition the treated mammal's redox state and phosphorylation potential tend to be normalized.

[0031] Another (second) class (preferred) of such solutions characteristically utilizes (contains) chloride anions and a class of carboxylate anionic mixture couples comprised of at least one member from the group consisting of (a) a mixture of l-lactate⁻ anions and pyruvate⁻ anions, (b) a mixture of d-betahydroxybutyrate⁻ anions and acetoacetate⁻ anions, and (c) a mixture of both (a) and (b). These solutions have a physiological pH which is as above defined in connection with such (first) class of solutions. When administered, these solutions not only tend to maintain the treated mammal's redox state within a normal range, but also tend to maintain that mammal's phosphorylation potential within a normal range.

[0032] Another (third) class (more preferred) of such solutions characteristically utilizes (contains) both chloride anions, and bicarbonate/carbon dioxide mixture, as in such (first) class of solutions, but also utilizes (contains) such class of carboxylate anionic couples, as in such (second) class of solutions. When administered, these solutions achieve the above indicated effects obtained from the use of such (first) class of solutions and the above indicated effects obtained from the use of such (second) class of solutions.

[0033] The specified milliequivalent ratio of sodium to chloride in normal mammalian blood generally is believed to be in the range from 1.24:1 to 1.47:1. In the case of a normal human adult, this range is now believed to extend (based on published information) from 1.24:1 to 1.45:1 and preferably from 1.33:1 to 1.42:1 and most preferably from 1.36:1 to 1.42:1. These ratios of $\text{Na}^+:\text{Cl}^-$ are typically employed in solutions used in the practices of this invention. Ratios above 1.47, i.e. from 1.47 to 1.6 can be used within the scope of this invention as when it is the physician's conscience intention to create an abnormal $\text{Na}^+:\text{Cl}^-$ ratio as, for example, to create an excess of alkali reserve; however, such higher ratios are generally not presently preferred for general usage. In the case of dialysis fluids or to create an alkalotic condition in a cell or to correct an existent acidosis, this $\text{Na}^+:\text{Cl}^-$ ratio could range from a normal value (1.24 to 1.45) to 1.6.

[0034] In using these couples, the important factor is the ratio of the concentration of [product] / [reactant] (see Eqns O, 1,2,3,4.5 & 7 hereinbelow). The absolute concentration becomes important in affecting the chemical activity of water (e.g. the osmotic pressure).

[0035] The total quantity, or sum (sigma), of the couple (d-betahydroxybutyrate / acetoacetate) present in a solution of this invention can range from 0.1 to 465 mMoles/liter of solution. However, in routine situations, the quantity of the couple commonly ranges from 0.1 to 25 to 60 mmoles/liter.

[0036] The ratio of bicarbonate milliequivalents per liter to dissolved carbon dioxide milliequivalents per liter in a solution of this invention can range from 0.1:1 to 55:0.1 and preferably 11:1 to 24:1. More preferably, such total ranges from 10 to 45 mmol/l and such ratio ranges from 18:1 to 26:1, and still more preferably such total ranges from 23 to 35 mmol/l while such ratio ranges from 19:1 to 21:1. A ratio of 19.95 for $[\text{HCO}_3^-]/[\text{CO}_2]$ gives a pH 7.4, which is presently particularly preferred.

[0037] The ratio of l-lactate anion milliequivalents per liter to pyruvate anion milliequivalents per liter in a solution can range from 20:1 to 1:1. Preferably, such total quantity ranges from 0.5 to 10 mmol/l and such ratio ranges from 3:1 to 15:1, and more preferably such total quantity ranges from 2 to 8 mmol/l while such ratio ranges from 5:1 to 12:1.

[0038] The ratio of d-betahydroxybutyrate anion milliequivalents per liter to acetoacetate milliequivalents per liter in a solution of this invention ranges from 6:1 to 0.5:1. Preferably, such total ranges from 1 to 10mmol/l and such ratio ranges from 4:1 to 1:1, and more preferably such total ranges from 2 to 5mmol/l while such ratio ranges from 3:1 to 1.5:1.

[0039] By the term "milliequivalent ratio" as sometimes used herein, reference is had to the ratio of milliequivalents per liter of one substance to milliequivalents per liter of another substance in an aqueous medium.

[0040] One of the three near equilibrium couples discloses herein (the bicarbonate/carbon dioxide couple) tends to regulate the concentration of hydrogen ions in blood (plasma) and in the treated mammal's cells, and each one of such couples tends to normalize the redox state of each of the three pyridine nucleotide couples. The phosphorylation potential also tends to be normalized. Also, each such near equilibrium couple when used as herein described constitutes a safe entry point into the metabolic system of a mammal.

[0041] By the term "safe entry point" as used herein reference is generally had to a metabolite which, in living tissue or cells:

- (1) does not cause a massive buildup of one or more of intermediate cellular metabolites,
- (2) does not cause a severe disruption of any one of the controlling nucleotide ratios in a living cell,
- (3) can be added to a physiological system of a living mammal at a concentration level which is greater than that which is found normally in such system (such as blood plasma of a fasting mammal) without causing any appreciable distortion in metabolism and without causing any pathological conditions to arise, and
- (4) may be found in normal variants of the physiological state as when the total of d-betahydroxybutyrate plus acetoacetate reaches a level of about 8 to 10 mmol/l in three- day fasting man, or the total of l-lactate plus pyruvate rises to a level of about 5 to 6 mmol/l in a jogging normal man.

[0042] Further, each such above described near equilibrium couple exhibits a distribution or permeability between intracellular fluid and extracellular fluid such that the ratio of the concentrations in, respectively, intracellular fluid to extracellular fluid ranges from 1.0:1 to 1.5:1 in most all mammalian cells.

[0043] These respective three pairs of permeant monocarboxylate near equilibrium couples are unique among metabolites in being osmotically neutral in respect to the water in intracellular and extracellular space. Administration of these three couples, as their appropriate cationic salts (individually or in some combination with one another as taught herein) necessarily results in no net change in the distribution of water between intracellular and extracellular spaces in most tissues. By administration of varying ratios of these couples, however, the physician may control the distribution of water by varying the redox state and hence the phosphorylation state as described in equation 7 herein below.

[0044] Osmotically active substances incorporated with the solutions of this invention preferably should each constitute a safe entry point. For example, glucose above 13mmol/l is higher than ever occurs under normal physiological conditions in a healthy man. Use of glucose above 13mmol/l (as in the widely used 5% glucose solution) as a calorie source is, apart from consideration of the source of pathology, and apart from the carboxylate couples, considered herein to be

an acceptable source of calories. The extreme ability of the mammalian body to regulate its glucose metabolism makes it far to be preferred over other possibly nonionics, such as fructose or glycerol, which enter the metabolic system in an uncontrolled manner causing pathologic changes such as are already referenced, and so such are not safe entry points.

5 [0044] Characteristically, a solution used in the practice of this invention contains from 1 to 2400 millimoles per liter of sodium cations, but, in routine situations, commonly ranges from 120 to 170 mmol/l; and more preferably from 129 to 163.5mmol/l and most preferably from 136 to 145mmol/l.

[0045] In addition, a solution contains sufficient chloride anions to produce a milliequivalent ratio of sodium cations to chloride anions in the range above defined.

10 [0046] Optionally, in addition to sodium, a solution of this invention can contain one or more of the following additional metallic cations each in a respective quantity as below indicated:

Table II

cation component	Quantity range (millimoles per liter)		
	broad	preferred	more preferred
potassium	0 - 90	0 - 40	0 - 5
calcium	0 - 60	0-10	0 - 1.5
magnesium	0-15	0-10	0-1

15 [0047] Optionally a solution of this invention can have additionally incorporated (dissolved) therein 0 to about 2400 millimoles per liter of at least one osmotically active substance which is preferably metabolizable and preferably substantially nonionic (including zwiterionic). A solution used in the practice of this invention is further characterized by generally having:

20 (1) sufficient total substances dissolved therein to produce an osmolarity ranging from 260 to 5000 milliosmoles/liter (mOs), and preferably from 265 to 550 mOs, and more preferably from 280 to 320 in mOs, and most preferably 311 milliosmoles/liter.

25 (2) the relationship between total (dissolved) ionic substances is such that the pH ranges from 5 to 9, and preferably from 6.9 to 8.6; and most preferably from 7.35 to 7.55;

(3) the charges of all cations equal the charges of all anions; and

30 (4) the minimum total concentration of the near equilibrium couples present is at least about 0.1 millimoles per liter, and preferably is at least 0.5 mmol/l, and more preferably 2 mmol/l, while the maximum concentration thereof is not more than 465 mmol/l and more preferably is not more than 65 mmol/l and most preferably is not more than 50 mmol/l.

35 [0048] Examples of usable osmotically active substantially nonionic substances include glucose, glycerol, fructose, sorbitol, and the like. Glucose is presently most preferred.

40 [0049] As hereinbelow explained, the solutions of the present invention find use in a wide variety of therapeutic applications, such as in electrolyte and fluid replacement, parenteral nutrition, and dialysis.

45 [0050] Various additional objects, aims, purposes, features, advantages, applications, variations and the like will be apparent to those skilled in the art from the teachings of the present specification taken with the claims.

DETAILED DESCRIPTION

50 [0051] This description is based upon best available information (including theory) known to the inventor. Any misdescription or the like, if such should exist, is not believed to alter the fundamentally correct basis and evidence supporting the present invention.

A. The Redox State

55 [0052] In biological cells, most reactions are catalyzed by enzymes of which an average cell may have of the order of 10^4 . In one classification, enzymes may be grouped in only six functional categories.

(1) dehydrogenases which transfer H⁺ and e⁻ from one substrate to another by the use of cofactors, such as NAD⁺ (nicotinamide adenine dinucleotide), or prosthetic groups, such as FAD (flavin adenine dinucleotide), or others;

(2) kinases or phosphotransferases which effect the group transfer of a phosphate to a substrate usually by using

a co-factor, such as ATP or other similar phosphate-containing compounds;

(3) carbon-carbon bond group transferases which either break carbon-carbon bonds using co-factors of the co-enzyme A type or occur on a solid state substrate, such as a glycogen particle, or the surface of a fatty acid synthase multi-enzyme complex;

(4) isomarases which effect internal rearrangements within a compound;

(5) hydratases which either add or subtract water from a substrate; and

(6) peptidases which break C-N bonds or create such bonds again usually taking advantage of a solid state synthetic matrix, such as a ribosome.

10 A special class of substrates taking part of biological reactions catalyzed by enzymes are called co-factors or co-enzymes. Co-enzymes, such as, for example, NAD, become attached and detached from an enzyme during a catalytic cycle, while prosthetic groups, such as flavin nucleotides or cytochromes, remain firmly attached during the catalytic cycle.

15 [0053] Since co-enzymes take part in multiple intracellular reaction within a given cellular compartment, the chemical potential of the co-enzyme couple becomes of central importance in energy transformation and oxide-reductions occurring in living matter. The thermodynamic characteristics of a particular whole set of oxide-reduction reactions is dependent upon the ratio of the free concentrations (strictly speaking, the activities) of the free $[NAD^+]$ and free $[NADH]$ ratio. The ratio $(NA(P)D^+)/[NAD(P)H]$, thus represents and defines the redox state, at a given pH, of a particular pyridine nucleotide couple, and this ratio then determines:

20 (1) the extent and direction of reversible reactions in near-equilibrium with that co-enzyme couple;
 (2) The extent to which a co-enzyme couple can be effective as an intracellular reducing agent, for example, in reducing the beta-oxpacyl co-enzyme A to beta-hydroxyacyl-coenzyme A; and
 25 (3) the magnitude of the free-energy changes of oxide-reductions in the electron transport chain responsible for the major portion of ATP synthesis.

[0054] The term "redox state" as thus used herein can be considered to refer to the oxidation-reduction state of any one or more of the three main pyridine nucleotide couples. Each of these couples are:

30 (A) The cytoplasmic $[NAD^+]/[NADH]$ linked dehydrogenase reactions of: (1) Lactate dehydrogenase (EC 1.1.1.27);
 (2) Malate dehydrogenase (EC 1.1.1.37); and (3) Glycerol 3-phosphate Dehydrogenase (EC 1.1.1.8).
 (B) The mitochondrial $[NAD^+]/[NADH]$ linked dehydrogenase reactions of: (1) Beta hydroxybutyrate dehydrogenase (EC 1.1.1.30); and (2) Glutamate dehydrogenase (EC 1.4.1.3).
 35 (C) The cytoplasmic $[NADP^+]/[NADPH]$ linked dehydrogenase reactions of: (1) Isocitrate dehydrogenase (EC 1.1.1.42); (2) 6-Phosphogluconate dehydrogenase (EC 1.1.1.44); and (3) Malic Enzyme (EC 1.1.1.40).

40 [0055] The three pyridine nucleotide couples or pools each achieve different redox potentials because of the chemical energies of the substrates to which they are linked by their respective enzymes since the standard redox potential of $[NAD^+]/[NADH]$ is about -0.32V. Thus, the near-equilibrium NAD-linked dehydrogenases have a K_{eq} of about $10^{-11}M$, the mitochondrial NAD-linked dehydrogenases have a K_{eq} of about $10^{-9}M$, and the cytoplasmic NADP linked dehydrogenases have a K_{eq} of about 1. The differences in pyridine nucleotide redox states within the cell may be considered to result from the fundamental properties of matter. Over time, enzymes have evolved which take advantage of these fundamental properties to organize the chemical reactions of the cell into coherent purposeful sequences we know as metabolism.

45 [0056] The oxidation of lactate anions to pyruvate anions (that is, the loss of $2H^+$ and $2e^-$ from lactate) is accompanied by the reduction of pyridine nucleotide NAD^+ . That is, NAD^+ gains two electrons and one H^+ with the other H^+ being liberated into the aqueous media where its activity is indicated and controlled by the HCO_3^-/CO_2 couple.

50 [0057] In general, the term "redox state" may also be defined as a ratio of [oxidized substrate]/reduced substrate]. The half or mid point potential E_h is conventionally measured as a potential in volts relative to a standard hydrogen electrode potential in accordance with the Nernst equation. The mid point potential of the NAD^+ system, that is, where the ratio of $[NAD^+]/[NADH]$ equals 1 at a pH of 7.0 and a temperature of 25 C is -0.32 volts under standard conditions. The midpoint potential of $[O_2/H_2O]$ is +0.816 volts. The cytoplasmic pyridine nucleotide system accepts H^+ and e^- from the organic compounds provided to mammalian organisms and transfers them to the mitochondrial pyridine nucleotide system where, by the electron transfer system, the $2H^+ + 2e^-$ reduce $1/2O_2$ to form water while conserving the energy of the oxidation reduction reaction by converting ADP + Pi to ATP. The reaction generates energy and heat. The redox state of cytoplasmic $[NAD^+]/[NADH]$ couple is about -0.19 volts, that of the mitochondrial $[NAD^+]/[NADH]$ couple is about -0.28 volts while that of the cytoplasmic $[NADP^+]/[NADPH]$ couple is about -0.42 volts. The last or $NADP^+$ couple is a much stronger reducing agent than the others and is used for reductive synthesis in the body, such as the making

of fatty acids from carbohydrates; (see Krebs and Veech, 1969) in The Energy Levels and Metabolic Control in Mitochondria (Papa S., Tager J.R., Quagliariello E. & Slater E.C. eds) pp 329-382, Adriatica Editrice, Bari.

[0058] In the case of a living cell, a plurality of oxidation-reduction reactions occur simultaneously. Under normal conditions, these reactions occur in a normal healthy cell in a predictable manner. How these various redox states are regulated has just been described in thermodynamic terms. The normal healthy cell keeps the redox state of its free cytoplasmic $[NAD^+]/[NADH]$ redox couple at a ratio of about 500 to 1500 which corresponds to a voltage of about -0.2 volts. In this way, the cytoplasmic pyridine nucleotides can accept the H^+ and e^- from the substrates or food presented to the cell so that the cell may convert this food or substrate into energy. When the cell is metabolizing very reduced substrates, such as fatty acids, the cytoplasmic $[NAD^+]/[NADH]$ is about 400-800. When the cell is metabolizing carbohydrates or amino acids, it is obvious that these compounds are already partially oxidized. Therefore, the free cytoplasmic $[NAD^+]/[NADH]$ reflects the oxidation level of its substrate and becomes more oxidized in the range of about 800 to 1500.

[0059] The redox state of the free cytoplasmic $[NAD^+]/[NADH]$ couple can be determined by various techniques, such as by measuring the ratio of $[lactate^-]/[pyruvate^-]$ (a) in freeze clamped tissue, (b) in the venous effluent leaving the organ in question, or (c) in the medium bathing the tissue in question. Alternatively $[l\text{-malate}^-]/[oxaloacetate^-]$ or $[-\text{glycerophosphate}]/[\text{dihydroxyacetone P}]$ ratios in tissue may be measured, if desired. The value of cytoplasmic $[NAD^+]/[NADH]$ can then be calculated.

[0060] In healthy living mammals, the ratio of $[L\text{-lactate}^-]/[pyruvate^-]$ is about 6, but can range, under special situations, such as starvation, to about 15 - 20. A $[L\text{-lactate}^-]/[pyruvate^-]$ ratio below about 20, as occurs after ethanol consumption, because of its links to the cytoplasmic $[NAD^+]/[NADH]$, is pathologic. A characteristic in all cells having a low $[NAD^+]/[NADH]$ ratio is believed to be demonstrable (observable) pathologic consequences, such as tissue swelling, low phosphorylation potential, low plasma membrane voltage, and abnormal electrolyte distribution between intracellular and extracellular H_2O .

[0061] Similarly, the redox state of the free mitochondrial $[NAD^+]/[NADH]$ can be determined by various techniques using tissues such as, for example, kidney or liver, by measuring the ratio of $[d\text{-beta-hydroxybutyrate}^-]/[acetoacetate^-]$ (a) in freeze-clamped tissue, (b) in the venous effluent leaving such tissue, or (c) in the fluid bathing isolated such tissues. A determination of the free mitochondrial $[NAD^+]/[NADH]$ in other tissues, such as brain or heart muscle, is more complex, but, in some cases, can be accomplished by measurement in freeze clamped tissue of the $[\alpha\text{-keto glutarate}^-][NH_3^+]/[glutamate^-]$ ratio (see Miller A.L., Hawkins R.A., and Veech R.L.; *J. Neurochem.* 20, 1393-1400, 1973).

[0062] The normal ratio of mitochondrial $[NAD^+]/[NADH]$ is between about 50 and 20, and the normal ratio of $[\beta\text{-hydroxybutyrate}^-]/[acetoacetate^-]$ is about 1.3 to 4. The value of mitochondrial $[NAD^+]/[NADH]$ can then be calculated.

[0063] The redox state of the free cytoplasmic $[NADP^+]/[NADPH]$ couple is, of course, affected by the $[CO_2]$ of surrounding fluids. Because of the lack of substrates which are permeable to the cell wall without significant and variable gradients, this redox state cannot at present be directly and totally regulated other than by the intracellular metabolic links with the cytoplasmic and mitochondrial $[NAD^+]/[NADH]$. (See Krebs H.A. and Veech R.L.; "Pyridine Nucleotide Interrelations", 1969 in The Energy Level and Metabolic Control in Mitochondria in Papa S., Tager J.M., Quagliariello E., and Slater E.C., eds. pp 329-383, Adriatic Editrice, Bari). Thus, for instance, because pyruvate reacts in both cytoplasmic $[NAD^+]/[NADH]$ and $[NADP^+]/[NADPH]$, administration of $[HCO_3^-]/[CO_2]$ and $[L\text{-lactate}^-]/[pyruvate^-]$ within certain narrow limits regulates these ratios because:

$$\frac{[NAD^+]_c}{[NADH]_c} = \frac{[NADP^+]_c}{[NADPH]_c} = \frac{K_{\text{malic enzyme}} \times [malate^{2-}]}{K_{LDH} \times [L\text{-lactate}^-] [CO_2]}$$

[0064] Pyruvate, L-lactate and CO_2 are permeable to cell wall in a simple fashion, as are D-betahydroxybutyrate and acetoacetate, while malate $^{2-}$ and other dicarboxylates are not.

[0065] While the importance of redox state to the maintenance and normalization of intracellular metabolic processes and bioenergetics has long been recognized, there has never been previously, so far as is now known, any attempt to regulate or to normalize the redox state in such mammals (including especially human patients) receiving intravenous therapy, in patients undergoing dialysis, or in patients receiving parenteral nutrition. The present invention provides compositions and methods for regulating and/or normalizing the redox state in mammals (including man) treated here-with.

[0066] Existing electrolyte fluids make no attempt to maintain or normalize cellular redox potentials in any way whatsoever. In fact, most existing electrolyte fluids actually severely distort or make abnormal the redox balance of the cells, resulting in multiple and definable abnormalities. In this way, existing electrolyte fluids distort, such things as, for example, the rate of fat oxidation, the rate of glucose production, the rate of uric acid excretion, the rate of galactose metabolism in milk fed infants, and the like. All of these abnormalities lead to respectively, accumulation of fat in tissue, such as, for example, liver, production of either hyperglycemia or hypoglycemia, gouty crisis, cataracts, and neurological

damage.

B. The phosphorylation Potential

- 5 [0066] Just as the $[NAD^+]/[NADH]$ ratio is defined as a "redox state", by analogy, it is customary to define the energy state of the adenine nucleotide co-enzyme couple as the "phosphorylation potential". Because in living cells ATP, ADP, and HPO_4^{2-} exist in several charged forms, and in various complexation states with Mg^{2+} , it is customary to define these forms as sigma ATP, sigma ADP, and sigma Pi. The phosphorylation potential is thus defined by the relationship $[\text{sigma ATP}]/[\text{sigma ADP}][\text{sigma Pi}]$.
- 10 [0067] It is clear that the reaction of oxidative phosphorylation contains both the redox state of mitochondria and the cytoplasmic phosphorylation potential. While the phosphorylation potential cannot apparently be controlled directly by addition of ATP and ADP to fluids contacting cells, since these compounds do not penetrate cell wall, there is, however, another reaction which is in near-equilibrium with the cytoplasmic $[\text{sigma ATP}]/[\text{sigma ADP}][\text{sigma Pi}]$ (see Veech et al. in *J. Biol. Chem.* 254, 6538-6547, 1979). The reaction involves the two most active enzymes in the glycolytic sequence found in nearly all living cells and catalyzed by the enzymes glyceraldehyde 3-phosphate kinase (EC 2.7.2.3). Veech et al. (reference just cited) provide an equation which defines the relationship between the free cytoplasmic $[NAD^+]/[NADH]$ or redox state and the cytoplasmic phosphorylation state or $[\text{sigma ATP}]/[\text{sigma ADP}][\text{sigma Pi}]$. This relationship is now and accepted by those familiar with this art and is (equation 5):

$$20 K_{G+G} = \frac{[\text{sigma 3-PG}][\text{sigma ATP}]}{[\text{sigma GAP}][\text{sigma ADP}][\text{sigma Pi}]} \cdot \frac{[NADH][H^+]}{[NAD^+]} = 1.83 \times 10^{-4}$$

or

$$25 \frac{K_{G+G}}{K_{LDH}} \frac{[\text{sigma 3-PG}]}{[\text{sigma DHAP}/22]} \cdot \frac{[\text{sigma ATP}]}{[\text{sigma ADP}][\text{sigma Pi}]} \cdot \frac{[l\text{-lactate}]}{[\text{pyruvate}]} = \\ 30 1.65 \times 10^{-7} M^{-1}$$

- [0068] Metabolism in any living cell may be considered to be an ordered process whereby $[H^+]$ and electrons $[e^-]$ are removed from substrates and passed to co-enzyme acceptors which are largely cytoplasmic NAD⁺. This co-factor thus has a potential in the cell for more oxidation at about -0.19 volts than its standard potential of about -0.32 volts so that 35 it may accept these electrons. The H^+ and e^- gathered in the cytoplasm, or even created in the mitochondria, may then be transferred to mitochondria by mechanisms involving other substrates to mitochondrial NADH which has a lower potential of about -0.28 volts in most mammalian cells. If e^- and H^+ are produced with a higher voltage, such as for example, from the oxidation of succinate or fatty acids, they form reduced FADH₂ from FAD which has a more oxidized potential and therefore less potential energy. H^+ and electrons produced from NADH-linked substrates produce 3 ATP for each $1/2 O_2$ consumed while those from flavo-protein (FAD) acceptors produce only 2. This difference in energy is due to the fundamental difference in the chemical reactions involved in producing the H^+ and e^- .

- [0069] The fundamental process of cell respiration where NADH is oxidized to form heat and energy is called oxidative phosphorylation. It occurs in cellular organelles called mitochondria in a series of redox reactions called the electron transport chain. The mitochondrial electron transport system takes two electrons $[2e^-]$ from substrates and passes 45 them up the chain to reduce $1/2 O_2$ forming H_2O . The energy realized in this process is conserved in the cell in a chemical form of anhydride bond in the terminal phosphate group of adenosine triphosphate (ATP). The formation of three pyrophosphate bonds of ATP leads to the formation of H_2O and requires $3H^+$ in addition to the formation of the $1 H_2O$ formed from NADH plus H^+ plus $2 e^-$ taken from the substrates being oxidized by the cell. The reaction of oxidative phosphorylation is a spontaneous one (see Veech et al in cited reference).

- [0070] The phosphorylation potential of living cells can be measured by determining the cellular contents of the components of certain metabolites (see Veech R.L., In *J. Biol. Chem.* 254, 6538-6547, 1979). In certain tissues, such as brain, heart, or skeletal muscle, measurement of the components of the creatine kinase reaction (EC 2.7.3.2) may be used as the preceding reference describes.

- [0071] Since on theoretical grounds Veech et al. in *J. Biol. Chem.* 254, 6538-6547, 1979 showed that $[\text{creatinine}]/[\text{creatinine-P}]$ is in near equilibrium with the cytoplasmic $[\text{sigma ATP}]/[\text{sigma ADP}]$, it follows that the phosphorylation potential in skeletal muscle or brain may be evaluated in living human patients by measuring the $[\text{sigma CRP}]/[\text{sigma Pi}]$ ratio without resorting to freeze-clamping of organs by the use of ^{31}P NMR (nuclear-magnetic resonance) as has been done by Chance and others (see Chance B., et al., *Proc. Nat'l. Acad. Sci., U.S.* 78 6714-6718, 1981). The agreement

between the necessarily destructive methods heretofore used in animals by Veech, and the somewhat less precise but nonharmful methods of sigma creatine-P/sigma Pi measurements with ^{31}P NMR, demonstrate that the normal value of the phosphorylation potential or $[\sigma\text{ATP}]/[\sigma\text{ADP}][\sigma\text{Pi}]$ as estimated by Veech is essentially correct (as stated above). Further, the increasing availability of ^{31}P NMR facilities in academic medical centers ensures that measurements in living human patients can be conducted without harming them.

[0072] Because the cytoplasmic $[\sigma\text{ATP}][\sigma\text{ADP}][\sigma\text{Pi}]$ or phosphorylation potential is related to the cytoplasmic $[\text{NAD}^+]/[\text{NADH}]$ or redox state by a near-equilibrium reaction catalyzed by glyceraldehyde-3-phosphate dehydrogenase and 3-phosphoglycerate kinase, it is possible to alter and regulate and normalize the phosphorylation potential of a living cell by affecting its redox state (as is believed to be accomplished in the present invention).

[0073] If a simple, reliable chemical means were known and/or could be devised to change the intracellular redox state, it would of necessity have to change the other components of the reaction which include the phosphorylation potential and would be of obvious fundamental importance in medicine and in many other related fields of biochemistry, physiology, molecular biology, tissue culture, veterinary medicine, and like endeavors. Such a chemical means is provided by the teachings of the present invention.

C. Redox Active Metabolites

[0074] As above indicated, a large portion of metabolism is devoted to energy generation which involves the removal of H^+ and e^- from substrates in cytoplasm or mitochondria for delivery to mitochondrial electron transport scheme for conversion of 2H^+ plus $2e^-$ with $1/2 \text{O}_2$ to yield H_2O with the liberation of about 1 volt or 54 kcal/mole of energy which is conserved in the $[\sigma\text{ATP}][\sigma\text{ADP}][\sigma\text{Pi}]$ couple. In mammalian cells, the $[\sigma\text{ATP}][\sigma\text{ADP}][\sigma\text{Pi}]$ has a delta G(free energy in kilocalories per mole) of between -13.6 and 014.1 Kcal/mole, the transfer to this H^+ and e^- is accomplished by a series of co-factors, the major one being NAD (nicotinamide adenine dinucleotide) and its phosphate (called NADP). Oxidation is defined as the removal of electrons, and reduction as the addition of electrons. The removal or addition of e^- plus H^+ from substrates is catalyzed by enzymes, the major group of which are called dehydrogenases, as indicated above. The enzymes (catalysts) control the rates at which reactions occur, but the extent and the direction of a reaction, and the amount of energy (delta G) which may be liberated by a reaction, is determined by the inherent energy in the chemical bonds (ΔG°) and the concentrations of the reactants and products.

[0075] Determination of any redox or energy state must always involve a ratio of chemical compounds, $[\text{oxidized product}]/[\text{reduced reactant}]$ and $[\text{oxidized co-factor}]/[\text{reduced co-factor}]$. The overall reaction is thus comprised of two individual redox systems, one of which is oxidized, while the other is reduced.

[0076] Those enzymes within a cell which are of sufficiently high activity relative to the flux through the enzyme to catalyze a state of near equilibrium are suitable for controlling the redox state. A reaction may be experimentally determined to be in a state of near-equilibrium by measuring the equilibrium constant (K_{eq}) under conditions which approximate those existing within a cell, that is, where the ionic strength I equals 0.25, the pH equals 7 to 7.2, the temperature equals 38°C , and the free $[\text{Mg}^{2+}]$ equals 0.5 to 1mM, and also where I equals $1/2$ sigma molarity of ions times the valence of ions. With knowledge of the value of K_{eq} , the concentration of the reactants in a tissue may be measured in rapidly frozen tissue. If the value of $[\text{product}]/[\text{reactant}]$ measured, in several different dehydrogenase reactions, gives the same calculated free $[\text{NAD(P)}^+]/[\text{NAP(P)}]$ ratio, then the reaction is said to be in "near-equilibrium" under in vivo conditions. In the case of near-equilibrium dehydrogenase reactions, addition of a predetermined amount of a ratio of product/reactant allows one to set the $[\text{NAD}^+]/[\text{NADH}]$ ratio within the cell at a predetermined level, provided the reactants penetrate-the cell wall freely or in a constant ratio one to another. The redox state or $[\text{NAD(P)}^+]/[\text{NAD(P)}]$ ratio may be set inside a cell by controlling the $[\text{CO}_2]$ and the redox state of the cytoplasmic free $[\text{NAD}^+]/[\text{NADH}]$ as described previously. Each of the three couples disclosed herein is a near equilibrium couple.

[0077] Various cytoplasmic and mitochondrial NAD-linked dehydrogenases appear to be capable of controlling or setting the $[\text{NAD}^+]/[\text{NADH}]$ ratio in each of cytoplasm and mitochondria. Because of the special permeability of the complete couple of L-lactate /pyruvate $^-$ for cytoplasm and D-B-hydroxybutyrate $^-$ /acetoacetate for mitochondria, these two redox couples are preeminently well suited for the practice of this invention. This is so because: 1) both manovalent anions in the pair distribute themselves equally between plasma and cellular H_2O ; 2) changes in distribution of anions between extracellular and intracellular H_2O during pathological states will effect both members of the couple equally through preserving the integrity of the given redox state; 3) both couples react with "dead end" branches off the main metabolic sequences; 4) the concentration of these normal transport metabolites can reach very high levels in plasma of normal healthy mammals under physiological conditions; and 5) the members of both couples each contain a charge which can be used to normalize the low $\text{Na}^+:\text{Cl}^-$ milliequivalent ratio characteristic of most I.V. (intravenous) solutions.

[0078] The near equilibrium redox active metabolite carboxylate couples disclosed herein, specifically, L-lactate/pyruvate $^-$ and d-betahydroxybutyrate $^-$ /acetoacetate $^-$, constitute safe entry points and appear to be unusual in their ability to not only normalize the redox state in cytoplasm through the reaction of L-lactate and pyruvate with LDH, but also to regulate the redox state in the mitochondria through reaction of d-betahydroxybutyrate and acetoacetate with the

enzyme d-betahydroxybutyrate dehydrogenase (EC 1.1.1.30) which is apparently present in most tissues at a high enough activity to maintain near-equilibrium conditions at most times.

[0079] As indicated above (see Table 1 and related text), previous attempts to normalize the sodium to chloride milliequivalent mole ratio of about 1.36 were usually done by adding either (d,l) lactate⁻ or acetate⁻, or a combination of lactate and acetate, or other inappropriately paired carboxylate anions, leading inevitably in all known instances to severe and measurable pathological consequences.

[0080] In the solutions disclosed herein, one employs at least one of the above indicated three different near-equilibrium couple mixtures. In each couple mixture, the two member components are employed in a definite milliequivalent ratio relative to one another. Such a ratio is needed in order to control either the plasma pH, or the redox state (and consequently the phosphorylation potential), or both.

[0081] Among the possible mixture couples which could be used, these three couples were selected because, for each couple:

1. The distribution of ions between extracellular fluid and intracellular fluid is predictable in all normal and pathological states.
2. It is capable of achieving and regulating a predetermined redox state and phosphorylation potential within most living cells.
3. At least one member thereof contains an anionic charge.
4. It can be given in aqueous solution form so that the total levels administered do not substantially exceed total levels found under normal physiologic conditions in mammalian blood (plasma).
5. Both members thereof constitute safe entry points which enter the metabolic sequence and pathways at a safe entry point and these safe entry points, are at dead end terminals in the metabolic pathways, thus avoiding any possibility of a pathologic buildup of metabolites with the consequence that a disordering of cellular metabolism would consequently result.
6. It need not induce a change in water distribution between intracellular and extracellular space.
7. It may be osmotically neutral in most tissues.
8. Administration permits control of water distribution as a result of changing redox and hence the linked phosphorylation state and the magnitude of the extracellular Na⁺ Donnan forces generated thereby.

[0082] When blood levels of, respectively, L-lactate/pyruvate, d-betahydroxybutyrate/acetoacetate, and bicarbonate/CO₂ are maintained within their normal limits, then the redox state, the phosphorylation state, and the plasma pH each tend to be normalized which is achieved as a result of administration of a solution of this invention.

[0083] Intracellular concentration of each member of each couple is achieved through the extracellular fluid because each of the monovalent anions chosen, namely, L-lactate and pyruvate, d-betahydroxybutyrate, and aceotacetate, and also bicarbonate, distribute themselves between plasma water, extracellular water, and intracellular water in concentration ratios or gradients which are the inverse of the hydrogen ion (concentration), thereby achieving a gradient or ratio of about 1.35 between extracellular and intracellular fluid. The nonionic dissolved CO₂ distributes itself substantially equally between extracellular fluid and intracellular fluid.

[0084] Those learned in the art realize a redox state must be defined at a certain pH, or [H⁺] ion concentration. The near-equilibrium couple [HCO₃⁻]/[CO₂] defines the cellular pH or [H⁺] concentration. This near-equilibrium couple is therefore an integral part of the redox state. Preferably the level of sigma [HCO₃⁻] plus [CO₂] present in any given solution disclosed herein may vary under normal physiological conditions from 10mmol/l to 40mmol/l, but in general, is (when present) in the range from 25 to 35 mmol/l. The milliequivalent ratio of [HCO₃]/[CO₂], of course, in effect, is defined so as to give a [H⁺] ion concentration, or pH, in the physiological range as defined above.

[0085] The redox and phosphorylation states in various tissues in the rat have been given by Veech et al. *J. Biol. Chem.* 254, 6538-6547, 1979 and for the redox states in Veech, Eggleston and Krebs, *Biochem. J.* 115, 609-619, 1969. The same general principles are believed to hold for man, but cannot be directly proved since freeze clamping is not possible. MNR measured estimates of the phosphorylation potential in brain and muscle in living humans, however, agree well with these figures derived by freeze clamping procedures.

[0086] By the term "plasma" or "blood plasma" as used herein conventional general reference is had to the liquid part of the blood as distinguished from the corpuscles. Plasma can be prepared by various techniques well known to those familiar with this art typically using centrifugal force to separate a supernatant (which is plasma) after non-coagulated blood is centrifuged.

[0087] By the term "extracellular fluid" as used herein conventional general reference is had to all body fluids in extracellular spaces outside of the circulatory system (e.g. the blood) and outside of intracellular fluid in a mammal (typically constituting about 15% of the weight of a mammal).

[0088] By the term "intracellular fluid" as used herein conventional general reference is had the fluid within cells which constitutes about 57% of total mammalian body weight.

[0089] It is well known that (see Black DAK. Lancet i 305-12 1953) infusions into a mammal of large amounts sodium and chloride in a solution milliequivalent ratio of 1 to 1 lead inherently to hyperchloremic acidosis. This knowledge lead to the development of such well known solutions as lactated Ringers, and also to the compositions used in most dialysis solutions, wherein, in a majority of cases, the sodium to chloride milliequivalent ratio is normalized compared to plasma values by the addition of various organic anions (as described above). These organic anions chosen in the prior art are as described above. In no known prior art case, however, were any solutions with a normalized Na:Cl milliequivalent ratio produced which did not use organic ions in such a way as to inherently lead to severe and measurable metabolic abnormalities and pathologic consequences. Mixtures of redox pairs nor $\text{HCO}_3^-/\text{CO}_2$ were not generally used to normalize the $\text{Na}^+:\text{Cl}^-$ ratio nor were the reasons known why a choice of near equilibrium matched couples was desirable.

5 Correction of this ratio between sodium cation and chloride anion by the mixture couples as disclosed herein eliminates the pathologic consequences of all the prior art electrolyte solution compositions. In addition, the solution compositions of this invention tend to normalize plasma inorganic electrolyte composition and to correct the anion gap which in many instances could not be accomplished by prior art electrolyte solutions.

[0090] Thus, in summary, the compositions of this invention tend to normalize (a) plasma pH, (b) composition of major plasma inorganic electrolytes, (including the milliequivalent ratio of Na^+Cl^- and the anion gap), (c) the redox state, and (d) the phosphorylation potential. These normalizations are obtained and achieved without the abnormal, pathological consequences inherent in all known prior art solutions. No other man-made solutions are presently known which will accomplish this combination of results.

20 D. Other Possible Benefits (Theorized).

[0091] It is theorized, and there is no intent to be bound by theory herein, that the solutions of the present invention, in addition to the properties above described, further tend to normalize at least one of the following states:

- 25 1. Distribution of water between intracellular and extracellular compartments,
 2. Distribution of major inorganic electrolytes between intracellular and extracellular fluid,
 3. Transmembrane cellular potential, and
 4. The degree of organization within the living cell or its entropy.

30 [0092] The ratio of the chemical activity of free water on each side of a typical normal mammalian cell membrane is always unity. Movement of water across such a cell membrane is achieved by the movement of osmotically active substances. Changing the cellular phosphorylation potential, through the NaK ATPase, therefore,

[0093] Transmembrane cellular potential can be measured by known (e.g. conventional) techniques: such as with electrodes or probes, and the like. Calculation of such cellular voltage can be achieved from a measurement of the distribution of chloride ions between intracellular and extracellular fluid following Nernst's law.

35 [0094] A quantitative relationship is theorized to exist involving redox state, phosphorylation potential and the above referenced three states. This relationship may be expressed by the following equation:

40 (7.)

$$\Delta G = 0 = \Delta G_{\text{ATPase}}^{\circ} + \Delta G_{\text{O}}^{\circ} \frac{[\text{Na}^+]_{\text{O}}}{[\text{Na}^+]_{\text{i}}} \dots + RT \ln \frac{[\text{ADP}] [\text{Pi}]}{[\text{ATP}]} + RT \ln \frac{[\text{Na}^+]_{\text{O}}^3 [\text{K}^+]_{\text{i}}^2 [\text{Cl}^-]_{\text{O}}}{[\text{Na}^+]_{\text{i}}^3 [\text{K}^+]_{\text{O}}^2 [\text{Cl}^-]_{\text{i}}} + T \Delta S$$

50

wherein

[0095] The values of the various terms in the foregoing equation of are given as follows (for muscle and brain):

55 (7.) $\Delta G = 0 - 7.73 \text{ kcal/mol} + 0 = (-6.3 \text{ kcal/mol}) + 8.4 \text{ kcal/mol} + 5.6 \text{ kcal/mol}$

[0096] In the foregoing equation, the phosphorylation potential is shown to be in a state of near equilibrium with the substrates of the sodium potassium ATPase. Since the chloride ion is cell wall permeable, this ion distributes itself in

conformity with the transmembrane cellular potential. Movement of three sodium ions out of the cell and two potassium ions into the cell across the cell membrane necessarily results, from the law of electrical neutrality, in the movement of one chloride ion from inside the cell to outside the cell across the cell membrane. This makes the sodium potassium ATPase, in effect, an osmopump resulting in the export of two milliosmoles per ATP hydrolyzed. This pump is electro-neutral.

[0097] The T delta S term, which is approximately 5.6 kilocalories per mole of ATP hydrolyzed, is an entropy term. It, therefore, refers the state of randomness within the cell. The positive nature of this entropy term indicates that a high degree of order is imposed on the intracellular environment. In terms of quantum and statistical mechanics, the number of ways of achieving a certain energy state is called its degeneracy (Ω). The Boltzmann equation defines S (or entropy) as $S = K_B \ln \Omega$, where Boltzmann's constant (which relates the gas constant to Avogadro's number), or $K_B = 1.38 \times 10^{-23} \text{ J}^\circ\text{K}$.

[0098] It follows from the foregoing equation 7, above, that the distribution of calcium inside the cell is a function of the cube of the respective sodium concentrations inside and outside of the cell because of the action of the high-activity sodium-calcium exchange enzyme. The following equation shows the relationship:

$$K_{\text{Na}/\text{Ca}} = \frac{[\text{Na}^+]_i^3 [\text{Ca}^{2+}]_o [\text{Cl}^-]_i}{[\text{Na}^+]_o^3 [\text{Ca}^{2+}]_i [\text{Cl}^-]_o}$$

where:

$[\cdot]_i$ ~intracellular concentration in cytoplasmic H_2O

$[\cdot]_o$ ~concentration in extracellular H_2O .

[0099] Unlike the simple NaK ATPase which moves 2 mOsmoles out of the cell thus moving H_2O with it, the result of moving Ca^{2+} out of the cell by the Na-Ca exchanger is to move a net of 3 mOsmoles into the cell, thus increasing the cells water content. The NaK ATPase must then operate again to move the excess sodium out in exchange for K^+ to restore osmotic equilibrium between extracellular space H_2O and cell H_2O .

[0100] The net result of the foregoing equation (7) is that the water of both intracellular and extracellular fluid is a function of the sodium/potassium ATPase (EC 3.6.1.3) and also of the phosphorylation potential.

[0101] It can be empirically seen that the voltage across a cell membrane is inversely related to the chloride distribution and the phosphorylation potential.

[0102] Correlation between phosphorylation potential, intracellular chloride and transmembrane cellular potential for various mammalian tissues is illustrated by Table II below:

Table IIIa
Correlation between Phosphorylation Potential,
Intracellular Chloride and Transmembrane Cellular
Potential.

	$[\Sigma \text{ATP}]$ $[\Sigma \text{ADP}] [\Sigma \text{Pi}]$	$[\text{Cl}^-]_i$ mEq/l M^{-1}	ΔE mV
red cell	7,000	90	- 9
liver	15,000	40	- 40
brain or muscle	30,000	7-9	- 70

[0103] From Table II, it is seen that low phosphorylation potential correlates with a high intracellular chloride, and a low transmembrane cellular potential correlates with the inherent setting of the potential as a function of the Donnan-active material within the cell with the phosphorylation potential merely overcoming the Donnan forces so as to export two milliosmoles, as described in equation 7.

[0104] Because of the voltage dependent permeant nature of chloride ion to most non-epithelial tissues (Ho, MK, Guidotti G. *J. Biol Chem* 250: 675-683, 1975) the induction of high extra cellular chloride, such as occurs, for example, in current intravenous electrolyte therapy, must have profound pathological consequences for the metabolism of the

cell, even though the purpose of such intravenous and dialysis therapy is to normalize the water and electrolyte concentrations of the various mammalian body cellular compartments. This is so because the ratio

$$\frac{[Na^+]_o^3 [K^+]_i^2 [Cl^-]_o}{[Na^+]_i^3 [K^+]_o^2 [Cl^-]_i}$$

and the TΔS term link the cellular phosphorylation and the cellular redox states to intracellular and extracellular water and the electrolyte concentrations of Na⁺, K⁺, Cl⁻ and also Ca²⁺.

E. Electrolyte Solution Preparation

[0105] The electrolyte solutions of the present invention can be prepared by any convenient or conventional procedure.

[0106] As a matter of accuracy, the compositions of this invention can be described in terms of their ion contents which can be expressed either in terms of millimoles per liter of solution, or milliequivalents per liter of solution. It is standard practice in this art in describing a given solution to separate anions from cations, and nonionics from ionic materials; this practice is followed herein in the main. As those skilled in the art will readily appreciate, a translation or conversion of millimoles per liter of solution, or of milliequivalents per liter of solution, into grams of a given salt added per liter of water is routine and is given in any standard text book in the field, such as, for example, "Data For Biochemical Research" (1969) (Dawson R.M.C., Elliott W.H., Jones KM., Eds). Clarendon Press, Oxford at pages 507 and 508. This reference illustrates not only the salt starting materials, but also the order of addition of same in the preparation of certain illustrative prior art electrolyte solutions shown therein. Solutions of this invention are readily prepared by this type of procedure. The particular salt combination used for a given solution may change from time to time in a manufacturing operation as those skilled in the art well know. The significant factor is that the final concentrations of respective component ions in any given solution remain as specified or desired. In view of the developed state of this art, no detailed description of electrolyte solution preparation procedures is believed to be necessary or desirable herein.

[0107] The solutions of this invention, and the component materials incorporated thereto, are, in general, formulated, as to contain a combination of the desired physiological Na⁺:Cl⁻ milliequivalent ratio normativity, the claimed near-equilibrium couple, and other components.

[0108] Thus, various initially existing pathological conditions can be ameliorated by practice of the processes and the compositions of the present invention, depending upon the particular solution used and the particular use conditions and circumstances in any given use situation. Thus, by this practice of this invention, one can accomplish in a physiologically acceptable manner the removable of metabolic products from cellular water, the replacement of body fluids and electrolytes, and the administration of nutrients, and the like, as desired. The solutions may be administered in any fashion desired so long as they contact living mammalian tissue. Administration can be accomplished by any convenient technique, such as for examples, intravenously, intraarterially, intradermally, intrathecally, orally (especially when the solution contains the nonbicarbonate containing couples), across a semi-permeable membrane, or the like, as those skilled in the art will readily appreciate. The solutions of this invention as prepared are, in general, well suited for the administration of therapeutic agents to living mammals.

[0109] When bicarbonate anions are not present, then the level of combined (or sigma) L-lactate/pyruvate and/or D-betahydroxybutyrate/acetoacetate present in a solution disclosed herein is optionally greater than when bicarbonate is present in order to achieve the desired milliequivalent ratio of sodium to chloride, as indicated. The concentration of either sigma L-lactate/pyruvate and/or of D-betahydroxybutyrate/acetoacetate in a given solution disclosed herein can thus range up to the full maximum quantity desired (within the limits described herein). It is presently preferred, particularly when no bicarbonate is present, to employ a mixture of L-lactate/pyruvate with a mixture of D-betahydroxybutyrate/acetoacetate.

[0110] Those skilled in the art will realize that in any given solution disclosed herein one can incorporate an excess of one or more individual members of any one mixture couple so that (a) the ratio of one member to the other of any given couple and (b) the total quantity of both mixtures or members lies outside of the ranges hereinabove described. Such a single member excess is not recommended when practicing the method disclosed herein. However, if such a single member excess does occur, the amount of the excess can be calculated by determining the maximum ratio of one couple member to the other which can be present in accord with the above teachings, and then the quantity of one couple member remaining (or present) which is outside of this ratio range may be considered to constitute an excess. The effect of such an excess is evidently merely to cut down, but not to eliminate, the efficacy of what effect would otherwise be obtained by using only a solution which contains mole ratios and quantities of respective mixture couples disclosed herein.

[0111] In the making of solutions disclosed herein, it is preferred to employ the optically active L-lactate salts or L-lactic acid (which will make the desired L-lactate anions in solution), and also similarly to employ D-betahydroxybutyric acid or D-betahydroxybutyrate salts (which will make the desired D-betahydroxybutyrate anions in solution). Choice of particular salt or acid (or mixture) used in any given case depends among various factors, such as upon the other starting inorganic salts which a formulator desires to use (based upon availability, cost, and like factors), all as will be readily appreciated by those skilled in the art. Racemic (D-L) mixtures could be used, but their use is preferably avoided since these unnatural isomers are known to be associated with specific toxic effects. Racemates can be metabolized. If such are used, the ratios of one member to another in the respective near equilibrium couples involved should be based upon the quantity of particular optically active form present (e.g. either [L-lactate⁻] or [D-betahydroxybutyrate⁻]), as the case may be.

[0112] In the sections disclosed herein at the pH ranges described, not all couple member material of any given couple will be in an ionized (anionic or dissociated) form; a portion of this material will be in an unionized (undissociated) form. Typically, the quantity of undissociated material (such as L-lactate acid, pyruvic acid, D-betahydroxybutyric acid, acetoacetic, sodium bicarbonate, carbonic acid, or the like) is not more than about 0.1% of the total quantity of all material of any given species (e.g. L-lactate, pyruvate, D-betahydroxybutyrate, acetoacetate, or bicarbonate). For purposes of calculating a milliequivalent ratio, molar concentration, or the like, it is preferred to base computations upon the total material of any given species which is present in a solution of this invention.

[0113] The carbon dioxide, when used, can be introduced either as a gas, preferably using conventional aeration apparatus to effect a solubilization of CO₂ in a solution, or it can be generated *in situ* from a dissolved metal (such as sodium (preferred), potassium, calcium or magnesium) salt of bicarbonate in combination with a dissolved acid (lactic, pyruvic, betahydroxybutyric, or acetoacetic) in respective proportions of each such that the total quantity of dissolved carbon dioxide so generated is within the ranges described herein for use in a solution of this invention.

[0114] As elsewhere indicated herein, if desired, a solution of this invention can also contain various known additives in concentrations taught by this art, but it is presently preferred not to employ anions and nonionics which will not be safe entry points.

[0115] In general, a solution disclosed herein should contain as a minimum a total of sigma (lactate/pyruvate and/or sigma betahydroxybutyrate/acetoacetate) and/or sigma bicarbonate/carbon dioxide which is at least about 0.5 millimoles per liter as indicated. Below these levels, benefits in normalization of body metabolism as explained above are apparently achievable, but such benefits become increasingly difficult to demonstrate and prove by state of the art techniques of measurement. Consequently, it is preferred to avoid, if possible, homeopathic possibilities by using minimum concentrations as above indicated.

[0116] When bicarbonate is present, the total quantity of sigma (lactate/pyruvate and/or betahydroxybutyrate/acetoacetate) used can generally be reduced which is now believed to be desirable. Thus, when bicarbonate is present, the total sigma (L-lactate/pyruvate and/or D-betahydroxybutyrate/acetoacetate) is preferably about 2 to 17 millimoles per liter.

[0117] When a solution of this invention contains at least one osmotically active substance (preferably metabolizable and nonionic), it is added to provide nutritional or osmotic requirements. Since it is uncharged, it does not therefore contribute to normalizing the Na⁺:Cl⁻ ratio or to correcting the anion gap.

40 F. Classification and Usage of Electrolyte Solutions

[0118] All of the formulations of this invention from a composition viewpoint fall into what can be regarded generally as being either one of two distinct classes:

45 Class I which comprises fluids containing at least one and not more than two metallic cations selected from the group consisting of sodium, potassium, calcium and magnesium, while
Class II which comprises solutions containing at least three and typically not more than four metallic cations selected from the same group.

50 [0119] Class I fluids are typically administered at dose levels which are not greater than about 1 liter per human adult patient per 24 hour day, one typical dose level being 500 ml per such patient per 24 hour day.

[0120] Class II fluids are typically administered at dose levels chosen by the physician, and these levels can range from 0 to greater than 100 liters per human adult patient per 24 hour day, depending upon circumstances.

55 [0121] Each of the inorganic electrolytes present in a solution of this invention is typically present in an amount of at least 0.5 mmol/l thus clearly qualifying them as "electrolytes" as such rather than as trace metals, such as is associated with levels of iron, manganese, zinc and the like in normal plasma and which trace metals can be present in normal plasma at levels less than 0.4 mmol/l. If desired, of course, trace materials can be added to solutions of this invention.

[0122] Each of the cations sodium, potassium, calcium, and magnesium and each of the anions bicarbonate, chloride, and phosphate are normally found in the plasma and tissue of mammals at concentration levels greater than or equal to about 1 millimolar per liter of body fluid (see Table I). The solutions of this invention, in general, contain respective inorganic electrolyte concentrations which resemble the corresponding concentrations of such electrolytes in plasma (when any one of such electrolytes is present in any given solution of this invention).

[0123] Class I solutions are useful as intravenous solutions for electrolyte and fluid therapy especially where no more than about 10% of total blood volume (about 500 ml in an adult human) is to be administered over a 24 hour day. Solutions of this type have been used in the treatment of hemorrhagic shock where 2400 mOsmolar NaCl solutions have been advocated. (See Velosco IT, Pontieri V, Rocha M, Silva E, Lopes OU. *Am J Physiol* 239: H664-673, 1980).

[0124] Class II solutions find use in intravenous applications where over 10% of total blood volume (about 500 ml in an adult human) is needed to be given to a human adult over a 24 hour day. Administration can be made, for example, to a normal human with an impairment or injury, such as loss of limb or the like, or to a human with impaired renal excretion: Class II solutions can be used as an improvement for lactated Ringer's solution.

[0125] Class II solutions also are useful in dialysis, peritoneal, ambulatory peritoneal dialysis or hemodialysis, where perhaps 120-160 liters per hemodialysis day per patient are used. Such solutions can be used improve existing acetate or lactate containing solutions, but use of acetate is not desired in the practice of this invention.

[0126] Given the solutions of this invention, a physician may henceforth wish to administer normal or hypertonic saline solution only to correct a condition of metabolic alkalosis since giving $\text{Na}^+:\text{Cl}^-$ in a 1:1 milliequivalent ratio causes acidosis and other disturbances recognized herein. The solutions described herein improve, normal saline solution.

[0127] Solutions of Class II can be used as such, or can be employed as diluent for plasma extenders or for reconstituted frozen blood. For example, dehydrated plasma can be dissolved and dispersed in a solution of Class II so as to produce an injectable solution, as those familiar with the art will appreciate.

[0128] Each one of these Class I and II solutions can be considered to be characteristically comprised of four sub-groups which can be stated briefly as follows:

- A. Solutions containing only inorganic ions and one or more of our near-equilibrium couples of organic anions pairs with which chloride anions are included.
- B. Solutions containing in addition to such inorganic ions and organic ion pairs a mixture of bicarbonate and carbon dioxide.
- C. Solutions containing such inorganic ions and organic ion pairs plus non ionic materials.
- D. Solutions containing in addition to the inorganic ionic material both mixtures of bicarbonate and carbon dioxide (as characterized in B above) plus other nonionics (of the type characterized in C above).

[0129] As indicated above, avoidance of substances in solutions of this invention which do not constitute safe entry points is preferred. For example, use of such nonionic osmotically active substances as fructose and glycerol are preferably avoided and are not recommended for use in the practice of this invention. Also, avoidance of the organic anions used in the prior art which are not safe entry points is recommended, including use of lactate alone, acetate alone, lactate and acetate together, gluconate, citrate, and the like.

[0130] Prior art in dialysis fluids show that the composition of the fluids now commercially used evidently is intended to approximate that of plasma with the proviso that the anion gap is typically corrected with abnormal amounts of typically acetate or lactate. The suggestion has also been made in the prior art dialysis fluid composition should approximate the composition of interstitial (extracellular) fluid. While such compositional approximations now appear to be essentially incorrect especially from the standpoint of achieving dialysis fluids of maximal safety and utility and patient benefit, it is submitted that such approximations can be substantially benefitted by compounding dialysis solutions in accord with the teachings of the present invention (both for hemo- and peritoneal dialysis).

[0131] Solution compositions of the present invention of Class I and Class II are generically characterized herein above. The following Table III includes preferred solutions of this invention in terms of composition at the time of administration (e.g., water having dissolved therein each of the indicated components in the respective amounts indicated).

[0132] With regard to the term "nonionics" in a solution or process of this invention, those skilled in the art will appreciate that this term connotes no net charge on the molecule involved at the particular solution pH specified.

[0133] Solutions of this invention can be prepared as concentrates which at 0.8 molar solutes or greater will inhibit bacterial growth, as those skilled in the art will appreciate, and such concentrates can then be diluted with water before administration to prepare compositions of this invention.

[0134] In general, solutions of this invention are believed to be preparable so as to be storage stable for periods of time at least sufficient to permit packaging, intermediate storage in sealed containers, followed by administered.

Table III

Generic Compositions of Class I and Class II Solutions			
Component	Composition of time of Administration Quantity Range (millimoles per liter)		
	broad	preferred	
Total cations (mEq/L)	1 to 2400	130 to 170	
(1) sodium +	1 to 2400	130 to 165	
(2) potassium+	0 to 90	0 to 5	
(3) calcium++	0 to 60	0 to 1.5	
(4) magnesium	0 to 15	0 to 1	
Total anions (mEq/L)	1 to 2400	130 to 170	
(5) chloride-	0.6 to 1940	80 to 130	
(6) bicarbonate-	0 to 465:	0 to 60	
(7) sigma 1-lactate- plus pyruvate	0 to 465	0 to 60	
(8) sigma d-betahydroxybutyrate/plus acetoacetate-	0 to 465	0 to 60	
(9) sigma (6+7+8)	0.1 to 465	25 to 65	
Total nonionics	0 to 2400	0 to 305	
(10) carbon dioxide	0 to 25	1 to 5	
(11) osmotically active substances*	0 to 2400	0 to 300	
In Table III solutions the component interrelationships are always such that the following holds:			
(12) mEq.ratio of bicarbonate- / CO ₂	0.1/1 to 55/0.1	0.1 to 55/0.1	
(13) mEq.ratio of 1-lactate / pyruvate	20/1 to 1/1	10/1 to 5/1	
(14) mEq.ratio of d-betahydroxybutyrate-/ acetoacetate	6/1 to 0.5/1	3/1 to 1.5/1	
(15) mEq.ratio of Na:Cl	1.24 to 1.60	1.24 to 1.6	
(16) Osmolarity of Solution	260 to 5000	280 to 545	
(17) pH of Solutions	5 to 9	5 to 9	

* Glucose preferred

[0135] Optionally, solutions as shown in Table III can additionally contain:

- (a) from 0 to 25 millimoles per liter of sigma inorganic phosphate (e.g. all inorganic phosphate, including mono-, di-, and trivalent phosphate ions), and
- (b) from 0 to 2 millimoles per liter of sigma inorganic sulfate (e.g. all inorganic sulfate including non ionized dissolved salts).

[0136] The electrolyte solutions of such Table III, as indicated above, are useful in such applications as intravenous administration for replacement of electrolytes and fluids, for parenteral nutrition, for dialysis, and the like. For a particular field of use and/or end use applications, the formulation of any given solution can be optimized in accord with the desires of the formulator. Thus, in general, there is disclosed an in vivo process which

- (a) tends to maintain a normal plasma milliequivalent ratio of sodium cations to chloride anions,
- (b) tends to maintain normal plasma and cellular pH, and
- (c) tends to maintain normal cellular cofactor ratios (that is, tends to maintain and regulate a normal cellular redox state and a normal cellular phosphorylation potential).

This process is practiced by introducing into a living mammal a physiologically effective amount of an aqueous solution as above characterized. Introducing can be accomplished by any known procedure as herein indicated. The physiologically effective amounts are as herein indicated.

[0137] Class I solutions which are particularly suited for electrolyte and fluid therapy are subgenerically characterized

in Table IV below. Each Table IV solution comprises water which has dissolved therein each of the indicated components in the respective amount indicated. In this Table IV the "preferred" class of embodiments (so identified) can be regarded as being usable either as such, or as concentrates which can be further diluted so long as nonionic material is included to keep the final osmolarity above about 260/mOsmoles/l. In the latter case, the diluted solutions should contain added dissolved nonionic material (preferably glucose) with care being taken to preserve in the product diluted solution the various ratios, osmolarity and pH values, all as shown in such Table IV.

[0138] Such Class I solutions can be used in an in vivo process for accomplishing electrolyte and fluid therapy in a mammal. This process:

- 10 (a) tends to maintain a normal plasma milliequivalent ratio of sodium cations to chloride anions,
- (b) tends to maintain normal plasma and cellular pH, and
- (c) tends to maintain normal cellular cofactor ratios.

[0139] This process comprises introducing intravenously into a mammal at a physiologically effective rate a quantity of such a solution in an amount which is not more than about 1 liter per 70 kilograms of mammal body weight per 24 hour day.

Table IV

Class I Solutions Particularly Suited for Electrolyte and Fluid Therapy			
Component	Composition at time of Administration Quantity Range (millimoles per liter)		
	broad	preferred	
Total cations (mEq/L)	1 to 2400	130 to 170	
(1) sodium ⁺	1 to 2400	130 to 165	
(2) potassium ⁺	0 to 90	0 to 10	
(3) calcium ⁺⁺	0 to 60	0 to 5	
(4) magnesium ⁺⁺	0 to 15	0 to 3	
Total anions (mEq/L)	1 to 2400	130 to 170	
(5) chloride ⁻	0.6 to 1935	80 to 130	
(6) bicarbonate ⁻	0 to 465	0 to 60	
(7) sigma 1-lactate/ plus pyruvate	0 to 465	0 to 60	
(8) sigma d-betahydroxybutyrate plus acetoacetate	0 to 465	0 to 60	
(9) sigma (6+7+8)	0.4 to 465	25 to 60	
Total nonionics	0 to 2400	0 to 300	
(10) carbon dioxide	0 to 25	0 to 5	
(11) osmotically active substances*	0 to 2400	0 to 300	
In Table IV solutions, the component interrelationships are always such that:			
(12) mEq.ratio of HCO ₃ / CO ₂	0.1/1 to 55/0.1	12/1 to 85/1	
(13) mEq.ratio of 1-lactate/ pyruvate	20/1 to 1/1	10/1 to 5/1	
(14) mEq.ratio of d-betahydroxybutyrate/ acetoacetate	6/1 to 0.5/1	3/1 to 1.5/1	
(15) mEq.ratio of Na:CL	1.24 to 1.6	1.26 to 1.6	
(16) Milliosmolarity of Solution	260 to 5000	260 to 540	
(17) pH of solution	5 to 9	7 to 8	

*glucose preferred

[0140] Class II solutions which are particularly suited for electrolyte and fluid therapy are subgenerically characterized in Table V below. As before, each Table V solution comprises water which has dissolved therein the indicated components in the respective amount indicated. In this Table V, the "preferred" class of embodiments (so identified) can be regarded as being representative of compositions which are now believed to be suitable for usage, for example, by hospitals and the like. In making and using all these solutions, care should be taken to preserve the various ratios, osmolarity, and pH values, all as shown in such Table V.

[0141] Such Class II solutions can be used in an *in vivo* process for accomplishing electrolyte and fluid therapy in a mammal. Parenteral nutrition optionally can be concurrently accomplished (depending upon the content of nutrients, such as nonionic osmotically active substances (like glucose, or other conventional additives, including amino acids). As with the process involving Class I solutions, this process:

- 5 (a) tends to maintain the normal plasma milliequivalent ratio of sodium cations to chloride anions, and
- (b) tends to maintain normal plasma and cellular pH ratios, and
- (c) tends to maintain normal cofactor ratios. This process comprises intravascularly introducing into the blood of a mammal a physiologically effective

10 amount of such a solution. The quantity introduced can vary per 24 hour day per patient depending upon the circumstances, patient condition, physicians purpose, and the like. No minimum or maximum definite limit on safe usage quantity is now known or believed to exist.

15 Table V

Generic Composition of Class II Solutions for Electrolyte and Fluid Therapy			
Component	Composition at time of Administration Quantity Range (millimoles per liter)		
	broad	preferred	
Total cations (mEq/L)	1 to 170	136 to 170	
(1) sodium ⁺	1 to 170	130 to 160	
(2) potassium ⁺	0 to 10	3 to 5	
(3) calcium ⁺⁺	0 to 5	1 to 1.5	
(4) magnesium ⁺⁺	0 to 5	0.5 to 1.0	
Total anions	1 to 170	136 to 170	
(5) chloride ⁻	0.6 to 137	81 to 129	
(6) bicarbonate ⁻	0 to 64	0 to 51	
(7) sigma l-lactate ⁻ and pyruvate	0 to 64	0 to 51	
(8) sigma d-betahydroxybutyrate /and acetoacetate	0 to 64	0 to 51	
(9) sigma (6+7+8)	0.4 to 64	25 to 51	
Total nonionics	0 to 625	0 to 305	
(10) carbon dioxide	0 to 25	0 to 5	
(11) osmotically active substances*	0 to 600	0 to 300	
In Table V solutions the component interrelationships are always such that:			
(12) mEq.ratio of HOO ₃ ⁻ / CO ₂	0.1/1 to 55/0.1	0.1/1 to 55/0.1	
(13) mEq.ratio of l-lactate ⁻ / pyruvate	20/1 to 1/1	10/1 to 5/1	
(14) mEq.ratio of d-betahydroxybutyrate ⁻ / acetoacetate	6/1 to 0.5/1	3/1 to 1.5/1	
(15) mEq.ratio of Na:Cl	1.24 to 1.6	1.24 to 1.6	
(16) Milliosmolarity of Solution	260 to 950	260 to 550	
(17) pH of Solution	5 to 9	5 to 9	

*glucose preferred

[0142] Class II solutions which are particularly suited for use in dialysis (whether hemo- or peritoneal) are subgenerically characterized in Table VI below.

Table VI

Class II Solutions Particularly Suited for Dialysis			
	(Hemo- & Peritoneal) Component	Composition at Time of Administration Quantity Range (millinoles per liter)	
		broad	preferred
5	Total cations (mEq/L)	130 to 170	136 to 155
10	(1) sodim ⁺	130 to 155	135 to 145
	(2) potassium ⁺	0 to 5	0 to 4
	(3) calcium ⁺⁺	0 to 3	0 to 1.7
	(4) magnesium ⁺⁺	0 to 2	0.3 to 1
15	Total anions (mEq/L)	130 to 170	136 to 155
	(5) chloride ⁻	81 to 125	86 to 104
	(6) bicarbonate ⁻	0 to 60	25 to 45
	(7) sigma 1-lactate ⁻ /plus pyruvate	0 to 60	2 to 10
20	(8) sigma d-betahydroxybutyrate ⁻ / plus acetacetate	0 to 60	1 to 5
	(9) sum (6+7+8)	25 to 60	27 to 55
	Total nonionics	0 to 525	11 to 280
25	(10) carbon dioxide	0 to 25	0.5 to 2
	(11) osmotically active substance*	0 to 500	10 to 280
30	In Table VI Solutions, the component interrelationships are always such that:		
	(12) mEq.ratio HCO ₃ ⁻ / CO ₂	0.1/1 to 55/0.1	19/1 to 8/1
	(13) mEq.ratio of l-lactate ⁻ / pyruvate	20/1 to 1/1	10/1 to 5/1
	(14) mEq. ratio of Na:Cl	1.24 to 1.6	1.36 to 1.5
	(16) Milliosmolarity of Solution	260 to 850	280 to 320
	(17) pH of Solutions	5 to 9	7.35 to 8

*glucose preferred

[0143] Class II solutions which are within the scope of Table VI above and which are particularly suited for hemodialysis are subgenerically characterized in Table VII below. As before, each Table VII solution comprises water which as dissolved therein the indicated components in the respective amounts indicated.

[0144] Such Class II solutions in Table VII are suitable for use in a hemodialysis process of the generally known and conventional type where renal function of a living mammal is replaced in whole or in part by dialysis. In hemodialysis, portions of the blood of such mammal are continuously passed over one face of a dialysis membrane (which is incorporated preferably a high surface area cartridge-like structure) while the opposed face of such membrane is contacted with a dialysis fluid, thereby to achieve a change in the chemical composition of the body fluids after the so dialyzed blood is returned to the mammal's vascular system. Duration of a conventional hemodialysis can vary, depending upon equipment, conditions, patient condition, and the like, but typically can extend for a time of from about 3 to 5 hours. Optionally, but preferably, the dialysis membrane used in combination with the associated apparatus is such that the blood so passed over such membrane can be pressurized during such passage (typically and conventionally up to about 300 grams per cubic centimeter), thereby to produce what is known in the dialysis art as "ultrafiltration". In the conventional hemodialysis procedure, the dialysis fluid is an aqueous solution which contains dissolved therein the same principal inorganic electrolytes at respective individual concentration levels which approximate such major plasma electrolytes and their concentrations.

[0145] In the patent hemodialysis one substitutes for the conventional dialysis fluid a solution of the present invention and as below characterized in Table VII. Conventional dialysis equipment can be used, but a deaerator, such as might tend to eliminate dissolved carbon dioxide from a dialysis solution of this invention, should not be present. During use in peritoneal dialysis, a solution of this invention:

- (a) tends to maintain a normal equivalent ratio of sodium cations to chloride anions, and

- (b) tends to maintain normal cellular and plasma pH, and
 (c) tends to maintain normal cofactor ratios.

The total quantity of such solution of this invention used in a given hemodialysis is comparable to the quantities used in prior art fluids employed under the same conditions (typically from about 35 to 160 liters of dialysis fluid per hemodialysis per man).

Table VII

Class II Solutions Particularly Suited for Hemodialysis			
Component	Composition at Time of Administration Quantity Range (millimoles per liter)		
	broad	preferred	
Total cations (mEq/L)	130 to 170	134 to 154	
(1) sodium ⁺	130 to 155	132 to 145	
(2) potassium ⁺	0 to 5	0 to 4	
(3) calcium ⁺⁺	0 to 3	1 to 1.75	
(4) magnesium ⁺⁺	0 to 2	0.3 to 0.75	
Total anions (mEq/L)	130 to 170	134 to 154	
(5) chloride ⁻	84 to 125	93 to 115	
(6) bicarbonate ⁻	0 to 55	25 to 35	
(7) sigma l-lactate ⁻ / pyruvate	0 to 55	0 to 12	
(8) sigma d- betahydroxybutyrate/ acetoacetate	0 to 55	0 to 5	
(9) sigma (6+7+8)	25 to 55	36 to 42	
Total nonionics*	0 to 525*	0 to 12	
(10) carbon dioxide	0 to 25	0 to 2	
(11) osmotically active substances**	0 to 500*	0 to 10	
In Table VII, the component interrelationships are always such that:			
(12) mEq-ratio of bicarbonate ⁻ / CO ₂	0.1/1 to 55/0.1	18/1 to 35/0.5	
(13) inEq.ratio of l-lactate ⁻ / pyruvate ⁻	20/1 to 1/1	10/1 to 5/1	
(14) mEq.ratio of d- betahydroxybutyrate/ acetoacetate	6/1 to 0.5/1	3/1 to 1.5/1	
(15) mEq.ratio of Na:Cl	1.24 to 1.6	1.26 to 1.55	
(16) milliosmolarity of Solution	260 to 800	260 to 350	
(17) pH of Solution	5 to 9	7.35 to 8	

*This upper limit used when the solution is being employed in an old type Kolff kidney where pressure cannot be exerted on the dialysis membrane.
 In a pressurized dialysis system the limit is 0 to 11 mmol/l for glucose; if other nonionics are added, then preferred limit Mould be below 20 mmol/l total.

**glucose preferred.

[0146] Class II solutions which are within this scope of Table VI above and which are particularly suited for peritoneal dialysis are subgenerically characterized in Table VIII below.

[0147] Such Class II solutions of Table VIII are suitable for use in a peritoneal dialysis process of the generally known and conventional type when renal function of a living mammal is replaced in whole or in part by dialysis. In peritoneal dialysis a quantity of a dialysis fluid is charged into the peritoneal cavity of such mammal for a time sufficient to achieve a change in the chemical composition of body fluids, after which the dialysate is drained or otherwise removed from the peritoneal cavity. Typical residence times for fluid in the peritoneal cavity range from about ½ to 1 hour, although longer and shorter times can be employed. Typically, peritoneal dialysis sessions last 4-½ hours, but continuous ambulatory peritoneal dialysis has recently been advocated. The patient's own peritoneum serves as a dialysis membrane.

In the conventional peritoneal dialysis procedure, the dialysis fluid is, as in the case of a hemodialysis fluid and aqueous solution which contains dissolved therein the same principal inorganic electrolytes and at respective individual concentration levels which approximate those of major plasma electrolytes and their concentrations, except that in the case of peritoneal dialysis fluids a higher concentration of nonionics, such as glucose, is typically employed in order to provide as osmolarity which is greater than that of mammalian plasma, thereby to promote ion and water transfer through the peritoneum, all as known to those skilled in the art. Chronic, so called "ambulatory" peritoneal dialysis may also benefit from these solutions.

[0148] One may substitute for the conventional dialysis fluid a solution of the present invention and as below characterized in Table VIII. During use in peritoneal dialysis, a solution of this invention:

- (a) tends to maintain a normal equivalent ratio of sodium cations to chloride anions,
- (b) tends to maintain normal plasma and cellular pH,
- (c) tends to maintain normal cofactor ratios.

[0149] The quantity of such solution employed is comparable to the quantity used in prior art peritoneal dialysis as is the residence time in the peritoneal cavity.

Table VIII

Class II Solutions Particularly Suited for Peritoneal Dialysis			
	component	Compositions at Time of Administration Quantity Range (millimoles per liter)	
		broad	preferred
	Total cations	130 to 170	135 to 150
(1)	sodium ⁺	130 to 165	130 to 145
(2)	potassium ⁺	0 to 5	0 to 4
(3)	calcium	0 to 2	1 to 1.5
(4)	magnesium ⁺⁺	0 to 1.5	0.3 to 1
	Total anions	130 to 170	135 to 150
(5)	chloride ⁻	81 to 130	93 to 102
(6)	bicarbonate ⁻	0 to 55	25 to 30
(7)	sigma l-Lactate ⁻ /plus pyruvate ⁻	0 to 55	2 to 12
(8)	sigma l-betahydroxybutyrate ⁻ /acetoacetate ⁻	0 to 55	1 to 5
(9)	sigma (6+7+8)	26 to 55	36 to 50
	Total nonionics*	40 to 252	84 to 238
(10)	carbon dioxide	0 to 25	0 to 2
(11)	osmotically active substance	40 to 250	83 to 237
In Table VIII, the component interrelationships are always such that:			
(12)	mEq.ratio of HCO ₃ ⁻ / CO ₂	0.1/1 to 160/1	19/1 to 21/1
(13)	mEq.ratio of l-lactate ⁻ / pyruvate ⁻	20/1 to 1/1	10/1 to 5/1
(14)	mEq. ratio of d-betahydroxybutyrate ⁻ /acetoacetate ⁻	6/1 to 0.5/1	3/1 to 1.5/1
(15)	mEq.ratio of Na:Cl	1.24 to 1.6	1.36 - 1.42
(16)	Milliosmolarity of Solution	310 to 615	350 to 520
(17)	pH of solution	to 8	7.36 to 7.6

* glucose preferred

EMBODIMENTS

[0150] The following examples are merely illustrative of the present invention and are not intended as a limitation upon the scope thereof.

Examples 1 through 27

[0151] The following compositions of this invention and comparative examples illustrate electrolyte solutions of Class I (above identified) which are suitable for intravenous administration to replace electrolytes and fluid in a human adult patient at dose rates of, for example, 500ml/patient/24 hour day. Each solution consists of water which has dissolved therein each of the identified in the respective specific per liter quantity shown components in the following Table IX.

[0152] Each solution is here prepared by dissolving substantially pure selected salt and nonionic material following the teaching of "Date for Biochemical Research", 1969, pp.507-508. Each solution can be made from many different materials depending upon manufacturing convenience, ease of sterilization, cost of raw materials, and the like; the only requirement is that the final ionic composition of each solution should be as described.

[0153] The footnote for each example in Table IX characterizes the composition and provides a suggested application or use.

[0154] Also shown in Table IX are further examples of prior solutions. All solutions are listed as Type 1 a, b, c, and d, in conformity with the classification herein developed.

Table IX Class Ia Solutions Containing 1 or 2 Cations from Among Na^+ , K^+ , Mg^{2+} or Ca^{2+} with no Nutrients (Glucose)

Units	Marcel Plaza Ref.J.A. 285, 1285 L. fluid	1.1 0.91 U.S. 1970	1.02 0.952 U.K.	1.03 NaCl U.S.	1.04 Isotonic Malactate Salt	1.05 Isotonic Malact/Pyr- BHB/acat	1.06 Isotonic Malact/Pyr- BHB/acat	1.07 Isotonic Malact/Pyr- BHB/acat	1.08 Isotonic Malact/Pyr- BHB/acat	1.09 Isotonic Malact/Pyr- BHB/acat
Na	136 - 145	155	162.5	160.3	157	155	155	152.5	152.5	152.5
K	3.5 - 5.0					2.5				
Ca free [Ca2+] [1.06]	2.1 - 2.6					1				
Mg free [Mg^{2+}] [0.55]	0.75 - 1.25									
2 Eq Cations 102.7-153.2 155			162.5	160.3	155	155	155	155	155	155
C1	160 - 166	155	162.5	160.3	166	166	166	166	166	166
HCO_3^-	26 - 29									
Cl^-	1 - 1.45									
SO_4^{2-}	0.22 - 0.94									
L-lactate	0.6 - 1.8									
pyruvate										
						5	5	5	5	5

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Lact/pyr	6.0	6.8	9.5	
0.0 Ethbutyrate			4.7	35
acetocetate			2.3	14
0.0 KBr / acac			2.0	2.5
acetate				
Other				
Z Afq inions 128.7-139.4	155	162.5	160.3	155
Na/Cl	1.28 - 1.45	1.60	1.00	1.48
Glucose	3.9 - 5.6			
or others				
CO ₂	0.99 - 1.39			
pH	7.35 - 7.45	5.5 - 6.5	4.5 - 6.5	6.5
Z solns	265 - 295	310	325	321
Use:	I.V. elec- trolyte replacement	same as I.V. electrolyte	Used to prevent acidosis	Improves IAT, IAT control of IAT cytoplas & with K with Ca ²⁺ mitochondria

1.a.1. Most common electrolyte solution given in U.S. tends to cause hyperchloric acidosis because of abnormal Na/Cl ratio.
 See Black Bk. Lancet i, 153, 1952.

1.a.2. Used in U.K. and Canada.

1.a.3. Barre et al. J Am Med Ass 143, 365, 432, 1944. Causes renal imbalance.

1.a.4. : - Solutions so about are now in this disclosure.

* — comparative example

Table II Class Ib Solutions Containing 1 or 2 Cations from Among Na^+ , K^+ , Mg^{2+} , Ca^{2+} with HCO_3^- or $\text{HCO}_3^-/\text{CO}_2$ and No Nutrients.

Units	Mg/mol	4		5		6		7	
		1 b 1	1 b 2	1 b 3	1 b 4	1 b 5	1 b 6	1 b 7	1 b 8
Plasma	Isotonic								
B.I.J.M.	NaHCO_3^-								
—	NaHCO_3^-								
1 fluid	203, 1285	Salt	$\text{NaHCO}_3^-/\text{CO}_2 + \text{NaAc}/\text{Prv}$	$\text{NaCl}/\text{K}/\text{A}$	$\text{NaCl} + \text{Ca}$	$\text{NaHCO}_3/\text{CO}_2$			
	1970								
Na	136 - 145	160.3		155		155		152	
K	3.5 - 5.0							3	
Ca free (Ca^{2+})	2.1 - 2.6 (1.06)								
Mg free (Mg^{2+})	0.75 - 1.25 (0.53)								
2 atq. Cations 142.7-153.2 160.3									
Cl	160 - 166	168.3	106	106	106	106	106	106	106
HCO_3^-	26 - 28	52	27	27	27	27	27	27	27
Zn	1 - 1.45								
SO_4^-	0.32 - 0.94								
L-lactate	0.6 - 1.8			19	19	13	13	13	13

	1	3	2	2
pyruvate				
Lact/pyr	6.3	6.3	6.5	6.5
D & DLbutyrate			5	4
acetoacetate			2.5	3
D HMP acac			2.5	1.3
acetate				
Other				
F Eq anions 128.7-139.4 160.3	155	155	155	155
Na/Cl	1.28 - 1.45 1.48	1.46	1.44	1.46
Glucose	1.9 - 5.6			
or others	0.99 - 1.39 *	1.3	1.3	1.3
CO ₂				
pH	7.35 - 7.75 8.6	7.35	7.35	7.35
ZnO ₁₀	285 - 295 321	311	311	311
Uses:				

16.1 Barone et al. J Am Med Res 103: 365, 1944, abnormal pH. Incompatible with Mg²⁺ and Ca²⁺.

* — comparative example

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Table IX. Class II Solutions Containing 1 or 2 Cations from Among Na^+ , K^+ , Mg^{2+} or Ca^{2+} with Non-ionic Nutrients.

Common non-ionic nutrients are 5%, 2.5%, 10% glucose. Additional similar fructose and glycerol solutions in over 20% amounts are prepared by commercial companies.

accounts are approved by FRS, but not recommended here. (See "Safe Entry Points")

c 1 - Most common I.V. fluid given. *Werth Haesbrouck 1956, p1067.* This is combined with isotonic NaCl in many proportions.
 c 2 - Used in the U.K. and Canada where "tonic" is different than in the U.S. - presumably. *Squier's Handbook 1974, p. 37*

6.3 - 2 parts isotonic glucose plus 1 part isotonic NaCl - Griggs *Nursing* 1970, p 334.
6.4 - Prevents hypochlorhydria but causes edema. *Primer of Internal Medicine*, 1970, p 331.

¹ See also his article 'Bult causes reborn' (ibidem).

Table II. Class Ic (Cont'd)
Case 1

	11	12	13*	14	15
Units	Normal 1 c 8	1 c 9	1 c 11	1 c 12	1 c 15
Plasma	2L 0.9% +	1L 0.9% +	1L Glucose 0.5% +	2.5% Glucose D 5 W +	D 10 W +
anoles	0.5L J.M. 283, 1285	0.5L Normal Saline +	0.9% NaCl + 0.9% NaCl + 0.15% NaCl	BuB/Heat + 1/F Saline	Redox
.....	283, 1285	Saline +			
L fluid	1970	Redox			
		Balance			
H ₂ O	136 - 105	31	31	154	154
K	3.5 - 5.0		5.0		
Ca	2.1 - 2.6				
free [Ca ²⁺] (1.06)					
Mg	0.75 - 1.25				
free [Mg ²⁺] (0.53)					
Σ anions 142.7-153.2	31	36	154	154	154
Cl	100 - 106	22	22	154	105
HCO ₃	26 - 28				
2 Pi	1 - 1.45				
SO ₄	0.32 - 0.94				
l - lactate	0.6 - 1.8	7	10	..	43
pyruvate		1	1.43		39

Lact/pyr	7	7	7.2	7
D & Dihydrate	0.66	1.57		4
acetoacetate	0.33	1.00		3
D Hb/ acac	2	1.6	1.5	1.3
acetate				
Other				
Σ eq anions 128.7-139.4	31	36	154	154
Na/Cl	1.28 - 1.45	1.41	1.00	1.00
Glucose	3.9 - 5.6	222.4	278	556
or others			139	278
CO ₂	0.99 - 1.39			
pH	7.35 - 7.45	*6.5	*5.5-6.5	*5.5-6.5
Σ a₀₂	285 - 295	284	561	293
Use:				
			Improves	Improves
			I c 10 with I c 11	I c 11
			Redox & Na/Cl	I c 13
			Balance	

I c 8. Improves with normal Na/Cl ratio and redox balance the most common routine I.V. order in the U.S.

I c 9. Replaces 12.5 eq of the 40 eq of K⁺ lost/ day when given at the usual rate of 2.5L/ day.

I c 10. Facts and Comparisons Oct '81, p.51,ippincott, St Louis

I c 11. Facts and Comparisons Oct '81, p.51,ippincott, St Louis

I c 12. Facts and Comparisons Oct '81, p.51,ippincott, St Louis

* — comparative example

Table II. Classical Solutions Containing 1 or 2 Cations from Among Na^+ , K^+ , Mg^{2+} , or Ca^{2+} plus Non-Ionic Nutritive Plus $\text{HCO}_3^-/\text{CO}_2$

L-lactate 0.6 - 1.0

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10		1	1	
15		2	2	
20				
25				
30				
35				
40				
45				
50				
55				
	D & G butyrate			
	acetocetate			
	9 Hg/ acac			
	acetate			
	Other			
	Z eff anions 128.7-139.4	155	160	156
	NaCl	1.28 - 1.45	1.45	1.45
	Glucose	3.9 - 5.6	10	10
	or others	(optional)	10	10
	CO ₂	0.99 - 1.39	2.7	2.71
	pH	7.35 - 7.45	7.35	7.35
	Z effs	285 - 295	320	330
	Use:			
		Improves	Replaces	Mg ²⁺
		coral NaCl	K loss	Ca ²⁺ does
		leaves pale		not ppt.
		agent alkalotic		precipitate as with HCO ₃ ⁻
				slight.

* — comparative example

Table II Class I_d Solutions Containing 1 or 2 Cations, to which is added HCO₃⁻/CO₂ and Non-ionic Nutrients
Case I (Cont'd)

	24	25	26	27
Units	Noral 2L D5W	1 d 9 2L D5W	1 d 10 R.B. Saline + 0.5L R.R.	1 d 11 R.B. Saline Like 1d11 with K & but RHB acid
anoles	M.F.J.M. 283, 1285	Saline	Saline + F	2.5% Gluc. & added to make No added CO ₂ CO ₂ in situ
L fluid	1970			
Na	136 - 145	28.2	28.2	140
K	3.5 - 5.0	5	4	4
Ca	2.1 - 2.6 free [Ca2+] [1.06]			
Mg	0.75 - 1.25 free [Mg2+] [0.53]			
Σ Eq Cations	142.7-153.2	28.2	33.2	144
Cl	100 - 106	20	20	104
HCO₃	26 - 28	5.8	10.8	29
Σ Pi	1 - 1.5			
SO₄	0.32 - 0.94			
L-lactate	0.6 - 1.8	1.4	1.4	5
pyruvate	0.2	0.2	1	1

5	Lact/pyr	7	7	7	7	
10	D Dibutyrate	0.4	0.4	2	2	
15	acetocetate	0.2	0.2	1	1	
20	D Hb / acac	2	2	2	2	
25	acetate					• 2 Mlactate + 2 d D hydroxybutyric acid
30	Other					
35	Σ Eq anions 120.7-139.4	28.2	33.2	144	144	
40	Na/Cl	1.20 - 1.45	1.41	1.35	1.35	
45	Glucose or others	3.9 - 5.6	222.4	139	139	
50	CO ₂	0.99 - 1.39	0.29	0.54	0 -	
55	pH	7.35 - 7.45	7.4	7.4	7.4	
60	Σ salts	265 - 295	279	289	277	
65	Use:			Replaces 2) D5W & K less 0.5) Normal Saline	Replaces 2) Replaces	

I 4 11 • L Lactic acid is added instead of CO₂ to generate CO₂ in situ.

I 4 12 • D D Hydroxybutyric acid is added to generate [L]₂ in situ.

Examples 28 through 41

[0155] The following compositions of this invention illustrate electrolyte solutions of Class II (above identified) which are suitable for (a) intravenous use to replace electrolytes and fluid (b) providing parenteral nutrition in a human adult patient, (c) peritoneal dialysis, and (d) hemodialysis. Dose rates can vary. Each solution consists of water which has dissolved therein each of the identified components in the respective specified concentrations per liter quantity shown in the following Table X. Each solution is prepared by conventional procedures. (See text of Examples 1 through 27).

[0156] The footnote for each example in Table X characterizes the composition and provides a suggested application or use.

[0157] These compositions demonstrate, as do Tables V through VIII (above), that there is no essential compositional difference between these various solutions.

[0158] Table XI shows prior art hemodialysis fluids for comparison purposes in dialyzing a human adult patient using, for example, an apparatus as described by Miller J.H., Schinaberger J.H., Kraut J.A., and Gardner P.S., Trans. Am. Soc. Artif. Intern. Organs 25, 404-408, 1979.

[0159] In these solutions which contain dissolved CO₂, no aerator should be used on the dialysis equipment.

Table I Class 2a Electrolyte Fluids Containing 3 or 4 Cations Suitable for Contracting Cells, Containing No $\text{HCO}_3^-/\text{CO}_2$ and No Glucose; e.g. after S.J. Ringer, *Physiol Rev* 4: 20, 223, 1885, and 7: 291, 1886.

Units	Na	K	Ca^{2+} free [Ca^{2+}] { 1.05 }	Mg^{2+} free [Mg^{2+}] { 0.53 }	2B*		2B		2B			
					Ringer's Injection 283, 1985 L. fluid 1970	Ringer's U.S. (Commercial)	Lactated Ringer's	Acetated Ringer's	Lact/Fyr Ringer's	Lact/Fyr Ringer's	dB-HB/acet Ringer's	dB-HB/acet Ringer's
Mm	136 - 145	147	129.8	130	130	140	130	130	130	130	130	130
K	3.5 - 5.0	4	5.4	4	4	4	10	4	4	4	4	4
Ca^{2+}	2.1 - 2.6	2.5	0.9	1.5	1.5	2.5	1.5	1.5	1.5	1.5	1.5	1.5
Mg^{2+}	0.75 - 1.25		1.0									
Zn^{2+}	Cations	142.7-153.2	156	139	137	137	158	137	137	137	137	137
C1	100 - 106	156	111.8	109	109	103	96	96	96	96	96	96
HCO_3^-	26 - 28											
ZPi		1 - 1.45										
SD ₄		0.32 - 0.94										
L-lactate	0.6 - 1.8		27.2 (d,l)	28 (d,l)		27.5 (d,l)	35.9		30			
pyruvate						5.1						

Lact/pyr	00	00	00	00	7	7.5
D & Oribulurate					27.1	5
acetooacetate					13.7	2
D Hb/ acat					2	2.5
acetate	28	27.5				
Other						
Σ afg anions	128.7-139.4	156	139	137	137	137
Na/Cl	1.29 - 1.45	0.91	1.16	1.19	1.36	1.35
Glucose	0.11.215-1.451					
or others	3.9 - 5.6					
CO ₂	0.99 - 1.39					
pH	7.35 - 7.45					
Σ eObs	285 - 295	309	276	272	312	272.5
User:	I.V. fluid	I.V. fluid	I.V. fluid	I.V. fluid	I.V. fluid	I.V. fluid
	2 a 3.	2 a 4	2 a 3.	2 a 4	2 a 3.	2 a 4

J. L. M. Pelt & Blood (1991) Guide, Revised Nov 1, '82.

2: In the Fields and Furnaces 850-900 (Innsbruck)

2.1.3. *Neurokinin-3 Receptor*

2. I. Hartmann *Br. J. Nutr.* 1955; 13(4): 183-191.

J. J. Ettinger and Co., 101, Lippincott.

2. 1. 1. Factors and Comparisons p50, Det '01, Lippincott

2, 3, 5, for et al., J. Ab. Med. Ass. 148: 827, 1952.

* = comparative example

COMPARATIVE EXAMPLE

Table I. Class 2a Electrolyte Fluids Containing 3 or 4 Cations Suitable for Contacting Cells, Containing No $\text{HCO}_3^-/\text{CO}_2$ and No Glucose; e.g. after S.J. Ringer, *Physiol. A*; 29, 223, 1881, and 7; 291, 1886.

Units	Normal Plasma	2. 4. 1.	2. 4. 2.	2. 4. 3.	2. 4. 4.	2. 4. 5.	2B*	2B	2B	2. 4. 7.	2. 4. 8.
		Ringer's M.S.J.B. Injection 283, 1285 U.S.	Lactated Ringer's U.S.	Acetated Ringer's (Commercial) U.S.	Ringer's Ringer's	Lact/acet Ringer's		Lact/Pyr Ringer's	Ringer's	dB-HB/acac Redox	Balanced Ringer's
H ₂ O	136 - 145	107	129.8	130	130	140	130	130	130	130	130
K	3.5 - 5.0	4	5.4	4	4	10	4	4	4	4	4
Ca	2.1 - 2.6	2.5	0.9	1.5	1.5	2.5	1.5	1.5	1.5	1.5	1.5
Mg free [Mg ²⁺] { 0.53 }	0.75 - 1.25	1.0				1.5					
Z Eq Cations 142.7-153.2	156	139	137	137	137	158	137	137	137	137	137
Cl	100 - 106	156	111.8	109	109	103	96	96	96	96	96
HCO_3^-	26 - 28										
ΣPi		1 - 1.45									
SO ₄	0.32 - 0.94										
L - lactate	0.6 - 1.6		27.2 (d,1)	28 (d,1)		27.5 (d,1)	35.9				
pyruvate							5.1				
Lact/Pyr		00	00			00	00	00	00	00	00
						7					7.5

N.I.H. Path & Blood Bank Guide, Revised May 1, '82.

7. 3. 1. *Facts and Comparisons* p50, Oct '01, Lippincott

Hartmann Af. J. Ab. Ned. Ass. 103: 1349, 1934.

2. *3. Facts and Comparisons* p50, Oct '81, Lippincott.

Facts and Comparisons 030 • Oct '91 • Lippincott

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* – comparative example

Table I. Class 2a (Cont'd) Solutions with **Bold numbers** and in boxes are new disclosures.

		31	2 a 9	2 a 10	2 a 11	2 a 12	2 a 13	2 a 14	2 a 15
Units	Normal	2 a 9	2 a 10	2 a 11	2 a 12	2 a 13	2 a 14	2 a 15	
Plasma	Redox	Ionalol	Plasalyte	Isolyte S	Isolyte E	DeBacco's	Krebs		
anoles	N.F.J.A.	Balanced	D-CM	(Travenol)	(McGaw)	Pi Buffered	Ringer		
	283, 1285	Ringer's &	Abbott	PolvenicR148	(Cutter)	Saline	Phosphate		
L fluid	1970	High K							
Na	136 - 145	140	139	140	140	140	140	152	150.76
K	3.5 - 5.0	10	12	10	5	10	4.17	5.92	
Ca	2.1 - 2.6	1.0	2.5	2.5		2.5	0.9	2.54	
free [Ca²⁺] [1.06]									
Mg	0.75 - 1.25	0.5	1.5	1.5		1.5	0.45	1.18	
free [Mg²⁺] [0.53]									
2 aEq Cations	142.7-153.2	153	158	148	158	159.15	164.12		
C1	100 - 105	103	108	103	98	103	140.5	131.51	
HCO₃		26 - 28							
2 Pi		1 - 1.45					9.8	17.30	
SO₄		0.32 - 0.94					0.45	1.10	
L - lactate	0.6 - 1.8	30	50 (d,1)	0 (d,1)					
pyruvate		5							
Lact/pyr		7.6					00	00	

D 0 Dihbutyrate	5							
acetoacetate	2							
BHB / acat	2.5							
acetate		47	27	49				
Other					23 gluconate 4 citrate			
Σ aEq anions 120.7-139.4	153	159	168	158		159.18	163.97	
Na/Cl	1.28 - 1.45	1.36	1.28	1.36	1.43	1.40	1.08	1.15
Glucose	3.9 - 5.6							
or others								
CO ₂	0.99 - 1.39							
pH	7.35 - 7.45							
Z nEq	265 - 295	304.5	312	312	294	315	308.3	311.65
Use:	Improves I.V. electrolyte 2 a 5, lowers Ca & therapy Mg to normal		Same as 2 a 10	Same as 2 a 10	Tissue culture pPi accou- ration	Tissue culture 2 a 10	Balance of salt air	Biochemical experiments

2410. *facts and convenient Ref.* p. 50

241 *Facts and Comparisons* Oct '81

2012, Vol. 12, No. 1, pp. 3–30

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63 13. WEBBS MM. 1877-1881 7 MARCH 1881

Table I. Class 2b Electrolyte Fluids Containing 3 to 4 Cations Suitable for Contracting Cells Also Containing $\text{HCO}_3^-/\text{CO}_2$ and No Glucose after Krebs H.A. & Hennefelt H.A., *Appar-Septier's J Physiol Chem* 210: 33-56, 1932.

	32	33	34 *	35 *	36
Units	Normal	2 b 1	2 b 2	2 b 3	2 b 4
Phases	Krebs	Krebs	Ringer-Bal-	High HCO_3^-	2 b 5
anoles	H.F.J.M.	Henseleit	anced	L/P Ringer's	2 b 6
.....	281, 1985			Ringer's 1/2	
L fluid	1970.			Lactate	
				Ketones	
				$\text{HCO}_3^-/\text{CO}_2$	
Na	134 - 145	141	130	136	130
K	3.5 - 5.0	5.9	4	4	4
Ca	2.4 - 2.6	2.5	1.5	1	1.5
Mg free (Mg^{2+})	{ 0.06 }				1.5
Mg free (Mg^{2+})	0.75 - 1.25	1.2	0.5	0.5	
3 Eq Cations 142.7-153.2 156.3					
Cl	100 - 106	127.8	96	100	96
HCO_3^-	26 - 28	25	29	43	29
Z Pi	1 - 1.45	1.18			
SO_4^-	0.32 { 0.94 }	1.2			
t-lactate	0.6 - 1.9		7	9	10.5
pyruvate			1	1	1.5

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Lact/pyr	7	9	7	
D D Bicarbonate	3	3		8
acetooacetate	1	1		4
B Hb/ acac	1	3		2
acetate				
Other				
Z Eq anions 128.7-138.4	137.3	143	143	137
Na/Cl	1.28 - 1.45	1.12	1.35	1.36
Glucose	3.9 - 5.6			
or others	0.99 - 1.39	1.24	1.5	1.5
CO ₂			2.46	1.5
pH	7.35 - 7.45	7.4	7.4	7.4
Z osm	285 - 295	308	274	286
Use:	Tissue	To replace incubation, all previous organ perfusion	For Rx of acidosis lactated Ringer's	Alternate to 2 b 2 to 2 b 5

2 b 2 to 2 b 6. All these solutions would be suitable, given added glucose, for peritoneal dialysis, ie like class 2 c.
As it is, these solutions would improve existing hemodialysis.

* — comparative example

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Table 1. Class 2c Electrolyte Fluids Containing 3 or 4 Cations Suitable for Contacting Cells Containing $\text{NaHCO}_3/\text{CO}_2$ to which are Added Non-ionic nutrients such as Glucose, Fructose, Glycerol etc.

Units	Normal	2 c 1	2 c 2	2 c 3	2 c 4	2 c 5	2 c 6	2 c 7
Plasma	Lactated	1/2 Strength Acetated	Ionsol B	Dianal	Peritoneal	Dianal	Peritoneal	Dianal
N.E.J.M.	Ringer's 5% Lactated	Ringers 5% b 51	b 51	b 1.51	Dialysis	K-141 b	Dialysis	K-141 b
283, 1285	Glucose	Ringer's 2.5% 5% Glucose	Glucose	4.25% Gluc	Glucose	4.25% Gluc	Glucose	4.25% Gluc
1. Fluid	1970		(Abbott)	(Travenol)	(Abbott)	(Abbott)	(Abbott)	(Abbott)
Na	136 - 145	130	65	130	57	141	141.5	132
K	3.5 - 5.0	4	2	4	25		4	
Ca	2.1 - 2.4	1.5	0.75	1.5	1.75	2.0	1.875	
free (Ca ²⁺)	{ 1.06 }							
Mg	0.75 - 1.25				2.5	0.75	0.75	0.75
free (Mg ²⁺)	{ 0.53 }							
2 Eq Cations	142.7-153.2	137	40.5	137	87	146	147	141
C1	100 - 106	109	55	109	49	101	102.5	106
HCO ₃	26 - 28							
2 Pi	1 - 1.45				6.5 H ₂ PO ₄ ⁻			
SO ₄	0.32 - 0.94							
1 - Lactate	0.6 - 1.0	28 (d,1)	14 (d,1)	25 (d,1)	45 (d,1)	35 (d,1)		
Pyruvate								

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Lact/pyr	00	00	00	00	00	00
0 0 Dihydrate						
acetooacetate						
B HB/ acac						
acetate	28			44.5		
Other						
Z Eq anions 128.7-139.4	137	69	137	87	146	147
Na/Cl	1.28 - 1.45	1.19	1.18	1.19	1.16	1.40
Glucose	3.7 - 5.6	278	139	278	83	236
or others						
CO ₂	0.99 - 1.39					
pH	7.35 - 7.45				*5.5-6.5	*5.5-6.5
Z adsa	285 - 295	5247 *(550.5)	263	523	443	366
Use:						
	I.V. therapy I.V. therapy I.V. therapy Parenteral				Peritoneal	Peritoneal
	for dehydr- seen as			nutrition	dialysis	dialysis
	ation	2 c 1	2 c 1			

* 2 c 1. Facts and Comparisons Oct '81, p 52. The osmolarity listed by the reference appears to be incorrect at 524 adsm.

The correct osmolarity appears to be 550.5 adsm.

2 c 2 - 2 c 3. Facts and Comparisons Oct '81, p52. Lippincott, St Louis

2 c 4. Facts and Comparisons Oct '81, p52. Lippincott, St Louis

2 c 5 - 2 c 7. Facts and Comparisons Oct '82, p704. Lippincott, St Louis

Table I. CLASS 2c (CONT'D.)

* — comparative example

Table I. Class 2d Electrolyte Fluids for Contacting Living Cells Containing 3 to 4 Cations plus Non-Ionic Nutrients plus HCO_3/CO_2

		39	40	41
Units	Normal Plasma M.F.J.M. 2B3, 12B5 L. fluid	2 d 1 Krebs Serum Substitute 1970	2 d 2 Tyrode's Solution Balanced Salt Solution	2 d 3 Veech's R.B.-Salt sine Pi cws 5% Glucose
Na	136 - 145	141	151.54	142
K	3.5 - 5.0	5.93	5.9	4.5
Ca	2.1 - 2.6	2.54	1.8	1.1
(free Ca^{2+})	[1.06]		[1.06]	[1.1]
Mg	0.75 - 1.25	1.18	0.45	0.56
(free Mg^{2+})	[0.53]		[0.53]	[0.56]
NaEq Cations	142.7-153.2	154.37	162.07	149.82
Cl	100 - 106	104.8	147.8	102
HCO_3	26 - 28	24.9	11.9	29
Z Pi	1 - 1.45	1.23	1.226	1.16 [0.7]
SO_4	0.32 - 0.94	2.36		
L-lactate	0.6 - 1.8		1.33	10.7
pyruvate	4.9	0.09	1.5	1.5

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Lact/pyr	14.8	7	7	7
D B Dihbutyrate		3	3	3
acetoacetate	2	2	2	2
B HB/ acac	1.5	1.5	1.5	1.5
acetate				
Other	2.45 glutamate 5.4 fumarate			
Σ eq anions 128.7-139.4	151.47	162.81	149.82	148.3
Na/C	1.28 - 1.45	1.35	1.03	1.39
Glucose	3.9 - 5.6	9.2	5.65	10
or others				
CO ₂	0.99 - 1.39	1.0	1.17	1.45
pH	7.35 - 7.45	7.4	7.1	7.40
Σ n/a	285 - 295	308.2	320	308.6
Use:	Media for tissue slices	For liver perfusion	for I.V. or general use neal dial. to replace I.V.	for I.V. & peritoneal dialysis 2 b 1 t 2 d 1

2 d 1. Krebs HA. Biochen Biophys Acta 4: 249-269, 1950

2 d 2. Tyrode MJ. Arch int Pharmacodyn 20: 205, 1910. ♦ For use in liver perfusion with albumin see

Schlaesek H. Bioches Z 336:460, 1963

2 d 3. ♦ The apparent charge on rna Pi in the presence of these cations is about 1.46 not 1.0 presumably due to cation binding.

Table II. Prior Art Hemodialysis Fluids. For recent discussion see Parsons FM, Stewart WK, Composition of Dialysis Fluid. In: *Replacement of Renal Function by Dialysis* (Drucker W, Parsons FM, Maher JP, eds.) Martinus Nijhoff, Hingham, pp 168-170, 1983.

Units	Normal Plasma M.F.J.T. 283, 1205 L fluid	2 d 6 Kolff 1947	2 d 7 Brigham 1932	2 a 16 Scribner's Acetate 1964	2 a 17 Commercial Acetate 1981	2 a 18 Bjälder "Low" Acet. 1981	2 a 19 Bjälder "High" Acet. 1981	2 b 2 Kraut HCO ₃ -Acetic Acid, 1981	2 b 3 CORE
K _a	136 - 145	126	140	135	140	134	136	140	135
K	3.5 - 5.0	5.6	4	1.5	2	2.2	2.2	2	2
Ca free [Ca ²⁺] { 1.06 J}	2.1 - 2.6	1.0	1.25	1.25	0.875	1.86	1.91	1.75	1.5
Hg free [Hg ²⁺] { 0.53 }	0.75 - 1.25	0.5	0.5	0.375	0	0	0	-	0.375
Eq Cations 142.7-153.2	133.6	147.5	140	144.5	139.88	147.02	145.5	140.75	
Cl	100 - 106	109	126.7	105	106	107.28	101.82	107	106.5
HCO ₃ _J	26 - 28	23.9	26.6				33	33	
P _i	1 - 1.45								
SO ₄	0.32 - 0.94								
L - lactate	0.6 - 1.0								
pyruvate									

	Lact/pyr	D B OHbutyrate	acetoacetate	B Hb/ acac	acetate	Other	35	38.5	32.6	JB.2	2 Hacetate	2 Hacetone
											23.5 gluconate	
Na/Cl	1.28 - 1.45	1.16	1.16		1.29	1.32		1.25	1.31		1.31	1.27
Glucose	3.9 - 5.6	76 - 151	10		0	0		0	0		0	0
or others	0.99 - 1.39	0			1.24	0		0	0		1.3	1.3
CO ₂												
pH	7.35 - 7.45	8.6	7.4		75.5-6.5	75.5-6.5		76.7	76.7		77.4	77.4
atm	285 - 295	343 - 410	304.8		270.25	287.75		277.92	282.97		289.3	280.4
2 d 6.	Kolff WJ.	New Ways of Treating Uremia, JIA Churchill, London, 1947										
2 d 7.	Murphy WP, Swan RC, Walter C, Weller JM, Merrill JP.	J Lab Clin Med 40: 436, 1952. Essentially Krebs Henseleit, but with lower Mg and Ca.										
2 a 16.	Parsons FM, Hegstrom RM, Boen SI, Scribner BH.	Trans Am Soc Artif Intern Organs 10: 110-113, 1964										
2 a 17.	Made in concentrates by numerous manufacturers.	The mean concentrations used are given in 2 d 17 according to Parsons FM and Stewart WK, listed above in title.										
2 a 18.	Bijlsma F et al Nephron 27: 142-145, 1981.	"Low" acetate leaves the patients acidotic, "high" acetate leaves them in normal. Bijlsma's interpretation for the reasons for the acidosis are incorrect.										
2 b 6.	Kraut J et al. Clin Neph 15: 101, 1981.	Used HCO ₃ ⁻ and acetic acid.										
2 b 3.	Commercial source. CUBE Laboratories, 1201 Oak Street, Lakewood Colorado.											

EXAMPLE 42

[0160] The following example illustrates usage of Class I solutions for electrolyte and fluid therapy.

[0161] The most commonly used electrolyte solution used today, by those skilled in the art, is so called "physiological" salt, or "normal saline" by which is means 0.9% NaCl in H₂O in the U.S. or 0.95% NaCl in H₂O in the United Kingdom. (See Table IX solutions 1a1 and 1a2 respectively). These solutions, wherein the milliequivalent ratio of Na/Cl is 1, are distinctly different from normal human plasma wherein the ratio of Na/Cl ranges from 1.28 to 1.45 (N.E.J.M. 283, 1285, 1970). Infusion of such solutions has long been recognized to be undesirable leading to a pathological condition known

as "hyperchlormic acidosis". (See Black D.A., Lancet 1, 353, 1953, and Harrison's Textbook of Medicine, pp 230 to 236, 1983). The degree of the pathology induced by solutions where the ratio of Na/Cl is below the ratio 1.28-1.45 depends upon:

- 5 1) the quantity of solution infused relative to the volume and electrolyte content of the extra-and intracellular H₂O volume of the cells being contacted;
- 2) the rate of infusion of solutions;
- 3) the degree of existing pathology in the organism being contacted with such fluid;
- 4) the efficiency of the kidney in excreting the excess of Cl⁻ and Na⁺ being administered.

10 [0162] In this example, the replacement of plasma H₂O and salt content in the rat serves as a model stimulating the situation which might occur in a human patient when a severe burn over 50% of the body exists resulting in the loss of plasma H₂O and electrolytes into transudates and blisters over the surface of damaged skin. Three solutions for therapy will be used: standard 0.9% aqueous NaCl (composition 1a1 of Table IX), standard lactated Ringer's U.S. (composition 2a3 of Table X) and a modified redox-balanced Ringer's Lactate solution containing, with near-equilibrium couples, (L-lactate⁻/pyruvate⁻ and d-betahydroxybutyrate⁻/acetoacetate⁻), HCO₃⁻/CO₂ (composition 2b2 of Table X) in accord with the present invention. The composition of the 3 fluids are given in Table XIII below.

Table XIII - Composition of Fluids

		Normal Plasma N.F.J.M. 283, 1970	1 a 1 Isotonic NaCl	2 a 3 Lactated Ringer's Fluid	2 b 2 K-β Lactated Ringer's HCO ₃ ⁻ /CO ₂
Units					
Na	136 - 145	155	130	130	130
K	3.5 - 5.0	4	4	4	4
Ca	2.1 - 2.6	1.5	1.5	1.5	1.5
Mg free [Mg ²⁺] [0.53]	0.75 - 1.25				
Σ Eq Cations	142.7-153.2	155	137	137	137
Cl	100 - 106	155	109	96	29
HCO ₃ ⁻	26 - 28				
Z Pi	1 - 1.45				
SO ₄	0.32 - 0.94				
L - lactate	0.6 - 1.6			28 (d,1)	7
pyruvate					1
Lact/pyr					100

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	2	1	3		137	1.35		
10	D & Dibutyrate	HB/ acetate	acetoacetate					
15								
20								
25								
	Na/Cl	1.28 - 1.45	1.00					
	Glucose	3.9 - 5.6						
	or others	0.99 - 1.39						
	CO ₂							
	pH	7.35 - 7.45	6.0					
	Z adso	285 - 295	310					

METHODS

30 [0163] 250 fed male Wistar rats are each anesthetized and systematically burned with gasoline over approximately the lower 50% of the body surface. A blood sample is taken from each rat prior to administration of the burn, and then again two hours after the burn from a venous canula inserted into the saphenous vein. Each animal is placed in a restraining case.

35 [0164] In the opposite saphenous vein, a canula is inserted to measure plasma electrolyte content. Five minutes after the administration of each electrolyte solution, blood is drawn for electrolyte analysis. Each rat's liver is removed, freeze clamped and the redox and phosphorylation states of liver measured by the methods previously described by Veech et al. (J. Biol. Chem. 254, 6538-6547, 1979).

RESULTS AND DISCUSSION

40 [0165] It is observed that 1/2 hour after the gasoline burn, a series of weeping blisters develop over the lower 1/2 of each rat's body. The volume of the transudate within these blisters is estimated by measurement of area and thickness to contain 4ml of transudate or (250 x 0.07 = 17.5 ml blood volume) or about 40% of the rat's average total plasma volume. This deduction is confirmed by measurement of the rat hematocrit which is 55% while the Na⁺ is 155 millimoles per liter plasma and Cl⁻ is 110 millimoles per liter plasma due to fluid loss. In the untreated controls rats, the hematocrit is 44%. Each treated animal's blood pressure is falling, heart rate is increasing, and urine output ceases.

45 [0166] Each treated animal is judged to be in hypo-volemic shock and 6mls of the three different solutions are infused, by venous canula, over the next 10 minutes, into three different animals. Five minutes after completion of the infusion, electrolytes are drawn from the canula, the animals sacrificed, and the liver freeze clamped. The average blood electrolyte level, in each of the three groups of animals so infused, is shown in Table XIV below.

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Table XIV Composition of Plasma After Infusion

	Units m moles/ L fluid	Normal Plasma H.E.J.H. 283, 1285, 1970	1a1 Isotonic NaCl	2a3 Lactated Ringers	2b2 R-B Lactated Ringers HCO ₃ ⁻ /CO ₂
5	Na	135-145	150	143	138
10	K	3.5-5.0	5	5	5
15	Ca free ²⁺ [Ca ²⁺]	2.1-2.6 [1.06]	2.0	2.2	2.5
20	Mg free ²⁺ [Mg ²⁺]	0.75-1.25 [0.53]	1.0	1.0	1.0
25	Σ meq Cations	142.7-153.2	158	153.2	147.5
30	Cl	100-106	103	105	102
35	HCO ₃ ⁻	26-28	18	13	27
40	Σ Pi	1 - 1.45	1.5	1.2	1
45	L-lactate	0.6-1.8	5.0	21	5
50	pyruvate		0.3	1.0	0.7
55	Lact/pyr			21	7
	D-B-OH butyrate				2
	acetoacetate				0.7
	BHB/acac				3
	acetate				
	others				
	Σ meq anions	128.7-139.4	146.3	141.2	138.65
	Na/Cl	1.28-1.45	1.22	1.34	1.36
	Glucose or others	3.9-5.6	8.2	10	7
	CO ₂	0.99-1.39	1.14	0.82	1.35
	pH	7.35-7.445	7.30	7.30	7.4
	Σ m Osm	285-295			

Having now fully described the invention, it will be apparent to one of ordinary skill in the art that many changes and modifications may be made thereto without departing from the spirit or scope of the invention as set forth herein. [0167] It is observed that the animals given 1a1 (0.9% saline) solution each have hyperchloremic acidosis with a Na/Cl ratio of 1.22 and plasma pH of 7.30. The animals given solution 2a3 Ringer's Lactate solution each have lactic

acidosis with a plasma pH of 7.3 and an elevated [lactate]/[pyruvate] ratio. Both groups of these animals have low serum $[HCO_3]$ and have a compensated metabolic acidosis which requires that they hyperventilate off their CO_2 . In contrast, the animals given solution 2b2 (Redox-balanced Ringers Lactate with HCO_3/CO_2) each have a normal [lactate]/[pyruvate] ratio, a normal $[HCO_3]/[CO_2]$ ratio and a normal plasma pH. More importantly, each of these animals achieves a replacement of H_2O and electrolytes as required for continued life, but without inducing an abnormal Na/Cl ratio, an abnormal redox state, or an abnormal phosphorylation potential. No change in respiratory pattern is observed in the grave life-threatening situation. Solution 2b2 is then an improvement over the state of the art.

[0168] In Table 3 is given the results of the freeze clamping of the liver to illustrate the effects of these solutions on the nucleotide ratios in liver cells. These results indicate that only in the liver cells of the rats treated with the redox-balanced Ringer's lactate solution (Table X, solution 2b2) of this invention do these ratios approach normal values. Here, it is seen that administration of Na/Cl in 1:1 ratio leads to no change in the cytoplasmic $[NAD]/[NADH]$ but does cause an increase in the cytoplasmic $[ATP]/[ADP][Pi]$. With no intention to be bound by theory, the elevation of $[ATP]/[ADP][Pi]$ would be expected from equation 7 given in another section. The conventional Ringer's lactate (2a3) gives a profound and pathological decrease in the cytoplasmic $[NAD^+]/[NAD]$ to levels associated with alcoholic fatty liver. There is, of course, a predictable falls in the $[ATP]/[ADP][Pi]$, since the redox state of the cytoplasmic NAD-couple is directly and inversely linked to the cytoplasmic $[ATP]/[ADP][Pi]$ ratio as equation 5 shows.

[0169] In contract, the new Redox Balanced Ringer's Lactate solution of the present invention does not change the cytoplasmic $[NAD^+]/[NADH]$ from out of the normal range and causes no change in the $[ATP]/[ADP][Pi]$. Replacement of needed H_2O and electrolytes has been accomplished without inducing acidosis or any other recognized pathologic effects which can be demonstrated by using NaCl in 1:1 ratio or standard Ringer's Lactate in this simulation of a very common clinical situation.

Table XV.

Example 42 Case 1	Metabolite Contents of Freeze-Clamped Rat Liver in Rats After Infusion with Normal Saline, Ringer's Lactate, and Redox Balanced Ringer's Lactate with HCO_3^-/CO_2			
	Values are in umoles/g wet weight.			
	Normal Rat	0.9% NaCl Infusion	Ringer's Lactate	New R-B Ringer's Lactate with HCO_3^-/CO_2
	Solution	1.a.1	2.a.3	2.b.2
Glucose	7.3	8.0	13	8
Glucose 6-F	0.12	0.18	0.26	0.16
Dihydroxy-acetone-F	0.029	0.051	0.078	0.039
3-Phosphoglycerate	0.309	0.309	0.56	0.35
L-Lactate	0.444	0.812	14.8	5.2
Pyruvate	0.086	0.165	0.70	0.74
L-Lactate/pyr	5.16	4.92	21	7.00
3-PG/DHAP	10.6S	7.24	7.14	6.93

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Example 42
Case 1Table XVI Co-Factor Ratios of Freeze-Clamped Liver of Rat After Infusions with 0.9% Normal Saline, Ringer's Lactate, and Redox-Balanced Ringer's Lactate with $\text{HCO}_3^-/\text{CO}_2$

	Normal Rat	0.9% NaCl Infused Rat	Ringer's Lactate Infused Rat	New R-B Lactate with $\text{HCO}_3^-/\text{CO}_2$
	1.a.1	2.a.3	2.b.2	
Free Cytoplasmic $[\text{NAD}^+]/[\text{NADH}]$	1750	1790	429	1290
Free Cytoplasmic $[\text{ATP}]/[\text{ADP}][\text{Pi}] \text{ M}^{-1}$	14,000	20,900*	5,000*	12,000

* indicates change is significant at $p < 0.05$.Comparative Example 43

Use of Solutions for Parenteral Nutrition

[0170] The procedure used is identical to that utilized by Woods, Eggleston and Krebs in Biochem. J. (1970) 119, 501-510.

Animals and Diets

[0171] Female Wistar rats, each weighing 170-215g, are obtained and are fed on a standard small-animal diet

Reagents

[0172] D-Glyceraldehyde, 1- α -Glycerophosphate (dicylohexyl-ammonium salt) having a purity of 96% of the calculate L-form and other substances, nucleotides, coenzymes, and crystalline enzymes.

Liver Perfusion

[0173] The method of liver perfusion used is that described by Hems, Ross, Berry & Krebs (1966). The perfusion medium is the physiological saline (Krebs & Henseleit, 1932), containing washed aged human erythrocytes. The bovine serum albumin is dialyzed as a 10% solution (at 4°C) against three changes of physiological saline (Krebs-Henseleit) and gassed with $\text{CO}_2 + \text{O}_2$ (5:95).

[0174] The perfusion medium described by Hems et al. (1966) is used, which contains initially about 1 mM L-lactate [0.87 ± 0.05 S.E.M. (14) umol/ml] derived from the erythrocytes. To decrease the initial lactate concentration, the erythrocytes are washed five times with ten times their volume of physiological saline. This lowers the initial lactate concentration in the perfusion medium to 0.23 ± 0.02 S.E.M. (16) umol/mol. The medium is gassed with $\text{CO}_2 + \text{O}_2$ (5:95) during perfusion.

[0175] Into the perfusion of 150ml is added a sufficient quantity of two parenteral nutrient solutions, one containing 10 mM D-Fructose from a commercial source (5% Fructose in Electrolyte #75, Travenol, Facts and Comparison, August '83, p52b) and a new parenteral solution composition using glucose in place of fructose, a normal Na:Cl ratio, redox-balanced lactate, pyruvate and excess K as does Electrolyte #75. Glucose enters the metabolic sequence at a "safe entry" point as herein defined. The composition of each solution is given in Table XVII below.

Sampling of Liver

[0176] For the analysis of liver, samples are rapidly frozen *in vivo* or during perfusion, by using the deep cooled clamps of Wollenberger, Ristau & Schoffa (1960). The resulting disc of liver tissue is ground to a fine powder in a cooled mortar with frequency additions of liquid N₂. The liver powder is transferred to a tared centrifuge tube cooled in liquid N₂ and 4 ml of ice-cold 6% (w/v) HClO₄ is then added to each gram of liver powder with constant stirring. The resulting slurry is allowed to thaw and then is homogenized in the centrifuge tube at a low speed with a glass pestle. The homogenate is kept ice-cold for 30 minutes, centrifuged, and the resulting supernatant is brought to pH 6-7 with 20% (w/v) KOH to precipitate the excess of HClO₄ as KClO₄. The assays are carried out on the clear supernatant.

Preparation of Liver Aldolase

[0177] Livers of large (300-450g) rats are bled by perfusion *in situ* with cold isoosmotic KCl and then homogenized with 4 vol. of KCl. After centrifugation at 30000 x g for 20 minutes, the supernatant is fractionated with (NH₄)₂SO₄ as described by Leuthardt & Wolfe (1955). The final precipitate is taken up in a small volume of water (0.3 ml/g of original liver) and dialyzed against 200 vol. of water at 0C, changed every hour for 4h. The cloudy preparation is centrifuged and 0.1ml of 0.1 M EDTA is added to every 4ml of clear supernatant. Incubation for 1h at 25 ° C completely inactivated sorbitol dehydrogenase (EC 1.1.1.14)

Table XVII

COMPOSITION OF FLUIDS			
UNITS	(1)	(2)	(3)
n moles/L			
Na	136 - 145	40	40
K	3.5 - 5.0	35	35
Ca	2.1 - 2.6		
free [Ca ²⁺]	[1.06]		
Mg	0.75 - 1.25		
free [Mg ²⁺]	[0.53]		
meq Cations	142.7 - 153.2	75	75
Cl	100 - 106	47.5	29
HCO ₃	26 - 28		26
Pi	1 - 1.45	7.5	1.4
SO ₄	0.32 - 0.94		
I-lactate	0.6 - 1.8	20(d,l)	15.64
pyruvate			1.56
Lact/Pyr		(inf.)	10
d-Beta OH butyrate			
Acetoacetate			
Beta HB/acac			
Acetate			
Others			
meq anions	128.7 - 139.4	75	75
Na/Cl	1.28-1.45	0.84	1.36
Glucose	3.9 - 5.6		278
Fructose		278	
CO ₂	0.99 - 1.39		1.5
pH	7.35 - 7.45	-	7.4

Footnotes for Table XVII

(1) Indicates: Normal Human Plasma as reported in N.E.J.M. 283, 1285, (1970).

(2) Indicates: 5wt % Fructose in Electrolyte #75 (commercially available from Travenol as shown in "Facts & Comparisons" Aug. '83, p.52b).

(3) Indicates 5% Glucose in Electrolyte Solution for parenteral nutrition from this patient following our outlines of safe entry points and a normalized Na/Cl ratio and redox state.

Such a solution improves Solution 2 in this table.

Table XVII (continued)

COMPOSITION OF FLUIDS			
UNITS	(1)	(2)	(3)
n moles/L			
m OsM	285 - 295	428	429.5

Footnotes for Table XVII

(1) Indicates: Normal Human Plasma as reported in N.E.J.M. 283, 1285, (1970).

(2) Indicates: 5wt % Fructose in Electrolyte #75 (commercially available from Travenol as shown in "Facts & Comparisons" Aug. '83, p.52b).

(3) Indicates 5% Glucose in Electrolyte Solution for parenteral nutrition from this patient following our outlines of safe entry points and a normalized Na/Cl ratio and redox state.

Such a solution improves Solution 2 in this table.

(Hers, 1956), which would otherwise react with fructose. The final preparation, containing 35-45 mg of protein/ml, is stored at -18°C and is found to lose only about 30% activity in one year. In addition to aldolase activity, it also contains glycerol I-phosphate dehydrogenase (EC 1.1.1.8) activity and triose phosphate isomerase (EC 5.3.1.1) activity.

Other Aldolase Preparations

[0178] Chilled fresh rat and rabbit tissues are homogenized with 14 vol. of 1 mM-EDTA and centrifuged for 20 minutes at 30000 x g. The supernatant obtained is used in assays without further purification. A crystalline preparation of rabbit muscle aldolase is supplied by the Boehringer Corp. (London) Ltd.

Analytical Methods

[0179] ATP is determined by the method described by Lamprecht & Trautschold (1963), ADP and AMP are determined in the combined assay of Adam (1963), Pi was determined by the method described by Berenblum & Chain (1938) as modified by Martin & Doty (1949). Fructose I-phosphate, is determined by the method of Eggleston (1970). Fructose I, 6-diphosphate, is measured together with total triose phosphates in the combined assay of Bucher & Hohorst (1963); pyruvate, phosphoenolpyruvate, 2- and 3-phosphoglycerate are determined in sequence (Czok & Eckert, 1963). The references to other analytical methods are as follows: α -glycerophosphate (Hohorst, 1963b); L-(+)-lactate (Hohorst, 1963c); glucose 6-phosphate and fructose 6-phosphate (Hohorst, 1963c); glucose I-phosphate (Bermeyer & Klotzsch, 1963); glucose and fructose (Klotzsch & Bergmeyer, 1963); the sum of D-glyceraldehyde and glycerol (Pinter, Hayashi & Watson, 1967). For the fluorimetric determination of very low concentrations of glyceraldehyde 3-phosphate and dihydroxyacetone phosphate by the method of Veech, Raijman, Dalziel & Krebs (1969), a portion of the neutralized supernatant is shaken for 1 minute with Florisil (100-200 U.S. mesh) to remove flavins and then re-centrifuged before use. In livers perfused with fructose where the concentration of dihydroxyacetone phosphate is increased, it is determined by the spectrophotometric method of Bucher & Hohorst (1963). IMP is determined by a combination of paper chromatographic separation (Krebs & Hems, 1953) and a spectrophotometric assay. A portion of deproteinized liver extract (0.1 or 0.2 ml) is dried onto a 1 cm area on Whatman no. 1 chromatograph paper under a current of hot air. Duplicates, with and without added IMP standards (10 ul, 2mM solutions) on the same spot, are developed by descending chromatography with the isobutyric acid-ammonia solvent mixture described by Krebs & Hems (1953) for 45-48h at room temperature. After drying in a current of air, the papers are examined under u.v. light from a Chromatolite lamp (Hanovia Ltd., Slough, Bucks, U.K.) and absorbent areas are ringed by pencil. Average distances run from the starting line are: IMP 23 cm, ATP 27 cm, ADP 32, cm, AMP and inosine 37 cm. IMP areas, and a blank area of similar size before the starting line, are cut out and dropped into 4ml of 10mM potassium phosphate buffer, pH 7.0. After gentle mixing at intervals for 1 h, 3ml is removed and the extinction at 248nm in 1cm wide silica cells in a Zeiss spectrophotometer is determined. At this wavelength, the Emax. $\times 10^3$ for IMP is 12.3 (Deutsch, 1952). Recovery of standards by the whole procedure is 93-104%.

RESULTS

[0180] The values of metabolites found in freeze clamped liver are given in Table XVIII. Infusion of a fructose solution at a rate sufficient to raise the blood fructose level to 10mM

TABLE XVIII

Liver Contents of Metabolites (After 10 Minutes of Perfusion) Values Are In μ moles/g Wet Weight			
	(1)	(2)	(3)
D-Glucose	6.99	2.29	10
D-Fructose	about 0	10	about 0
Glucose 6-P	0.25	0.14	0.30
Fru-tose 1-P	0.23	8.72	0.25
Dihydroxyacetone -P	0.04	0.16	0.04
3 Phosphoglycerate	0.26	0.16	0.26
Lactate	0.79	1.34	0.79
Pyruvate	0.08	0.15	0.08

Footnotes for Table XVIII

(1) Indicates liver before perfusion.

(2) Indicates perfusion with solution 1 from commercial sources.

(3) Indicates perfusion with solution 2 from this patient.

20 drops liver and hence blood glucose level to 2.29mM and raises fructose 1, P, over 35 fold to 8.7 umoles/g. in contrast, using a glucose solution so as to raise the blood level to 10 mM glucose has no appreciable effects except for a small elevation of glucose 6-P.

25 [0181] In Table XIX, we see that raising blood fructose causes a three fold drop in ATP and a seven fold increase in IMP. The phosphate is simply stripped off the nucleotides to put on fructose 1-P. In addition, the inorganic Pi in liver drops from 4.2 to 1.7 umoles/g weight. Taken together, this is a picture of profound metabolic disorder in intracellular energy metabolism which may be avoided by using the alternative NaCl balanced, redox-balanced solution which uses nutrients of the "safe entry point class".

Table XIX

Liver Content of Nucleotides and Pi			
	Values are in μ moles/g wet weight		
	Control	Fructose Solution (1)	Glucose Solution (2)
ATP	2.22	0.51	2.22
ADP	0.78	0.66	0.78
AMP	0.26	0.20	0.26
IMP	0.165	1.14	0.165
Pi	4.25	1.67	4.25
metabolically active Pi	13.75	13.88	13.80

45 [0182] In Table XX, we see the $[NAD^+]/[NADH]$ ratio calculated from the $[l\text{-lactate}]/[\text{pyruvate}]$ ratio or the $[\text{malate}]/[\text{oxaloacetate}]$ ratio increases with fructose by two fold. As predicted by the equation of the K_{G+G} reaction, this is accompanied by an incredible elevation of the free $[\sum \text{ATP}]/[\sum \text{ADP}] [\sum \text{Pi}]$ ratio to $150,000 \text{M}^{-1}$, the highest values ever recorded. Whether near-equilibrium is reached in such an abnormal situation is not the point here. Rather, it is clear fructose abnormally decreases not only the total amounts of the adenine nucleotides (Table XIX) but also severely distorts their thermodynamic relationship thereby profoundly disordering the normal metabolic state of liver. In contrast, solution 2 has no effect, firstly because it does not violate the "safe entry point" concept, and also, because it has pH, redox and NaCl balance.

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TABLE XX

Example 2: Using Class 1 Solutions for Parenteral Nutrition

Liver Nucleotide Ratios

Control Liver	Perfused with Parenteral Nutrient (1)	Perfused with Parenteral Nutrient (2)
---------------	---------------------------------------	---------------------------------------

Free Cytoplasmic

$\frac{[NAD^+]}{[NADH]}$	912	1812	912
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Free Cytoplasmic*

$\frac{[\Sigma ATP]}{[\Sigma ADP][\Sigma Pi]}$	M ⁻¹	11,517	151,000	11,517
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*The free cytoplasmic $\frac{[\Sigma ATP]}{[\Sigma ADP][\Sigma Pi]}$ is calculated from equation 5 in this disclosure as described by Veech R. L., et al, J. Biol. Chem. 254, 6538-6547, 1979.

[0183] The example also illustrates the concept of "safe entry points" discussed herein: Compounds which may be included in solutions which directly contact living cells, without, for instance, first passing through the gut wall to be metabolically changed, constitute the group herein identified by having "safe entry points". Members of the "safe entry point group" where levels over 3mM may be used in fluids directly contacting cells are:

I-Lactate
pyruvate

d B-Hydroxybutyrate
acetoacetate

D-Glucose

The upper limits to which even these may be used depends upon the metabolite and medical situation and no upper limit can be set absolutely without such considerations. However, the sum of lactate and pyruvate is generally in the level of 10-12 mM in healthy, jogging adults. The sum of betahydroxybutyrate and acetoacetate is in the range 5-7 mM/L plasma in healthy individuals undergoing reducing three day fasts. (See Cahill G. F. and Aoki T.T. in Cerebral Metabolism and Neural Function (1980) Passonneau J.V., Hawkins R.A., Lust W.D. and Welsh F.A. eds; pp 234-242, Williams & Wilkins, Baltimore). Such levels may therefore be considered to be in a "Normal" range and used safely in most normal conditions excepting perhaps ketones in pregnant women where the decision by the physician will depend upon the medical necessity. (See Rudolf M.C.J. and Sherwin R.S., Clinics in Endocr. & Metab. 12, pp 413-428, 1983).

[0184] The toxicity of elevating blood glucose above 13 mM/l is well documented in the studies of the University Diabetes Group and must be balanced in the physician's judgment by the need for calories in the patient. Glucose is herein demonstrated, however, to be much less toxic than fructose.

[0185] Compounds which may not be used parenterally as "safe entry points" into the metabolic sequence, as currently practiced in the art, are:

Acetate
Glycerol

Lactate (without pyruvate)
 Pyruvate (without lactate)
 Fructose

- 5 [0186] The methods used in this example are found in the following reference: Woods HF, Eggleston LV, Krebs HA. The cause of the accumulation of fructose 1-P on fructose loading. Biochem J. 119: 501-510, 1970.

Example 44

- 10 Use of Class II Solutions for Peritoneal Dialysis

[0187] The procedure used here is similar to that utilized by Klim and Williamson in Biochem. J. (1982) 214, 459-464

Materials

- 15 [0188] Male Wistar rats weighing 213 + 35g (66), at time of death, are used: there are no significant differences between the mean body weights of the experimental groups. They are maintained on a standard small animal diet, and water ad libitum in an animal house with lights on from 08:00 to 20:00h. Chronic uremia is induced by the five-sixths bilateral nephrectomy technique (Morrison, 1966). Uremic rats are allowed approximately 14 days to recover
 20 from the last operation before use.

Peritoneal-Dialysis Solutions

- 25 [0189] A commercial peritoneal dialysis solution is used, containing 45 mM acetate and 1.5% glucose (83mM) and compared to a new dialysis solution of the present invention (Example 3). The composition of the two solutions is comparatively shown in Table XXI. Control rats are simply given glucose to raise their blood levels to those occurring in dialyzed animals.

- 30 [0190] The methods of measurement of liver metabolites are those of Veech and are described amply in the literature such as Veech et al. J. Biol. Chem. 254 6538-6547, 1979; Veech, Eggleston & Krebs Biochem. J. 115, 609-619, 1969 and Veech et al. FEBS Letts., 117, K65-72, 1980.

TABLE XXI

Composition of Dialysis Fluids			
Units m moles	Normal Plasma	Commercial Fluid	New Fluid
L Fluid	(1)	(2)	(3)
Na	136 - 145	140	140
K	3.5- 5.0	4	4
Ca	2.1- 2.6	2.0	2.0
free [Ca ²⁺]	[1.06]		
Mg	0.75- 1.25	0.75	0.75
Sigma mEq.Cations	142.7 - 153.2	150	150
Cl	100 - 106	105	105
HCO ₃	26 - 28		29
Sigma Pi	1 - 1.45		
SO ₄	0.32- 0.94		
L-lactate	0.6 - 1.8		8.21
pyruvate			1.79

Footnotes for Table XXI

(1) indicates: Normal plasma N.E.J.M. 283, 1285, 1970.

(2) indicates: Commercial Fluid-Peritoneal dialysis with 1.5% Glucose. American McGraw, Facts and Comparisons, October 1982, page 704.

(3) indicates: New fluid-improved peritoneal dialysis fluid formulated in this disclosure is meant to mimic the ideal commercial fluid. This new fluid is not to be taken as "ideal" but is simply a way of illustrating why acetate should not be used. A better fluid would also contain HCO₃/CO₂, Lactate/pyr & Beta-HB-/AcAc but would have an increased Na:Cl ratio of between 1.38 to 1.41 to increase alkali reserve in the chronically acidotic uremics. Cl could be 100, HCO₃⁻ of 34 with [CO₂] of 1.7mM as an example of a fluid designed in conformity with the principles outlined herein. Such fluids have 1) redox balance and hence normal phosphorylation state achieved with 2) pair of ratioed couples so as to achieve a normal M desired NaCl ratio 3) while causing less pathological consequences than present art allows.

TABLE XXI (continued)

Composition of Dialysis Fluids				
	Units m moles	Normal Plasma	Commercial Fluid	New Fluid
5	Lact/pyr			4.6
	d-Beta-OH butyrate			3.24
	Acetoacetate			2.76
	BetaHB/acac			1.17
10	Acetate		45	
	Sigma mEq anions	128.7 - 139.4	150	150
	Na/Cl	1.28-1.45	1.33	1.33
	Glucose	3.9 - 5.6	83	83
15	CO ₂	0.99 - 1.39		1.5
	pH	7.35- 7.45	5.5-6.5	7.4
	Sigma m OsM	285 - 295	379.75	379.75

The values of metabolites in rat liver are given after seventy minutes of peritoneal dialysis in Table XXII.

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(Electrolyte 14)

Table XXII

	Control	(1) Acetate Peritoneal Dialysis	(2) Redox-Balanced Dialysis Fluid
N	(13)	(10)	(10)
Values are given in n moles/g wet weight liver.			
Dihydroxy-	46	53	69
acetone P	+_3	+_5	
3-Phospho-	294	405	294
glycerate	+_15	+_27	
l-Lactate	727	743	6081
	+_36	+_70	
Pyruvate	158	98	1326
	+_13	+_9	
d-Beta Hydroxy-	117	151	2400
butyrate	+_20	+_12	
Acetoacetate	100	117	1380
	+_19	+_8	
Acetate	20	33000	20

[0191] In Table XXIII are given the changes in liver content of divalent cations, Pi, PPi and total metabolizable phosphate containing compounds after such treatment.

TABLE XXIII
Changes in Mg, Ca, Pi and PPi Content in Rat Liver During Dialysis

Values in umoles/g wet weight liver.					
	(1)		(2)		
	Acetate	Change	New	Change	
	Dialysis	Induced	Dialysis	Induced	
	by Acetate		by new		
15	Control		Dialysis		Dialysis
	(16)	(16)			
20	Ca	1.06	1.76 +.70	1.06	0
Mg	11.76	12.94 +1.18	11.8	0	
25	Inorganic Pyrophosphate				
(PPi)	.018	0.198 +0.18	0.018	0	
30	Sigma Adenine				
Nucleotides	7.95	9.43 +1.48	7.95	0	
35	Sigma Guanine				
Nucleotides	1.56	1.97 +0.41	1.56	0	
40	Sigma Glycolytic				
Pi	0.65	1.65 +0.06	0.85	+.2	
45	Sigma Metabolic				
Pi	13.75	17.97 +4.22	13.95	+.2	
50	from all measured				
55	Metabolites				

[0192] It should be remembered that normal hemodialysis with 35 mM acetate makes the abnormal elevation in PPi reach 100 times normal with a quadrupling of liver Ca at the expense of bone stores of calcium. It is thus exaggerated in every way. Solutions containing 35mM Na Acetate currently account for about of 80% of hemodialysis in the U.S. The increased Pi demonstrated herein during acetate dialysis is "hidden" in liver and flows out (into blood) after dialysis accounting for why such patients remain persistently hyperphosphotemic leading to much current pathology found in chronic dialysis patients.

[0193] The data presented in Table XXIII clearly show that peritoneal dialysis, with acetate containing fluids, leads to gross elevations of liver inorganic pyrophosphate and liver calcium. While not widely appreciated, inorganic pyrophosphate (PPi) is an important controller of cellular metabolic pathways of many types. See Lawson J.W.R. et al. in Gluconeogenesis, 1975 (Hanson R.W. & Mehlman M.A. Eds) pp 481-511, John Wiley & Sons, New York). Changes in PPi are therefore likely to be of widespread significance. The 70% increase in liver calcium is, of course, clearly large and of potential significance because of the importance calcium plays as an activator of many intracellular protein kinases.

[0194] Finally, Table XXIII shows that acetate induces a rapid increase of 4.2 umoles/g wet weight of the liver's rapidly metabolizing phosphate compounds. It derives this excess Σ Pi from the blood and other phosphate stores. When the acetate is finally metabolized, this phosphate returns to the blood where Pi is 1-1.45mM. Since liver and blood are

roughly equal in weight in the normal adult, this movement of Σ Pi out of liver must inevitably lead to the hyperphosphatemia which is a major and persistent pathological sequela of uremia treated by current dialysis practice. This persistent elevation of blood Pi leads to chronic hyperparathyroidism, hypocalcemia, accelerated bone disease, ectopic calcification of tissue and many other causes of morbidity and even mortality in chronic renal disease. Because the phosphate accumulates in the liver during acetate dialysis, it is effectively "hidden" from the beneficial effects which dialysis is trying to obtain, namely the removal of excess dietary Σ Pi which is taken in by the patient during the intra-dialysis periods.

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(Electrolyte-14)

TABLE XXIV

Table XXIV gives the results obtained for the redox and phosphorylation states calculated, as described in Equations 4 and 5. Values are given as means + S.E.M.

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		(1)	(2)
	Control	Acetate Dialysis	New Dialysis
25 N	(5)	(6)	(6)
Cytoplasmic free			
[NAD ⁺]	1944	1209*	about 1944
[NADH]	+ 94	+ 88	
Mitochondrial free			
[NAD ⁺]	18.2	17.4	about 18.2
[NADH]	+2.3	+2.6	
cytoplasmic			
$\frac{[\Sigma ATP]}{[\Sigma ADP][\Sigma Pi]}$ M ⁻¹	25,800	13,700*	about 25,800
	+ 3,200	+ 2,600	

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*indicates significant difference at $P > 0.05$.

[0195] The use of acetate in a peritoneal dialysis fluid obviously causes a significant decrease in the free cytoplasmic [NAD⁺]/[NADH] and an even more profound decrease in the cytoplasmic $[\Sigma ATP]/[\Sigma ADP][\Sigma Pi]$ ratio. This is so because the free [NAD⁺]/[NADH] ratio of cytoplasm is directly linked to the free cytoplasmic $[\Sigma ATP]/[\Sigma ADP][\Sigma Pi]$ by equation 5. (See Veech, et al. *J. Biol. Chem.* 254, 6538-6547, 1979). On page 704 of Facts and Comparisons, October, 1982, are listed 16 peritoneal dialysis solutions, using 35 to 45 mMolar (d,l)-lactate in commercial peritoneal dialysis solutions made by four different commercial manufacturers. These solutions, in addition to the 7 commercial acetates containing peritoneal dialysis solutions, make up the current state of the art. None achieve the normal Na/Cl ratio they desire in the manner described herein.

[0196] No example of the effects of using 35 to 45 mM L-lactate alone, in a peritoneal dialysis solution, need be given. It is by now obvious, from the teachings here presented, that such solutions are entirely without redox balance but indeed induce a profound lactic acidosis with a pathological decrease in the free cytoplasmic [NAD⁺]/[NADH] and the free cytoplasmic $[\Sigma ATP][\Sigma ADP][\Sigma Pi]$ to which it is linked by equation 5. It is also obvious that redox-balanced solutions, made by the principles outlined here, would be an advance in the present art.

Example 45

Hemodialysis

- 5 [0197] Using hemodialysis equipment, which is the current major type in use, (see Keshaviah et al., CRC Critical
Reviews in Biomedical Engineering 9, 201-244, 1983) and using the most common type of dialysis fluid currently in
use in the art, which uses between 35 to 45 mMoles/L of Na acetate to correct the anion gap, (see Parsons F.M. &
Stewart W.K., The Composition of Dialysis Fluid in Replacement of Renal Function by Dialysis, 2nd edition (1983)
(Drukker W., Parsons F.M. & Maher J.F., eds) pp 148-170, (Martinus, Nijhoff, Hingham) we may obviously predict the
10 effects, upon body organs such as the liver, of such treatment.

Methods

- 15 [0198] Rats are made uremic as described in the previous example. After five days, they are fasted, attached to a
miniature hemodialysis apparatus, heparinized and dialyzed with two different solutions, one representing the most
common types of currently used hemodialysis solutions, and another where the anion gap is made up without the use
of $\text{HCO}_3^-/\text{CO}_2$, but instead, with the use of L-lactate/pyruvate and D-β-Hydroxybutyrate/acetoacetate as are given in
the class 2-a solutions in this disclosure, as for example 2-a-8, Redox-Balanced Ringers. It should be understood that
I do not conclude such a solution as 2-a-8 is the best solution for such a purpose, but I shall show it is superior to the
existing art and may be used in the bulk of existing apparatus which contain deaerators* and currently use acetate
containing hemodialysis fluids. (Keshaviah et al. CRC Critical Reviews in Biomedical Engineering 9, 201-244, 1983).
A few current machine, typically 1 out of 10 in the dialysis centers I have surveyed have dialysis machines of the type
described by Miller J.H. et al. Trans Am Soc Artif Internal Organs 25, 404-408, 1979. Such machines can use HCO_3^-
containing solutions. Such $\text{HCO}_3^-/\text{CO}_2$ solutions are preferred.
25 [0199] The compositions of the two example solutions are given in Table XXV.

Example 4 Table XXV Solution for Hemodialysis of a Uremic Rat.

Units	(1) Usual Hemo- dialysis Solution	(2) Redox- Balanced Hemodialysis Solution	
K	3.5 - 5.0	0 - 1.5	4
Ca free [Ca ²⁺] (1.06)	2.1 - 2.6	1.25	1.5
Mg free [Mg ²⁺] (0.53)	0.75 - 1.25	0.5	-
Zn ²⁺ Cations (142.7-153.2	133.5-140	137	136
C1	100 - 106	100.5	96
HCO ₃ ⁻	26 - 28		
Cl ⁻	1 - 1.45		
SO ₄ ²⁻	0.32 - 0.94		
L-lactate ⁻	0.6 - 1.8		
pyruvate		1.9	
Lact/pyr			17

D β-β-hydroxybutyrate	5
acetoacetate	2
B Hb/ acac	2.5
acetate	33.5-40
Other	
Σ Eq anions	128.7-139.4
[H/C]	1.28 - 1.45
Glucose or others	3.9 - 5.6
CO ₂	0.99 - 1.39
NH	7.35 - 7.45
Σ solns	285 - 295
	375
	272.5

(1) The composition of the usual hemodialysis solution is taken from Farson's and Stewart, 1983, cited above.

(2) Composition of solution 2-3-B is taken from this application except that the lactate/pyruvate ratio is decreased to 17 to account for the absence of glucose since most current hemodialysis fluids use acetate without glucose. This composition is chosen to compare with current acetate hemodialysis practice. This solution should not be taken as "ideal" or even as recommended, but rather illustrative.

[0200] The rats are dialyzed with solutions 1 and 2 for four hours; the animals are sacrificed and the livers freeze clamped. A group of normal rats, starved 48 hours, are also sacrificed and their livers freeze clamped to serve as controls. Metabolites are measured, as previously described.

[0201] In Table XXVI, we see that both acetate and new redox-balanced dialysis fluids elevate liver sugar and the first portion of the gluconeogenic pathway. During acetate dialysis, changes occur throughout the gluconeogenic sequence and the ratio of one metabolite to another changes.

Table XXVI

Liver Metabolites from Rats Dialysed with Acetate Dialysis Fluid versus New Redox-Balanced Dialysis Fluids without HCO ₃ ⁻ /CO ₂			
Values are given as means + S.E.M. in nmoles/g wet weight. A * indicates a significant difference from normal rats at P < 0.05 as judged by Student's T Test			
	Untreated Starved Rats	Commercial Acetate Dialysis	New Redox-Balanced Dialysis
N	13	10	
10 ⁻³ x glucose	4.81+0.21	7.94+0.42	7.2*
glucose 6-P	59+2	99*+10	88.5*
glucose 1-P	7+1	11*+1	10.5*
fructose 6-P	17+1	32*+3	25.2*
fructose 1,6 bis-P	4.6+0.4	23*+6	6.9
CHAP	11+1	36*+4	16.5
3-phosphoglycerate	156+14	581*+62	234
PEP	73+5	330*+40	110
pyruvate	10+1	27*+6	1260*
L-lactate	171+17	721+208	21300*
L-malate	268+28	592*+84	402
α-ketoglutarate	118+13	86+17	177

Table XXVI (continued)

Liver Metabolites from Rats Dialysed with Acetate Dialysis Fluid versus New Redox-Balanced Dialysis Fluids without HCO ₃ ⁻ /CO ₂			
Values are given as means + S.E.M. in nmoles/g wet weight. A * indicates a significant difference from normal rats at P < 0.05 as judged by Student's T Test			
	Untreated Starved Rats	Commercial Acetate Dialysis	New Redox-Balanced Dialysis
N	13	10	
isocitrate	17+2	41*+3	25.5
citrate	308+42	944*+85	462
acetoacetate	638+33	643+66	1330*
D-B OHbutyrate	1643+75	983*+83	3300*
LDP-glucose	350+15	367+25	350
UTP	205+9	186+8	205
acetate	20	25000	20

[0202] In Table XXVII are presented the changes in the controlling co-factor ratios after the two types of dialysis.

TABLE XXVII

Free Nucleotide Ratios in Freeze Clamped Rat Liver After Acetate and Redox-Balanced Hemodialysis

Values are given as mean + S.E.M. An * indicates a significant difference from control values of P < 0.02 as judged.

	Starved Control	Acetate Dialysis	Redox-Balanced Dialysis
(n)	(13)	(10)	
Cytoplasmic [NAD ⁺] [NADH]	587 + 86	391 + 35	587
$10^3 \times$ [NADP ⁺] [NADPH]	7.3 + .7	2.1* + .3	7.3
$\frac{[\Sigma ATP]}{[\Sigma ADP][\Sigma Pi]M^{-1}}$	3710 + 580	2090 + 280	3710
mitochondrial [NAD ⁺] [NADH]	8.1 + 0.7	13.8* + 1.4	8.1

[0203] In Table XXVII we see that acetate dialysis causes oxidation of the mitochondrial [NAD⁺]/[NADH] ratio and reduction of the free cytoplasmic [NADP⁺]/[NADPH] ratio while redox-balanced dialysis causes no change as judged by the isocitrate/α-ketoglutarate ratio.

[0204] In Table XXVIII are presented the results of the measurement of the Ca, Mg, phosphate and pyrophosphate content of rat liver after acetate versus redox-balanced hemodialysis.

TABLE XXVIII

Changes in Mg, Ca and Phosphate Compounds in Liver Following Acetate versus Redox-Balanced Hemodialysis.			
n	Control 13	Acetate Hemodialysis 10	Redox-Balanced Hemodialysis
Ca	1.33	+2.89	0
Mg	10.1	+1.8	0
P Pi	0.024	+ 2.00	0
Pi	4.22	+3.73	0
Σ Adenine Nucleotide Pi	9.32	+0.07	0
Σ Guanine Nucleotide Pi	1.76	+0.19	0
Σ Glycolytic Pi	0.36	+0.86	.50
Σ Pi Increased from All measured metabolites	15.71	+8.85	.50

[0205] We see in Table XXVIII that acetate dialysis raises inorganic pyrophosphate 200 times while redox-balanced dialysis makes no change. Acetate hemodialysis increases liver calcium three fold; redox-balanced dialysis makes no change. Acetate hemodialysis increases total liver metabolizable phosphate by 8.8 m moles/g, while redox-balanced dialysis increases liver metabolizable phosphate by only 0.5 m moles/g, or 16 times. The "hidden" phosphate, inaccessible to dialysis after acetate hemodialysis, is the largest ever seen. The metabolic pathology is therefore even greater than that in Example 44.

Example 46

[0206] Solutions of this invention when administered not only regulate redox state and phosphorylation, but also further tend to normalize the following states:

- (1) Distribution of water between intracellular and extracellular fluid.
- (2) Distribution of the inorganic electrolytes Na^+ , K^+ , Cl^- and Ca^{2+} between intracellular and extracellular fluid, and
- (3) Transmembrane cellular potential. ΔE

[0207] The following equations state the governing scientific laws involved:

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0. Eqn 0 - The Second Law

J. Willard Gibbs. On the equilibrium of heterogeneous substances. *J. Conn. Acad. Sci.* 1876 III : 543.

0 - 1 Definition of Gibbs Free Energy and Other Properties of State:

$$G = H - TS$$

where:

- G ~ Gibbs free energy
- H ~ Enthalpy or heat content
- T ~ absolute temperature
- S ~ Entropy, or state of randomness or disorder

0 - 1 a Entropy may be more rigorously defined by statistical and quantum mechanics in the Boltzmann Equation:

$$S = k_B \ln \Omega$$

where:

- S ~ Entropy
- k_B ~ Boltzmann constant = $\frac{R}{\text{Avogadro's number}}$

$$\Omega \sim \text{Degeneracy} = 1.38 \times 10^{-23} \text{ J/K}$$

$$\Delta G = \Delta H - T \Delta S$$

where ~ change in

0 - 3 Standard Free Energy ~ ΔG°

$$\Delta G = \Delta G^\circ + RT \ln \frac{[products]}{[reactants]}$$

where:

$$R \sim \text{gas constant} \\ = 1.987 \text{ calories}/\text{mole}$$

$$\text{and } \frac{R}{R_K} \sim 273 + {}^\circ C$$

$$T = \frac{R}{R_K} \\ \ln \sim 2.303 \log_{10}$$

$$\Delta G^\circ = -RT \ln K_{eq}$$

where:

$$K_{eq} \sim \frac{[products]}{[reactants]}$$

0 - 4 At equilibrium, $\Delta G = 0$, so in $A + B \rightleftharpoons C + D$

$$\Delta G = -RT \ln K_{eq} + RT \ln \frac{[C][D]}{[A][B]}$$

where:

$$[] \sim \text{activity or } \sim \text{concentration}$$

"A theory is the more impressive the greater the simplicity of its premises, the more different are the kinds of things it relates, and the more extended is its range of applicability... It is the only physical theory of universal content which I am convinced, that within the framework of applicability of its basic concepts, will never be overthrown."

A. Einstein

I Eqn 1 - The Henderson-Hasselbalch Equation

[0208] The major buffer and controller of extra and intracellular pH. Henderson LJ. Blood, A Study in General Physiology. Silliman Lectures, Yale University Press, 1928

1.a

$$pH = pK_a + \log \frac{[HCO_3^-]}{[CO_2]}$$

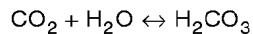
where:

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$pK_a = 6.10$ at 38°C and serum concentrations of electrolytes

1.b The solubility of CO_2 in fluid, i.e. dissolved CO_2 gas plus H_2CO_3 from:

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$$[\text{CO}_2] \text{ in mmol/liter} = \frac{\text{pCO}_2 \text{ in mmHg}}{760 \text{ mmHg}} \cdot \frac{\alpha \text{ ml CO}_2/\text{ml of H}_2\text{O}}{22.26 \text{ L/mole}} \cdot \frac{1000 \text{ mmol}}{\text{mole}}$$

$$\alpha_{\text{CO}_2} = 0.553/\text{ml serum H}_2\text{O at } 38^\circ\text{C from:}$$

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van Slyke DD. J Biol Chem 73: 765-799, 1928

1.c The pH of a bicarbonate containing solution to which has been added a carbocyclic acid such as acetic, lactic, acetoacetic acid with a pK' in the 3 to 4 range and where the concentration of HCO_3^- is much larger than the concentration of carboxycyclic acid:

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$$\text{pH} = pK_a' - \log \left\{ \frac{[\text{HCO}_3^-]}{2([\text{HCO}_3^-] - [\text{HA}])} - \frac{1}{2} \right\}$$

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Thus adding 1.8 mM Hlactate and 0.2 mM Hpyruvate to 25 mM NaHCO_3 yeids what pH?

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$$\begin{aligned} \text{pH} &= pK_a' - \log \left\{ \frac{[25]}{2([\text{HCO}_3^-] - [\text{HA}])} - \frac{1}{2} \right\} \\ &= 6.1 - (1.36) \\ &= 7.46 \end{aligned}$$

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III Donnan Equilibrium Equation

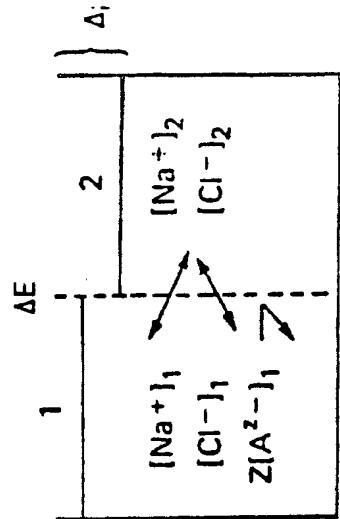
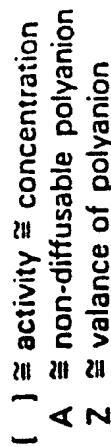
Donnan FG. - Electrochem 17: 572, 1911
 Donnan FG. Chem Rev 1: 73-90, 1924.

1. From Gibbs (Eqn 0)

$$RT \ln \frac{[Cl^-]_1}{[Cl^-]_2} + RT \ln \frac{[Na^+]_1}{[Na^+]_2} = 0$$

Or:

$$1. \therefore \frac{[Cl^-]_1}{[Cl^-]_2} = \frac{[Na^+]_2}{[Na^+]_1}$$



$$\text{Therefore: } \frac{[Cl^-]_1}{[Cl^-]_2} = \frac{[Cl^-]_2}{[Cl^-]_1 + Z[A^{z-}]_1} = \frac{[Na^+]_2}{[Na^+]_1 + Z[A^{z-}]_1} = \frac{[Na^+]_2}{[Na^+]_1}$$

and for polyvalents:

$$\left\{ \frac{[Anions]_1}{[Anions]_2} \right\}^{1/z} = \left\{ \frac{[Cations]_2}{[Cations]_1} \right\}^{1/z} \text{ cations}$$

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2. From the Law of Electrically Neutrality:

$$[\text{Na}^+]_2 = [\text{Cl}^-]_2$$

$$[\text{Na}^+]_1 = [\text{Cl}^-]_1 + z [\text{A}^{z-}]_1$$

3. Quadratic equation:

$$x = \frac{-b \pm \sqrt{b^2 - 4ac}}{2a}$$

Example:
 Consider albumin dialysed against 100% CO_2 / 3.13 NaHCO₃ buffer with 1.17 mM albumin (i.e. 8% solution). Hypothetically keep charge on albumin at -20/mole.

$$\frac{[\text{HCO}_3^-]_i}{[\text{HCO}_3^-]_0} = \frac{[\text{HCO}_3^-]_0}{[\text{HCO}_3^-]_i + 20[\text{Alb}^{-20}]_i} = \frac{[\text{Na}^+]_0}{[\text{Na}^+]_i}$$

$$\frac{[\text{HCO}_3^-]_i}{[3.13 \times 10^{-3}]_i} = \frac{[3.13 \times 10^{-3}]_0}{[3.13 \times 10^{-3}]_i + 20[1.17 \times 10^{-3}]_i}$$

$$[\text{HCO}_3^-]_i = 0.4 \times 10^{-3} \text{ M}$$

II Eqn 2 Multicomponent Donnan Equilibrium System for Solutions Such as the Hemodialysis of Blood Plasma Electrolytes

where $\Delta p = 0$ and all components but albumin are permeant. Subscript 0 ~ in dialysis fluid, subscript 1 ~ in patient's plasma. Δp ~ difference in pressure.

$$2.3. \quad \frac{[Na^+]}{[Na^+]_0} = \frac{[K^+]}{[K^+]_0} = \frac{[(Ca^{2+})_1]}{[(Ca^{2+})_0]} = \frac{[(Mg^{2+})_1]}{[(Mg^{2+})_0]} = \frac{[Cl^-]}{[Cl^-]_0} = \frac{[(HCO_3^-)_1]}{[(HCO_3^-)_0]} = \frac{[(OH^-)_1]}{[(OH^-)_0]} = \frac{[(H_3O^+)_0]}{[(H_3O^+)_{0'}]} = \frac{[(Pyr^-)_0]}{[(Pyr^-)_{0'}]} = \frac{[(Ac^-)_0]}{[(Ac^-)_{0'}]} = \frac{[(NH_3)_0]}{[(NH_3)_{0'}]} = \frac{[(acet^-)_0]}{[(acet^-)_{0'}]} = \frac{[(acet^-)_1]}{[(acet^-)_{1'}]} = \frac{[(acet^-)_{0'}]}{[(acet^-)_1]} = \frac{[(acet^-)_{0''}]}{[(acet^-)_{1''}]}$$

Statement of electrical neutrality on two sides of an uncharged membrane

2.b.1.
$$[\text{Na}^+]_0 + [\text{K}^+]_0 + 2[\text{Ca}^{2+}]_0 + 2[\text{Mg}^{2+}]_0 = [\text{Cl}^-]_0 + [\text{HCO}_3^-]_0 + 1.81 \cdot \text{Pi}^{-1} \cdot \theta_0 + [\text{acet}^-]_0 + [\text{DMSO}]_0 + [\text{acet}^-]_0 + [\text{proton}]_0$$

2.b.2.
$$[\text{Na}^+]_i + [\text{K}^+]_i + 2[\text{Ca}^{2+}]_i + 2[\text{Mg}^{2+}]_i = [\text{Cl}^-]_i + [\text{HCO}_3^-]_i + 1.81 \cdot \text{Pi}^{-1} \cdot \theta_i + [\text{acet}^-]_i + [\text{DMSO}]_i + [\text{acet}^-]_i + [\text{proton}]_i$$

Distribution of cations on two sides of the membrane:

$$[X_4^+] = [X_4^+]_0 \frac{e^{-k_2 t}}{[Mg]^2} ; \quad [Ca^{2+}] = [Ca^{2+}]_0 \frac{e^{-k_2 t}}{[Mg]^2} ; \quad [Mg^{2+}] = [Mg^{2+}]_0 \frac{e^{-k_2 t}}{[Mg]^2}$$

Distribution of Anions:

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$$2.4 \quad [Cl^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [Cl^-]_0; \quad [HCO_3^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [HCO_3^-]_0; \quad [acet^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [acet^-]_0; \quad [Pi]_i = \frac{[Na^+]_0}{[Na^+]_i} [Pi]_0;$$

$$[tar^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [tar^-]_0; \quad [pyr^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [pyr^-]_0; \quad [acet^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [acet^-]_0; \quad [BHB^-]_i = \frac{[Na^+]_0}{[Na^+]_i} [BHB^-]_0$$

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Now solving for $[Na^+]_i / [Na^+]_0$ for a dialysis fluid₀ of known composition:

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$$\frac{[Na^+]_i}{[Na^+]_0} [Cl^-]_0 + [K^+]_0 + 2\frac{[Na^+]_0}{[Na^+]_i} [(Ca^{2+})_0 + (Mg^{2+})_0] =$$

$$\frac{[Na^+]_i}{[Na^+]_0} [Cl^-]_0 + [HCO_3^-]_0 + [acet^-]_0 + [Pi]_0 + [BHB^-]_0 + 1.0 \frac{[Na^+]_0}{[Na^+]_i} + 0.8 \frac{[Na^+]_0}{[Na^+]_i} + \frac{[Na^+]_0}{[Na^+]_i} [prot^+]$$

and:

$$\frac{2.4}{[Na^+]_0 + [K^+]_0} = \frac{2[(Ca^{2+})_0 + (Mg^{2+})_0]}{[Na^+]_i^2 + [HCO_3^-]_i^2 - [Na^+]_i^3 - [Na^+]_i \cdot [Na^+]_0 \cdot [Na^+]_i (-0.8)} =$$

$$\frac{[Na^+]_0^2}{[Na^+]_0} \frac{[Na^+]_3}{[Na^+]_0}$$

$$[Cl^-]_0 + [HCO_3^-]_0 + [acet^-]_0 + [Pi]_0 + [BHB^-]_0$$

Plasma [concentration] = 0.935 × plasma H_2O [concentration]

III Eqn 3. Nernst Equation - ΔE

50 [0209] Nernst W. Theoretical Chemistry 4th Edition, 1904, McMillan, London. See also Silliam Lecture, 1906, Yale U. Press, New Haven.

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3. $\Delta E = - \frac{RT}{nF} \ln \frac{[\text{anion}^-]_{\text{outside}}}{[\text{anion}^-]_{\text{inside}}}$

or:

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$$\Delta E = -\frac{RT}{nF} \ln \frac{[\text{cation}^+]\text{inside}}{[\text{cation}^+]\text{outside}}$$

5 where:

at 38°C T ~ 311 °K

R, the gas constant ~ 8.314 hiykes/degree/mole

n ~ number of equivalents

10 F, the Faraday, ~ 96,494 coulombs

AE ~ potential in volts

[0210] To convert ln to \log_{10} , multiply by 2.303 From Cornell N, Anal Biochem 1980; 102: 326-331, for isolated hepatocytes from starved rats incubated in Krebs-Henseleit.

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$$\Delta E = -0.0617 \log \frac{[0.128 \text{ M Cl}^-]\text{outside}}{[0.041 \text{ M Cl}^-]\text{inside}}$$

20 $\Delta E = -0.0305 \text{ V or } -30.5 \text{ mV}$

and for cat brain, from Eccles JC. The Physiology of Nerve Cell, 1957, Johns Hopkins U Press, Baltimore.

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$$\Delta E = -0.0617 \log \frac{[0.125 \text{ M Cl}^-]\text{outside}}{[0.009 \text{ M Cl}^-]\text{inside}}$$

$\Delta E = -0.0705 \text{ V or } -70.5 \text{ mV}$

3.b Redox Potential of Half Reactions

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$$E_h = E^\circ + \frac{RT}{nF} \ln \frac{[\text{oxidized}]}{[\text{reduced}]}$$

where:

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R ~ 8.31431 J/°K/mole

T ~ °K

n ~ number of electrons

F ~ Faraday ~ 96,494 coulombs

ln ~ 2. 03 \log_{10}

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IV Eqn 4 Redox State Equations. $[NAD^+]/[NADH]$ or $[NADP^+]/[NADPH]$.

Near equilibrium reactions are given a number depending upon location. The E°' of the $[NAD^+]/[NADH]$ couple at pH 7 is -0.37V. That of the $[NADP^+]/[NADPH]$ couple, -0.35 V.

Abbreviation Definition of K_{eq}	Value of K_{eq} at pH = 0 \pm eq at pH 7 at pH 7.0 <u>oxidized</u> <u>reduced</u>	Value of E°' at pH 7.0 $CO_2 = 1.5 \text{ mM}$ or 0.5 mM NH_4^+ or 1 mM Pi^-
Enzyme No.		

Cytoplasmic NAD - linked Dehydrogenases

4 c 1 $K_{LDH} = \frac{[\text{Pyruvate}]^{1/2}[\text{NADH}]^{1/2}}{[\text{l-lactate-}]^{1/2}[\text{NAD}^+]}$	$1.1 \times 10^{-11} \text{ M}$	1.1×10^{-4}	-0.201
EC 1.1.1.27			
4 c 2 $K_{MDH} = \frac{[\text{D-oxaloacetate-}^{2-}]^{1/2}[\text{NADH}]^{1/2}}{[\text{l-Balate-}^{2-}]^{1/2}[\text{NAD}^+]}$	$2.86 \times 10^{-12} \text{ M}$	2.86×10^{-5}	-0.184
EC 1.1.1.37			
4 c 3 $K_{GPDH} = \frac{[\alpha\text{-glycerol-P}^{2-}]^{1/2}[\text{NADH}]^{1/2}}{[\text{DHAP-}^{2-}]^{1/2}[\text{NAD}^+]}$	$1.3 \times 10^{-11} \text{ M}$	1.3×10^{-4}	-0.203
EC 1.1.1.94			
4 c 4 $K_{GAldDH} = \frac{[\text{l,3 DiGlc A-}]^{1/2}[\text{NADH}]^{1/2}}{[\text{GAldP-}^{2-}]^{1/2}[\text{Pi-}^{2-}]^{1/2}[\text{NAD}^+]}$	$5.3 \times 10^{-8} \text{ M}$	5.3×10^{-1}	-0.302
EC 1.2.1.12			
$K_{ADH} = \frac{[\text{acetaldehyde}]^{1/2}[\text{NADH}]^{1/2}}{[\text{ethanol}]^{1/2}[\text{NAD}^+]}$	$1.94 \times 10^{-11} \text{ M}$	1.94×10^{-4}	-0.209
EC 1.1.1.1			
$K_{1DPDH} = \frac{[\text{D-fructose}]^{1/2}[\text{NADH}]^{1/2}}{[\text{D-sorbitol}]^{1/2}[\text{NAD}^+]}$	$1.14 \times 10^{-9} \text{ M}$	1.14×10^{-2}	-0.252
EC 1.1.1.14			

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Mitochondrial NAD - Linked Dehydrogenases

$$4 \text{ a } 1 \quad K_{NADH} = \frac{[\text{acetooacetate}^{\cdot}][\text{NADH}]^{+}}{[\text{d}-\beta\text{-hydroxybutyrate}^{\cdot}][\text{NAD}^{+}]}$$

EC 1.1.1.30

$$4 \text{ a } 2 \quad K_{GDH} = \frac{[\text{L}-\text{glutamate}^{\cdot}][\text{NADH}]^{+}}{[\text{L}-\text{glutamate}][\text{NAD}^{+}]}$$

EC 1.4.1.3

$$K_{AIDH} = \frac{[\text{acetate}^{\cdot}][\text{NADH}]^{+}^2}{[\text{acetaldheyde}][\text{NAD}^{+}]}$$

EC 1.2.1.3

Cyttoplasmic NADP - Linked Dehydrogenases

$$4 \text{ b } 1 \quad K_{ICDH} = \frac{[\alpha - \text{KG}^{2-}][\text{ICD}][\text{NADPH}]}{[\text{L}-\text{isocitrate}^{\cdot}][\text{NADP}^{+}]}$$

EC 1.1.1.42

$$4 \text{ b } 2 \quad K_{Malic Enz} = \frac{[\text{pyruvate}^{\cdot}][\text{ICD}][\text{NADPH}]}{[\text{malate}^{\cdot}][\text{NADP}^{+}]}$$

EC 1.1.1.40

$$4 \text{ b } 3 \quad K_{APGDH} = \frac{[\text{ribulose 5-p}^{2-}][\text{ICD}][\text{NADPH}]}{[\text{6-phosphogluconate}^{\cdot}][\text{NADP}^{+}]}$$

EC 1.1.1.43

* See ref.

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Liking Isoenzymes

$$4 L 1 \quad K_{BD} = \frac{[\alpha'-K6^{2-}][\text{-aspartate}^-]}{[1-\text{glutamate}][\text{oxaloacetate}^-]} \quad 6.61$$

EE 2.6.1.1

$$4 L 2 \quad K_{BPI} = \frac{[\alpha'-K6^{2-}][\text{-alanine}]}{[1-\text{glutamate}][\text{pyruvate}^-]} \quad 1.52$$

EE 2.6.1.2

$$4 L 3 \quad K_{PPI} = \frac{[\text{dihydroxyacetone-P}^{2-}]}{[\text{glyceraldehyde 3-P}^{2-}]} \quad 22$$

EE 5.3.1.1

References for Values of Near-Equilibrium Reactions in Equation 4

Equation Abbreviation Reference

4 C 1	K_{LDH}	Williamson DH, Lund P, Krebs HA. <i>Biochem J</i> 195: 514-527, 1967
4 C 2	K_{MDH}	Guyon R, Selberg H, Veech RL. <i>J Biol Chem</i> 248: 6957-6965, 1973
4 C 3	K_{GPDH}	Ruscan W. <i>Thesis</i> , Munich, 1969.
4 C 4	K_{GAFDH}	Cornell W, Leadbetter M, Veech RL. <i>J Biol Chem</i> 254: 6522-6527, 1979
4 M 1	K_{NBBH}	Williamson DH, Lund P, Krebs HA. <i>Biochem J</i> 105: 514-527, 1967
4 M 2	K_{GLDH}	Engel P, Dalziel K. <i>Biochem J</i> 105: 691-695, 1967

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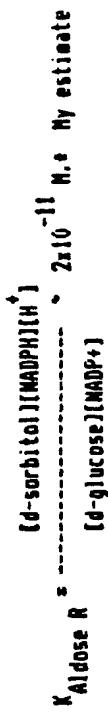
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4 L 1	K_{ICDH}	Londesborough J, Dalziel K. <i>Biochem J</i> 110: 217-222, 1968
4 L 2	$K_{H.E.}$	Veeth R, Eggleston LV, Krebs HA. <i>Biochem J</i> 115: 609-619, 1967
4 L 3	K_{6PGDH}	Villet R, Dalziel K. <i>Biochem J</i> 115: 633-638, 1969
4 L 4	K_{GOT}	Krebs HA. <i>Adv Enz Reg</i> 13: 449-472, 1975
4 L 5	K_{GPT}	Krebs HA. <i>Adv Enz Reg</i> 13: 449-472, 1975
4 L 6	K_{TPI}	Veeth RL, Rajean L, Dalziel K, Krebs HA. <i>Biochem J</i> 115: 837-842, 1969

- The enzyme aldose reductase EC 1.1.1.21 may be redox active during fructose infusion in certain tissues.
The reaction is:



For description, see Hayanai S, Kinoshita JH. *J Biol Chem* 240: 877, 1965

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V Eqn 5 Phosphorylation State Equations - $\frac{[\text{ATP}]}{[\text{ADP}][\text{Pi}]}$

Veeth RL, Larson JK, Cornell NW, Krebs HE, J Biol Chem 254: 6538-6547, 1979

5a. The equilibrium constant of the glyceraldehyde 3-phosphate dehydrogenase (EC 1.1.1.291) and 3 phosphoglycerate kinase reactions (EC 2.7.2.3) at 38°C, $K_{GAP} = 0.25$, and free $[\text{Mg}^{2+}] = 1 \text{ mM}$ is:

$$K_{GAP} = \frac{[\text{ATP}]}{[\text{ADP}][\text{Pi}]} \cdot \frac{[\text{NADH}]^+}{[\text{NAD}]} = 1.03 \times 10^{-4}$$

5b. Combining the above reaction with K_{LDH} and substituting $[\text{DHAP}] = [\text{GAP}]/22$

$$\frac{K_{GAP}}{K_{LDH}} = \frac{[\text{ATP}]}{[\text{ADP}][\text{Pi}]} \cdot \frac{[\text{l-lactate}]}{[\text{pyruvate}]} = 1.65 \times 10^{47} \text{ M}^{-1}$$

5c. Or:

$$\frac{\text{free Cytoplasmic}}{[\text{ADP}][\text{Pi}]} = \frac{[\text{ATP}]}{[\text{ADP}][\text{Pi}]} \cdot \frac{[\text{DHAP}]}{[\text{GAP}]} \cdot \frac{[\text{pyruvate}]}{[\text{l-lactate}]} = 7.5 \times 10^{45} \text{ M}^{-1}$$

5d. Alternatively, from the creatine phosphokinase reaction (EC 2.7.3.2)

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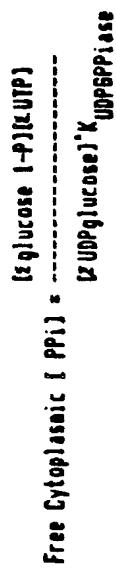
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$$K_{CK} = \frac{[\text{ATP}]}{[\text{ADP}]} \cdot \frac{[\text{Creatine}]}{[\text{Creatine-P}][\text{H}^+]} = 1.66 \times 10^{-9} \text{ M}^{-1}$$

For the Pyrophosphorylation State or [PPi]/[Pi]:

Lawson JWR, Guynn RW, Cornell MM, Veech RL. In *Glucoseogenesis* (Hanson RN, Mehlhaas MA eds) pp 461-511, John Wiley, New York, 1976

5e. From the UDPG Pyrophosphorylase reaction (EC 2.7.7.9):



$$\text{where } K_{\text{UDPGPPiase}} = 4.55$$

5f. For liver and blood glucose:



$$K_{\text{G-6-P-PPi Trans Pass}} = \frac{([\text{free F}_1,6\text{-diPi}][\text{Pi}])}{([\text{fructose-6-P}][\text{Pi}])} = 29.0$$

5g.

VI Eqn 6 Determination of Osmotic Pressure - π

[0211] Van't Hoff JH. *Arch Neerl Sci* 20: 239-303, 1886

where:

- π ~ osmotic pressure in atmospheres (relative to pure H₂O)
 $\sum[C]$ ~ [concentrations] of solutes in mole/liter
R ~ gas constant = 0.082 liter atmospheres/mole/ degree K

T~ 273 + °C

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VII Eqn 7 The Equation of State of the Cell

Relating the E across the cell membrane, the distribution of $[Na^+]$, $[K^+]$, $[Cl^-]$, and $[Ca^{2+}]$ between extracellular fluid and cytoplasmic H_2O and hence cell volume to the cytoplasmic $[ATP]/[ADP] \cdot p_i$

$$\Delta G_{Na/K} \text{ ATPase} = \Delta G_{ATPase} + \Delta G_{ions} + RT \ln \frac{[ATP] \cdot p_i}{[ATP]} + RT \ln \frac{[Na^+]_i^3 [K^+]_i^2 [Cl^-]_0}{[Na^+]_0^3 [K^+]_0^2 [Cl^-]_i} + TAS$$

Since $\Delta G = 0$, then:

$$0 = -7.73 \text{ kcal/mole} + 0 + (-6.3 \text{ kcal/mole}) + 8.5 \text{ kcal/mole} + 5.5 \text{ kcal/mole}$$

$$\text{As } 1 \text{ kcal/mole} = \frac{0.082 \text{ liter atmos/mole/K}}{1.98 \times 10^{-3} \text{ kcal/mole/K}} \times \frac{1}{22.4 \text{ l/mole}} = 1.05 \text{ atmospheres}$$

then the $T S$ term = $5.5 \times 1.05 = 10.2$ atmospheres.

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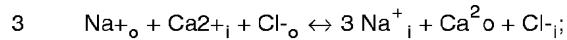
And further from Van't Hoff (Eqn 6)

$$\frac{z[C]_n - z[C]_{out}}{RT} = \frac{\Delta E}{2}$$

$$2[C]_n - 2[C]_{out} = 0.40 \text{ moles/l}$$

Eqn 7 states that since H_2O outside = H_2O inside, the cell is prevented from swelling by the Na^+/K^+ ATPase which electroreutrally pumps out 2 osmoles/ ATP hydrolysed. The ΔE across the cell membrane is reflected by the distribution of $[\text{Cl}^-]_0 / [\text{Cl}^-]_i$ in accordance with the Nernst equation (Eqn 3). The ΔS or decreased entropy within the living cell represents the increase "order" characteristic of the living cell. See Eqn 6.

7b. From the high capacity $\text{Na}^+/\text{Ca}^{2+}$ exchanger written in an electroneutral manner reflecting the free permeability of Cl^- in accordance with the dictates of the Nernst equation, (Eqn 3):



The net osmolar movement of eqn 7a is 2 osmoles \rightarrow outside. In contrast, the net movement of eqn 7b is 3 osmoles \rightarrow inside, requiring the Na^+/K^+ ATPase to cycle 3 times for each 2 times the $\text{Na}^+/\text{Ca}^{2+}$ exchange mechanism operates

in order to maintain osmotic equilibrium.

[0212] The gradient $[Ca^{2+}]_i/[Ca^{2+}]_o$ is thus a direct function of the $[Na^+]_o^3/[Na^+]_i^3$, (the $[Cl^-]_o/[Cl^-]_i$), and a function of the phosphorylation and entropy state of the cell.

[0213] It will be clear to those skilled in the art that equation 7 is the statement of the reaction which links the external environment of the cell to its internal environment and metabolic machinery. Extracellular fluid is thus a creation of the metabolic process of the cell. Changing the external $[Na^+]$, $[K^+]$, $[Cl^-]$, or $[Ca^{2+}]$, or the $[H_2O]$ must necessarily effect the same parameters inside the cell.

[0214] Additionally, the redox and phosphorylation states, the ΔE , and the $T\Delta S$ of the cell are all related and therefore manipulable by the relationships given.

[0215] To control these parameters one needs to use solutions as provided herein which include defined concentrations of Na^+ , K^+ , Cl^- and Ca^{2+} and the related ions HCO_3^- , H^+ , at a defined Mg^{2+} concentration and with a defined osmotic pressure.

[0216] Thus, the present invention provides a process for regulating:

- 15 1) Distribution of water between intracellular and extracellular fluid.
- 2) Distribution of the inorganic electrolytes Na, K, Cl and Ca between intracellular and extracellular fluid.
- 3) and transmembrane cellular potential

[0217] This process is practiced by contacting cells with aqueous near-equilibrium couples as taught by this inventor or by varying the external concentration of Na^+ , K^+ , Cl^- or Ca^{2+} . For example a solution with low $Na:Cl$ ratio raises the phosphorylation potential (See Table III above). In other circumstances, raising $Na:Cl$ outside may raise cellular $[Ca^{2+}]$ for example in rat liver.

25 Claims

1. A physiologically compatible aqueous salt solution for mammalian administration which (a) maintains a normal plasma millequivalent ratio of sodium cations to chloride anions in a normal range, and (b) maintains normal plasma and cellular pH and maintains normal cellular co-factor ratios, said solution containing no plasma proteins and comprising water which has dissolved therein:

(A) the following near equilibrium coupled in the quantity indicated: from 0.1 to 465 millimoles per liter of a couple mixture consisting of d-betahydroxybutyrate anions and acetoacetate anions wherein the milliequivalent ratio of said d-betahydroxybutyrate to said acetoacetate ranges from 6:1 to 0.5:1.

(B) from 1 to 2400 millimoles per liter of sodium cations.

(C) sufficient millimoles per liter of chloride anions to produce a milliequivalent ratio of sodium cations to chloride anions in the range from 1.24 to 1.6.

(D) optionally from 0 to 2400 millimoles per liter of at least one osmotically active substance,

(E) optionally at least one of the following additional cations in a respective quantity as indicated:

cation	quantity (in millimoles/liter)
potassium ⁺	up to 90
calcium ⁺⁺	up to 60
magnesium ⁺⁺	up to 15

(F) optionally up to 25 millimoles per liter of sigma inorganic phosphate.

(G) optionally up to 2 millimoles per liter of sigma inorganic sulfate.

50 the relationship between said water and all solutes in said water being such that said solution has:

(1) an osmolarity ranging from 260 to 5000 milliosmoles:

(2) a pH in the range from 5 to 9; and

(3) the charges of all cations equal the charges of all anions.

55 2. A solution according to claim 1, additionally containing sigma inorganic phosphate

(a) sigma inorganic phosphate in an amount up to 25 millimoles per liter, and/or

(b) sigma inorganic sulfate in an amount up to 2 millimoles per liter.

3. A solution according to claim 1 or 2 further containing an organic non-ionic nutrient in a concentration of up to 2400 millimoles per liter and from 0.1 to 55 millimoles per liter of a further couple mixture consisting of bicarbonate anions and dissolved carbon dioxide wherein the millequivalent ratio of said bicarbonate anions to said carbon dioxide ranges from 0.1:1 to 55:0.1, whereby both normalization of cellular co-factor ratios and normalization of both plasma and intracellular fluid pH is achieved.
4. A solution according to claim 3, wherein the amount of said nonionic substance ranges up to 15 millimoles per liter of at least one dissolved metabolizable nonionic osmotically active substance, said solution being such as to produce a milliosmolarity therein in the range from 280 to 320.
5. A solution according to claim 4, wherein said nonionic substance comprises glucose.
- 15 6. A solution according to claim 4, wherein said nonionic substance is selected from glucose, fructose, glycerol and sorbitol.
7. A solution according to any of claims 3 to 6, wherein said mole ratio of said bicarbonate anions to said carbon dioxide ranges from 0.1:1 to 55:0.1.
- 20 8. A solution according to any preceding claim, wherein the only cation present is sodium.
9. A solution according to any of claims 1 to 7, containing not more than two cations one of which is said sodium while the other thereof is selected from potassium, magnesium and calcium.
- 25 10. A solution according to any of claims 1 to 7, containing three cations, one of which is sodium while the others thereof are selected from the group consisting of potassium, magnesium and calcium.
11. A solution according to claim 10, wherein said three cations are sodium, potassium and calcium.

Patentansprüche

1. Physiologisch verträgliche wässrige Salzlösung zur Anwendung bei Säugern, die (a) ein normales plasmatisches milliäquivalentes Verhältnis von Natriumkationen zu Chloridanionen im normalen Bereich und (b) einen normalen plasmatischen und zellulären pH und ein normales zelluläres Co-Faktorverhältnis aufrechterhält, wobei die Lösung keine Plasmaproteine enthält und Wasser umfasst, in dem gelöst ist
 - (A) das folgende nahezu im Gleichgewicht stehende Paar in der angegebenen Menge:
 - 40 von 0,1 bis 465 mMol pro 1 eines Gemischpaars, bestehend aus d-β-Hydroxybutyratanionen und Acetacetatanionen, wobei das milliäquivalente Verhältnis des d-β-Hydroxybutyrats zum Acetacetat in einem Bereich von 6:1 bis 0,5:1 liegt,
 - (B) 1 bis 2400 mMol pro 1 Natriumkationen,
 - (C) ausreichend mMol pro 1 Chloridanionen, um ein milliäquivalentes Verhältnis von Natriumkationen zu Chloridanionen in einem Bereich von 1,24 bis 1,6 herzustellen,
 - (D) gegebenenfalls 0 bis 2400 mMol pro 1 wenigstens einer osmotisch aktiven Substanz,
 - (E) gegebenenfalls wenigstens eines der folgenden zusätzlichen Kationen in den entsprechend angegebenen Mengen:

Kation	Menge (in mMol/l)
Kalium ⁺	bis zu 90
Calcium ⁺⁺	bis zu 60
Magnesium ⁺⁺	bis zu 15

- (F) gegebenenfalls bis zu 25 mMol pro 1 sigma-anorganischen Phosphats,
(G) gegebenenfalls bis zum 2 mMol pro 1 sigma-anorganischen Sulfats,

5 wobei das Verhältnis zwischen Wasser und allen im Wasser gelösten Bestandteilen derartig ist, dass die Lösung aufweist:

- (1) eine Osmolarität in einem Bereich von 260 bis 5000 Milliosmol;
(2) einen pH im Bereich von 5 bis 9; und
(3) Gleichgewicht der Ladung aller Kationen zur Ladung aller Anionen.

10 2. Eine Lösung nach Anspruch 1, zusätzlich enthaltend sigma-anorganisches Phosphat

- (a) sigma-anorganisches Phosphat in einer Menge bis zu 25 mMol pro 1, und/oder
(b) sigma-anorganisches Sulfat in einer Menge bis zu 2 mMol pro 1.

15 3. Lösung nach Anspruch 1 oder 2, wobei sie ferner einen organischen nicht-ionischen Nährstoff in einer Konzentration bis zu 2400 mMol pro 1 und 0,1 bis 55 mMol pro 1 eines weiteren Gemischpaars, bestehend aus Bicarbonatanionen und gelöstem Kohlendioxid, wobei das milliäquivalente Verhältnis der Bicarbonatanionen zum Kohlendioxid in einem Bereich von 0,1:1 bis 55:0,1 liegt, wobei Normalisierung der zellulären Co-Faktorverhältnisse und Normalisierung des pH von Plasma und intrazellulärer Flüssigkeit erreicht wird, enthält.

20 4. Lösung nach Anspruch 3, wobei die Menge der nicht-ionischen Substanz in einem Bereich bis zu 15 mMol/l wenigstens einer gelösten metabolisierbaren nicht-ionischen osmotisch aktiven Substanz liegt und wobei die Lösung in der Lage ist, eine Milliosmolarität im Bereich von 280 bis 320 herzustellen.

25 5. Lösung nach Anspruch 4, wobei die nicht-ionische Substanz Glucose umfasst.

30 6. Eine Lösung nach Anspruch 4, wobei die nicht-ionische Substanz ausgewählt wird aus Glucose, Fructose, Glycerin und Sorbit.

35 7. Lösung nach einem der Ansprüche 3 bis 6, wobei das Mol-Verhältnis des Bicarbonatanions zu Kohlendioxid in einem Bereich von 0,1:1 bis 55:0,1 liegt.

8. Lösung nach einem der vorhergehenden Ansprüche, wobei das einzige vorhandene Kation Natrium ist.

35 9. Lösung nach einem der Ansprüche 1 bis 7, enthaltend nicht mehr als zwei Kationen, wobei eines der Kationen Natrium ist und das andere ausgewählt wird aus Kalium, Magnesium und Calcium.

40 10. Lösung nach einem der Ansprüche 1 bis 7, enthaltend drei Kationen, wobei eines Natrium ist und die anderen ausgewählt werden aus der Gruppe, bestehend aus Kalium, Magnesium und Calcium.

11. Lösung nach Anspruch 10, wobei die drei Kationen Natrium, Kalium und Calcium sind.

45 **Revendications**

1. Solution saline aqueuse, compatible sur le plan physiologique, pour une administration à un mammifère qui (a) conserve un rapport plasmatique normal en milliéquivalents des cations de sodium aux anions de chlorure dans une gamme normale, et (b) conserve un pH plasmatique et cellulaire normal et conserve des rapports de co-facteurs cellulaires normaux, ladite solution ne contenant aucune protéine de plasma et comprenant de l'eau dans laquelle est dissous:

- (A) le couple quasi-équilibré suivant dans la quantité indiquée:

- 55 - de 0,1 à 465 millimoles par litre d'un mélange de couple composé d'anions d-bétahydroxybutyrate et d'anions acétoacétate dans lequel le rapport, en milliéquivalents, dudit d-bétahydroxybutyrate audit acétoacétate est dans la gamme de 6:1 à 0,5:1 ;

- (B) de 1 à 2 400 millimoles par litre de cations de sodium ;
 (C) une quantité suffisante de millimoles par litre d'anions chlorure pour produire un rapport en milliéquivalents des cations de sodium aux anions de chlorure dans la gamme de 1,24 à 1,6 ;
 (D) éventuellement de 0 à 2 400 millimoles par litre d'au moins une substance osmotiquement active ;
 (E) éventuellement au moins un des cations supplémentaires suivants en une quantité respective telle qu'indiquée :

Cation	Quantité (en millimoles/litre)
potassium*	jusqu'à 90
Calcium**	jusqu'à 60
Magnésium**	jusqu'à 15

- (F) éventuellement jusqu'à 25 millimoles par litre de phosphate inorganique sigma ;
 (G) éventuellement jusqu'à 2 millimoles par litre de sulfate inorganique sigma ;

la relation entre ladite eau et tous les solutés dans ladite eau étant telle que ladite solution possède :

- (1) une osmolarité dans la gamme de 260 à 5 000 millimoles ;
 (2) un pH compris entre 5 et 9 ; et
 (3) les charges de tous les cations égalent les charges de tous les anions.

2. Solution selon la revendication 1, contenant de plus un phosphate inorganique sigma :
- 25 (a) un phosphate inorganique sigma en une quantité allant jusqu'à 25 millimoles par litre ; et/ou
 (b) un sulfate inorganique sigma en une quantité allant jusqu'à 2 millimoles par litre.
3. Solution selon la revendication 1 ou 2, contenant de plus un nutriment non ionique organique en une concentration allant jusqu'à 2 400 millimoles par litre et de 0,1 à 55 millimoles par litre d'un mélange de couple supplémentaire composé d'anions bicarbonate et de dioxyde de carbone dissous dans lequel le rapport en milliéquivalents desdits anions bicarbonate audit dioxyde de carbone est dans la gamme de 0,1:1 à 55:0,1, de cette manière la normalisation des rapports des co-facteurs cellulaires et la normalisation du pH à la fois du plasma et du fluide intracellulaire sont toutes deux réalisées.
- 35 4. Solution selon la revendication 3, dans laquelle la quantité de ladite substance non ionique va jusqu'à 15 millimoles par litre d'au moins une substance osmotiquement active, non ionique, métabolisable, dissoute, ladite solution étant telle de façon à produire une milliosmolarité dans la gamme de 280 à 320.
- 40 5. Solution selon la revendication 4, dans laquelle ladite substance non ionique comprend du glucose.
6. Solution selon la revendication 4, dans laquelle ladite substance non ionique est choisie parmi le glucose, le fructose, le glycérol et le sorbitol.
- 45 7. Solution selon l'une quelconque des revendications 3 à 6, dans laquelle ledit rapport molaire desdits anions bi-carbonate audit dioxyde de carbone est dans la gamme de 0,1:1 à 55:0,1.
8. Solution selon l'une quelconque des revendications précédentes, dans laquelle le seul cation présent est le sodium.
- 50 9. Solution selon l'une quelconque des revendications 1 à 7, ne contenant pas plus de deux cations, dont l'un est ledit sodium alors que l'autre est choisi parmi le potassium, le magnésium et le calcium.
10. Solution selon l'une quelconque des revendications 1 à 7, contenant trois cations, dont l'un est le sodium alors que les autres sont choisis dans le groupe constitué du potassium, du magnésium et du calcium.
- 55 11. Solution selon la revendication 10, dans laquelle lesdits trois cations sont le sodium, le potassium et le calcium.